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Sulfonamide and carbamate derivatives of 6-chloropurine: synthesis, characterization and antimicrobial activity evaluation

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Abstract: A series of new sulfonamide derivatives, 9-(substitutedbenzenesulfonyl)-6-chloro-9*H*-purines 7(a-e) and carbamate derivatives, 6-chloro-purine-9-carboxylic acid substituted alkyl/arylester 9(a-d), have been synthesized through an intermediate, sodium salt of 6-chloro-9(*H*)-purine (6) which was prepared by the treatment of 6-chloro-9(*H*)-purine (4) with sodium hydride. Structures of the newly synthesized compounds were elucidated by IR, NMR (¹H and¹³C), mass spectra and elemental analysis. Antimicrobial activity against three bacterial strains and three fungal strains at two different concentrations, 100 and 200 μ g/mL including MIC values was investigated. Bio-screening data disclosed that most of the sulfonamide derivatives, 7a, 7c and 7d, and one carbamate derivative 9a showed promising antimicrobial activity having MIC values in the range of 18.0-25.0 μ g/mL.

Keywords: 6-Chloro-9(H)-purine; sulfonamides; carbamates; antimicrobial activity; minimum inhibitory concentration. © 2014 ACG Publications. All rights reserved.

1. Introduction

It is well known that nitrogen heterocyclic compounds, particularly, purine derivatives are important targeting scaffolds for many biosynthetic, regulatory and signal transduction proteins including cellular kinases, G proteins and polymerases.¹⁻⁶ Also, they are basic structures in nucleic acids like adenine and guanine which are essential for genetic and many metabolic processes in living organisms. Purine derivatives have been playing indispensible role in different phases of the cell cycles, in cell signaling and other fundamental reactions in the biological system due to a great number of enzymes and receptors are associated with them.^{7,8} Some purine derivatives like N^6 -[(3-methylbut-2-en-1-yl)amino]-purine (1) showed cytokinin activity,⁹ isoprenoid cytokinins are able to influence plant growth development and cell differentiation,¹⁰ and agelasimines¹¹ (2 and 3) (Figure 1)

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exhibited promising biological activities such as antimicrobial and cytotoxic effects. Purine nucleosides and their substituted analogues also exhibited numerous biological activities such as enzyme inhibitors,¹² cytotoxic,¹³ antihypergylcemic,¹⁴ antiviral,¹⁵ immunostimulant¹⁶ antifungal and antibacterial agents.¹⁷



Figure 1. Some biologically active purine derivatives

Active research has been going on sulfonamide derivatives from the last five decades due to their unique importance in both chemical and biological aspects. Generally, the preparation of sulfonamide and carbamate derivatives of 6-chloropurine requires as harsh conditions, however, 9-arylsulfonyl derivatives of 6-chloropurine were synthesized using KOH (aq) as a base in acetone at 0 $^{\circ}$ C.¹⁸ They exhibited high antimycobacterial activity (MIC 0.39 µg/mL against M. tuberculosis). Primarily, sulfonamides (-SO₂-N-) are bacteriostatic agents or an acetylated metabolite finds use in both human therapy and animal husbandry.¹⁹ The sulfonamide derivatives have been found to be used in various biological application such as antitumor,²⁰ hypoglycemic,²¹ anti-thyroid,²² anti-carbonic anhydrase,²³ anti-inflammatory,²⁴ diuretic,²⁵ COX-inhibitors, the enzyme dihydropteroatesynthetase (DHPS)-the key enzyme involved in folate synthesis, anti-impotent drugs.²⁶ Organic carbamates are valuable synthetic intermediates and found in a variety of biologically active compounds.²⁷⁻²⁸ For example, *O*-alkyl and *O*-aryl carbamate derivatives of the antimalarial drug, primaquine is a potential pro-drug that prevent oxidative deamination to the inactive metabolite carboxyprimaquine.²⁹ Recently, our group have been synthesized sulfonamide and carbamate derivatives of (2'-(1*H*-tetrazol-5-yl)-biphenyl-4-yl)methanamine and carvedilol and evaluated their antimicrobial activity, a few of them exhibited potent antimicrobial activity.^{30,31}

By considering an overview facts and our continuing research in the synthesis of biologically active sulfonamide/carbamate derivatives, herein, we report the synthesis of a series of new sulfonamide and carabamate derivatives of 6-chloropurine for biological interest and their antimicrobial activity was screened.

2. Results and Discussion

Recently, our group synthesized phosphoramidate derivatives of 6-chloropurine showed potent antimicrobial activity³² and these consequences encouraged us to design and synthesize a new series of 6-chloropurine derivatives. As in the continuation of our medicinal chemistry programme and

considering the biological potency of sulfonamide and carbamate derivatives, we synthesized a series of new sulfonamide and carbamate derivatives of 6-chloropurine and the schematic representation is depicted in **Figure 2**.



Figure 2. Synthesis of sulfonamide and carbamate derivatives of 6-chloropurine.

The title compounds were synthesized by *in situ* in two steps. Initially, 6-chloropurine (4) was treated with a base, sodium hydride at 5-20 °C in THF to get an intermediate, sodium salt of 6-chloropurine (5). Subsequently, the intermediate 5 was directly reacted with substituted phenyl sulfonyl chlorides 6(a-e) and substituted alkyl/phenyl chloroformates 8(a-d) at 10-40 °C to obtain the title sulfonamides 7(a-e) and carbamates 9(a-d) of 6-chloropurine, respectively. After completion of the reaction, NaCl salt was filtered and the filtrate was concentrated under vacuum to get the crude products. Column chromatography was adopted to purify the crude compounds using 15-25% of methanol and chloroform as mobile phase (Table 1).

Structures of the newly synthesized pure compounds were elucidated by spectroscopic (IR, ¹H-, ¹³C NMR and mass) data and elemental analysis. In IR spectra, absorption bands in the range of 1360-1385 cm⁻¹ (str) and 1180-1195 cm⁻¹ (ben) in the sulfonamide derivatives **7(a-e)**, and 1750-1770 cm⁻¹ (str) in carbamate derivatives **9(a-d)** were confirmed the presence of $-SO_2$ and -CO functionalities, respectively. In ¹H NMR, two protons as singlet in the region of 8.65-8.75 ppm and 8.85-8.97 ppm were assigned to 6-chloropurine moiety and other protons of the corresponding structures were observed in their expected region. The carbon chemical shift values in the range of 158-163 ppm in the carbamate derivatives **9(a-d)** are confirmed -C(O)-. The aromatic and aliphatic carbon signals are recognized in their corresponding region. In addition, the molecular ion peaks of the corresponding mass of the compounds in their mass spectra and the relative CHN composition of the title compounds in the elemental analysis were given further evidence to elucidate the structures of the newly synthesized compounds.

The antibacterial and antifungal activities of the synthesized sulfonamide derivatives 7(a-e) and carbamates 9(a-d) of 6-chloropurine were investigated using disc diffusion^{33,34} and agar discdiffusion³⁵⁻³⁷ methods, respectively. The bacterial strains like *Streptococcus aureus* (ATCC-25923),

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Pseudomonas aeruginosa (ATCC-25619) and *Escherichia coli* (ATCC-9637), and fungi such as *Aspergillus flavus* (MTCC-1884), *Aspergillus niger* (MTCC- 1881) and *Candida albicans* (ATCC-2091) were chosen to screen the antimicrobial activity at two different concentrations, 100 and 200 μ g/mL. Ciprofloxacin and Fluconazole were used as the standards in the antibacterial activity and antifungal activity, respectively and the samples of the title compounds were prepared in DMSO. The microbial growth of zone of inhibition data are tabulated in **Table 2** (antibacterial activity) and **Table 3** (antifungal activity).

Compd.	Time (h)	Yield (%)	M.p. ^O C	Compd.	Time (h)	Yield (%)	M.p. ^o C
7a	4.5	80	157-159	9a	5.5	70	88-90
7b	5.5	72	178-181	9b	6.0	65	74-76
7c	4.0	82	>300	9c	5.0	68	172-175
7d	5.5	79	210-213	9d	5.0	79	189-193
7e	6.0	70	206-208				

Table 1. Physical data of the synthesized sulfonamides 7(a-e) and carbamates 9(a-d).

Table 2. Bacterial	growth of inhibition	of the synthesized c	compounds 7(a-e	e) and 9(a-e)

	Bacterial zone of inhibition in mm							
Compd.	Streptococcus aureus (ATCC-25923)		Pseudomono (ATCC	as aeruginosa C-25619)	Escherichia coli (ATCC-9637)			
	100 µg/mL	200 µg/mL	100 µg/mL	200 µg/mL	100 µg/mL	200 µg/mL		
7a	13.4	20.1	12.8	18.5	14.2	18.9		
7b	6.5	12.7	7.5	14.3	14.7	20.2		
7c	12.3	19.7	11.4	19.2	13.5	21.8		
7d	13.0	19.0	12.6	19.1	15.7	21.2		
7e	8.8	14.6	6.2	10.3	9.4	12.5		
9a	13.1	20.4	11.9	18.5	14.9	22.4		
9b	4.4	11.3	7.2	13.4	8.0	15.2		
9c	5.9	9.7	3.4	8.6	3.9	11.3		
9d	5.0	13.6	9.1	15.9	7.1	12.7		
Std.	18.4	23.5	16.9	22.8	19.4	25.3		

Std. - Standard: Ciprofloxacin was used as standard.

	Fungal zone of inhibition in mm							
Compd.	Aspergillus flavus (MTCC-1884)		Aspergi (MTCO	llus niger C- 1881)	Candida albicans (ATCC- 2091)			
	100 µg/mL	200 µg/mL	100 µg/mL	200 µg/mL	100 µg/mL	200 µg/mL		
7a	13.5	21.2	12.3	20.9	13.0	19.5		
7b	3.7	9.3	10.5	15.2	7.4	10.9		
7c	14.3	21.8	12.4	19.7	14.0	20.5		
7d	12.6	22.1	10.9	20.2	11.3	19.6		
7e	7.7	11.3	5.2	12.1	10.1	18.4		
9a	9.7	20.5	13.1	19.4	12.8	19.0		
9b	7.2	13.0	10.5	17.5	8.3	10.7		
9c	9.6	13.9	4.8	10.3	9.0	18.5		
9d	9.5	17.3	8.0	12.7	10.2	17.6		
Std.	18.1	24.6	16.3	23.5	16.9	22.8		

Table 3. Fungal growth of inhibition of the synthesized compounds 7(a-e) and 9(a-e).

Std. – Standard – Fluconazole was used as standard.

Compd	Minimum inhibitory concentrations in µg/mL						
	S. aureus	P. aeruginosa	E. coli	A. flavus	A. niger	C. albicans	
7a	17.5	23.5	18.0	22.0	21.5	23.5	
7b	NT	NT	19.5	NT	NT	NT	
7c	24.5	18.0	20.0	19.0	24.5	22.0	
7d	21.0	19.5	23.5	25.0	19.0	24.5	
7e	NT	NT	NT	NT	NT	25.0	
9a	24.0	23.5	25.0	19.5	23.5	26.5	
9b	NT	NT	NT	NT	NT	NT	
9c	NT	NT	NT	NT	NT	23.5	
9d	NT	NT	NT	30.0	NT	27.0	
Std ^a .	7.5	9.0	7.0	NT	NT	NT	
Std ^b .	NT	NT	NT	6.0	9.5	8.5	

Fable 4. Minimum inh	hibitory concentrations	of the active	synthesized compounds.
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NT – Not Tested; Std^a. – Ciprofloxacin; Std^b. – Fluconazole; *S. aureus - Streptococcus aureus* (ATCC-25923); *P. aeruginosa - Pseudomonas aeruginosa* (ATCC-25619); *E. coli - Escherichia coli* (ATCC-9637); *A. flavus - Aspergillus flavus* (MTCC-1884); *A. niger - Aspergillus niger* (MTCC-1881); *C. albicans - Candida albicans* (ATCC-2091).

Minimum inhibitory concentration (MIC) was also examined to the active synthesized compounds using micro broth dilution technique³⁵ and summarized the results in **Table 4**. The biological data disclosed that some of the compounds exhibited potential antimicrobial activity. Whereas, the sulfonamide derivatives, **7a** bonded with 4-chloro-3-nitro phenyl ring, **7c** bearing with 4-nitro phenyl ring and **7d** connected with 4-fluoro phenyl ring, and carbamate derivative, **9a** bonded with trichloro ethyl group were exhibited promising bacterial or fungal growth of inhibition and which are closer to the standards. Also, the derivatives **7b** against *E. coli*, **7e** and **9c** against *C. albicans*, **9b** against *A. niger*, and **9d** against *A. flavus* and *C. albicans* showed promising activity. For these active compounds, the MIC values were determined to know their potency. The data revealed that the active compounds displayed minimum inhibitory concentration values in the range of 18.0-30.0 μ g/mL as compared with the standards (6-10 μ g/mL). In the whole observation, the sulfonamide derivatives of 6-chloropurine exhibited promising activity as compared with carbamate derivatives.

3. Experimental

All chemicals and reagents were purchased from *Sigma-Aldrich* and *Merck* and used without further purification. Silica gel 60-120 mesh was used as solid phase in the column chromatography to purify the compounds. Melting points were determined in an open capillary tube by *GUNA* digital melting point apparatus and are uncorrected. IR spectroscopic data were recorded on *Bruker ALPHA* interferometer FT-IR spectrophotometer. ¹H and ¹³C NMR spectroscopic data were recorded on a *Bruker AV* 400 spectrometer in CDCl₃, DMSO- d_6 solvents, 400 MHz and 100 MHz were used for recording ¹H NMR and ¹³C NMR respectively, and tetramethylsilane (TMS) was used as internal standard. E.S.I mass spectra were recorded on *Agilent* 1000 mass spectrometer. Elemental analysis was performed on *Thermo Finnigan FLASH EA* 1112 instrument. Chemical shifts were recorded in parts per million (ppm) and multiplicities are represented as abbreviations: s (singlet), brs (broad singlet), d (doublet), triplet (t) and m (multiplet).

3.1. General procedure for synthesis of compounds (7a-e) and 9(a-d)

6-Chloropurine (1.5 mmol, 231 mg) (4) was taken into a round-bottomed flask (50 mL) containing 15 mL of THF: pyridine (2:1). The reaction mass was cooled to 5-10 °C using ice bath and then sodium hydride (2.0 mmol, 48 mg) was added. The reaction mixture was stirred for 1.5 h at 10-20 °C to obtain sodium salt of 6-chloropurine (5). The completion of the reaction was confirmed by disappearance of starting material in the TLC. The phenyl sulfonyl chlorides and substituted alkyl/phenyl chloro formates (1.5 mmol) was added to a solution of compound **6(a-e)** (for **7a-e)** and compound **8(a-d)** (1.5 mmol) (for **9a-d)** in dry tetrahydrofuran (5.0 mL). The mixture was stirred at 10 °C. Later slowly increased the reaction temperature to 40 °C and stirred for 4-6 h. After completion of the reaction as checked by TLC using 25% methanol in chloroform, the reaction mass was cooled to 25 °C and filtered to remove the salt, NaCl. The bed was washed with THF (5 mL). Column chromatography was adopted to purify the crude compounds using 15-25% of methanol and chloroform as mobile phase (**Table 1**).

3.1.1. 6-Chloro-9-(4-chloro-3-nitro-benzenesulfonyl)-9H-purine (7a):



Yield: 80% , pale yellow solid, m.p. 157-159 °C; ¹H NMR (DMSO- d_6) (δ /ppm) : 7.85 (d, 1H, J = 8.0 Hz, Ar-H), 8.21(d, 1H, J = 8.0 Hz, Ar-H), 8.73 (s, 1H, Ar-H), 8.83 (s, 1H, -N-CH-N-), 9.21(s, 1H, Cl-C-N-CH-N); ¹³C NMR (DMSO- d_6) (δ /ppm) : 124.5, 129.5, 131.2, 132.9, 134.4, 138.0, 146.1, 147.9, 149.8, 150.6, 154.2; IR(v/cm⁻¹): 1442.5 (NO₂), 1400.3 (SO₂), 1188.9 (SO₂), 690.1 (C-Cl); ESI-Ms m/z (%): 374 (M+H)⁺ (100), 376 (M+H⁺+2) (67); Analysis (% Calculated/found) for C₁₁H₅Cl₂N₅O₄S C:35.31/35.26, H:1.35/1.32, N:18.72/18.67.

3.1.2. 9-(4-Bromo-benzenesulfonyl)-6-chloro-9H-purine (7b):



Yield: 72%, light brown solid, m.p. 178-181 °C, ¹H NMR (CDCl₃) (δ /ppm) : 7.77 (d, 2H, J = 8.0 Hz, Ar-H), 8.18 (d, 2H, J = 8.0 Hz, Ar-H), 8.56 (s, 1H, -N-CH-N-), 8.85 (s, 1H, Cl-C-N-CH-N); ¹³C NMR (CDCl₃) (δ /ppm): 128.2, 130.8, 131.0, 131.2, 133.6, 144.8, 149.5, 150.9, 153.8; IR (v/cm⁻¹): 1367.0 (SO₂), 1189.7 (SO₂, bending), 688.5 (C-Cl, str), 569.2 (C-Br, str); ESI-Ms m/z (%): 373 (M+H) ⁺ (77.9), 375 (M+H⁺+2) (100); Analysis (% Calculated/found) for C₁₁H₆BrClN₄O₂S C:35.36/35.31, H:1.62/1.56, N:15.00/14.94.

3.1.3. 6-Chloro-9-(4-nitro-benzenesulfonyl)-9H-purine (7c):



Yield:82%, yellow solid, m.p. 300 °C, ¹H NMR (CDCl₃) (δ /ppm): 8.47 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.53 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.92 (s, 1H, -N-CH-N-), 9.20 (s, 1H, Cl-C-N-CH-N); ¹³C NMR (CDCl₃) (δ /ppm) : 125.2, 130.9, 131.6, 143.1, 148.5, 149.2, 151.3, 152.6, 153.5; IR (v/cm⁻¹): 1442.5 (NO₂), 1366.4 (SO₂), 1187.7 (SO₂), 629.8 (C-Cl); ESI-Ms *m*/*z* (%): 340 (M+H)⁺ (100), 342 (M+H⁺+2) (33); Analysis (% Calculated/found) for C₁₁H₆ClN₅O₄S C:38.89/38.83, H:1.78/1.76, N:20.62/20.59.

3.1.4. 6-Chloro-9-(4-fluoro-benzenesulfonyl)-9H-purine (7d):



Yield: 79%, pale brown solid, m.p. 210-213 °C, ¹H NMR (CDCl₃) (δ /ppm): 7.33 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.17 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.69 (s, 1H, N-CH-N-), 9.03 (s, 1H, Cl-C-N-CH-N-); ¹³C NMR (CDCl₃) (δ /ppm) :119.5, 131.2, 132.1, 134.9, 148.5, 152.2, 152.9, 154.1,168.3 (d, *J* =184.5 Hz); IR

(v/cm⁻¹): 1366.0 (SO₂),1188.5 (SO₂), 1091.4 (C-F), 626.4 (C-Cl); ESI-Ms m/z (%): 313.2 (M+H)⁺ (32.1), 315 (M+H⁺+2) (33), 288.3 [M+H⁺-25] (30.9), 274.4 [M+H⁺-39] (100); Analysis (% Calculated/found) for C₁₁H₆CIFN₄O₂S C:42.25/42.19, H:1.93/1.92, N:17.92/17.89.

3.1.5. 4-(6-Chloro-purine-9-sulfonyl)-phenylamine (7e):



Yield: 70%, brown solid, m.p. 206-208 °C, ¹H NMR (CDCl₃) (δ /ppm): 5.62 (brs, 2H, Ar-NH₂), 6.93 (d, 2H, *J* = 8.0 Hz, Ar-H), 7.76 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.78 (s, 1H, -N-CH-N-), 8.96 (s, 1H, Cl-C-N-CH-N-); ¹³C NMR (CDCl₃) (δ /ppm): 115.1, 126.8, 130.4, 132.9, 148.5, 152.0, 151.8, 153.7, 155.1 ppm; IR (v/cm⁻¹): 3368.4 (NH₂), 1531.1 (NH₂), 1366.4 (SO₂), 1187.7 (SO₂), 629.8 (C-Cl); ESI-Ms *m/z* (%): 310.3 (M+H)⁺ (31.3), 312.4 (M+H⁺+2) (9.7), 274.4 [M+H⁺-36] (100); Analysis (% Calculated/found) for C₁₁H₈ClN₅O₂S C:42.66/42.61, H:2.60/2.57, N:22.61/22.56.

3.1.6. 6-Chloro-purine-9-carboxylic acid 2, 2, 2-trichloro-ethyl ester (9a):



Yield:70%, off-white solid, m.p. 88-90 °C, ¹H NMR (CDCl₃) (δ /ppm): 4.87 (s, 2H, CH₂, aliphatic), 8.64 (1H, s, -NCHN-), 8.95 (1H, s, Cl-C-N-CH-N-); ¹³C NMR (CDCl₃) (δ /ppm): 76.3, 98.0, 138.2, 143.4, 148.3, 151.8, 152.6, 154.6; IR (ν /cm⁻¹): 1764.4 (C=O), 1367.5 (C-N), 1146.7 (C-O), 685.7 (C-Cl); ESI-Ms *m*/*z* (%) : 330.5 [M+H]⁺ (11.6), 353.5 [M+Na] (100), 302.5 [M+H⁺-28] (23), 288.3 [M+H⁺-41] (35.6), 274.4 [M+H⁺-55] (40); Analysis (% Calculated/found) for C₈H₄Cl₄N₄O₂ C:29.12/29.09, H:1.22/1.20, N:16.98/16.96.

3.1.7. 6-Chloro-purine-9-carboxylic acid isobutyl ester (9b):



Yield:65%, pale brown solid, m.p. 74-76 °C, ¹H NMR (CDCl₃) (δ /ppm): 1.10 (d, 6H, J = 6.7 Hz, (CH₃)₂-CH-CH₂-O), 2.25-2.18 (m, 1H, (CH₃)₂-CH-CH₂-O), 4.38 (d, 2H, J = 8.0 Hz, (CH₃)₂-CH-CH₂-O), 8.65 (1H, s, -N-CH-N-), 8.93 (1H, s, Cl-C-N-CH-N-); ¹³C NMR (CDCl₃) (δ /ppm): 18.9 (CH₃)₂-CH-CH₂-O), 27.8 (CH₃)₂-CH-CH₂-O), 75.3 (CH₃)₂-CH-CH₂-O), 143.7, 146.9, 151.9, 154.2, 159.9; IR (v/cm⁻¹): 1758.7 (C=O), 1288.5 (C-N), 1141.9 (C-O), 634.5 (C-Cl); ESI-Ms m/z (%): 255.2 (M+H)⁺ (100), 256.1 (M+H⁺+2) (33); Analysis (% Calculated/found) for C₁₀H₁₁ClN₄O₂ C:47.16/47.19, H:4.35/4.35, N:22.00/22.03.



Yield: 68%, pale yellow solid; m.p. 172-175 °C, ¹H NMR (CDCl₃) (δ /ppm): 5.53 (s, 2H, -CH₂-C₆H₅), 7.58 (d, 2H, *J* = 8.0 Hz, Ar-H), 7.96-7 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.67 (s, 1H, -N-CH-N-), 9.05 (s, 1H, Cl-C-N-CH-N-); IR (v/cm⁻¹): 1758.1 (C=O), 1448.1 (NO₂), 1140.3 (C-O), 1281.9 (C-N), 681.4 (C-Cl); ESI-Ms *m*/*z* (%): 334.4 (M+H)⁺ (100), 336.2 (M+H⁺+2) (33); Analysis (% Calculated/found) for C₁₃H₈ClN₅O₄ C:46.79/46.74, H:2.42/2.40, N:20.99/21.00.

3.1.9. 6-Chloro-purine-9-carboxylic acid 4-nitro-phenyl ester (9d):



Yield:79%, off-white solid; m.p. 189-193 °C, ¹H NMR (CDCl₃) (δ /ppm) : 7.66 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.21 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.62 (s, 1H, -N-CH-N-), 8.96 (s, 1H, Cl-C-N-CH-N-); ¹³C NMR (CDCl₃) (δ /ppm): 123.0, 125.4, 136.2, 138.9, 143.2, 145.5, 149.1, 152.3, 152.8, 156.4; IR (v/cm⁻¹): 1614.6 (C=O), 1447.1 (NO₂), 1288.5 (C-N), 1166.4 (C-O), 692.5 (C-Cl); ESI-Ms *m/z* (%): 320.4 (M+H)⁺ (100), 322.0 [M+H⁺+2] (32.8); Analysis (% Calculated/found) for C₁₂H₆ClN₅O₄ C:45.09/45.03, H:1.89/1.85, N:21.91/21.89.

3.2. Antimicrobial activity

3.2.1. Antibacterial activity

The newly synthesized sulfonamides **7(a-e)** and carbamates **9(a-e)** of 6-chloro-9*H*-purine were evaluated for their antibacterial activity using disc diffusion method.³⁴ The bacterial strains such as *Streptococcus aureus* (ATCC-25923), *Pseudomonas aeruginosa* (ATCC-25619) and *Escherichia coli* (ATCC-9637) were selected to investigate the activity and Ciprofloxacin was used as a standard drug. The title compounds (1 mg) were dissolved in methanol and adjusted their concentration to 100 and 200 µg/mL to examine the activity. A standard inoculum of 1-2 x 10⁻⁷CFU/mL (0.5 McFarland standards) was introduced onto the surface of sterile agar plates and made even distribution of the inoculums. The dry sterilized discs of 6 mm were soaked in the test solutions (100 and 200 µg/mL) and placed in nutrient agar medium. Blank test showed that, methanol used in the preparation of the test solutions does not affect the bacteria. The inoculated plates were inverted and incubated for 24 h at 37 ± 3 °C. The bacterial growth of inhibition as zone of inhibition around the disc was measured in millimeters. The experiments were repeated three times and average data was taken as final result.

3.2.2. Antifungal activity

The synthesized sulfonamides **7(a-e)** and carbamates **9(a-e)** were evaluated for their antifungal activity against fungal strains such as *Aspergillus flavus* (MTCC-1884), *Aspergillus niger* (MTCC-

1881) and *Candida albicans* (ATCC- 2091) using agar disc-diffusion method.³⁶ Fluconazole was used as a standard drug for antifungal study. The fungal strains were maintained on Potato Dextrose Agar (PDA) medium (Hi-Media). The culture from the slant was inoculated into the Potato Dextrose broth and incubated at 37 °C for 48-72 h. This culture (0.1 mL) was spread on the potato dextrose agar plate. The compounds (1 mg) were dissolved in methanol and adjusted their concentration to 100 and 200 μ g/mL. Sterile discs of 6 mm diameter soaked into the test solutions and these are impregnated on the surface of the media and incubated for 48-72 h at 37±3 °C. The zone of inhibition around the disc was measured in millimeters. The tests were repeated three times and average value was taken.

Minimum inhibitory concentrations (MICs) were examined using micro broth dilution technique.³⁵ In different test tubes 0.1 mL of standardized inoculum (1-2 x 10^7 CFU/mL) was added and, incubated 24 h for bacterial inoculums and 48-72 h for fungal inoculum at 37 ± 3 °C. Two controls were maintained for each test sample. The growth was monitored visually and spectrophotometrically. The lowest concentration (highest dilution) required arresting the growth of bacteria or fungi was regarded as minimum inhibitory concentration (MIC).

4. Conclusion

In the present study as in the part of medicinal chemistry programme, we have been synthesized a series of new sulfonamide derivatives, 9-(substitutedbenzenesulfonyl)-6-chloro-9*H*-purines and carbamate derivatives, 6-chloro-purine-9-carboxylicacid substituted alkyl/arylester by in situ fashion. Antimicrobial activity against three bacterial strains and three fungal strains at two different concentrations, 100 and 200 μ g/mL including MIC values was investigated. Bio-screening data disclosed that all the title compounds exhibited promising antimicrobial activity at both the concentrations. Sulfonamide derivatives **7a**, **7c** and **7d**, and one carbamate derivative **9a** showed promising growth of inhibition of selected bacterial and fungal strains and these compounds showed MIC values in the range of 18.0-25.0 μ g/mL as compared with other title compounds and near activity of the standards. In whole comparison, the sulfonamide derivatives. The study of results encouraged us to design new library of purine derivatives as antimicrobial agents in future endeavors and might be worthy to medicinal chemistry.

References

- [1] Legraverend, M.; Grierson, D.S. The purines: potent and versatile small molecule inhibitors and modulators of key biological targets. *Bioorg. Med. Chem.* **2006**, *14*, 3987-4006.
- [2] Chang, Y.T.; Wignall, S.M.; Rosania, G.R.; Gray, N.S.; Hanson, S.R.; Su, A.I.; Merlie, J.Jr.; Moon, H.S.; Sangankar, S.B.; Perez, O.; Heald, R.; Schultz, P.G.J. Synthesis and biological evaluation of myoseverin derivatives: microtubule assembly inhibitors. *Med. Chem.* 2001, 44, 4497-4500.
- [3] Chiosis, G.; Timaul, M.N.; Lucas, B.; Munster, P.N.; Zheng, F.F.; Sepp-Lorenzino, L.; Rosen, N. A small molecule designed to bind to the adenine nucleotide pocket of Hsp90 causes Her2 degradation and the growth arrest and differentiation of breast cancer cells. *Chem. Biol.* **2001**, *8*, 289-299.
- [4] Gundersen, L.L.; Nissen-Meyer, J.; Spilsberg, B. Synthesis and antimycobacterial activity of 6arylpurines: The requirements for the *N*-9 substituent in active antimycobacterial purines. *J. Med. Chem.* **2002**, *45*, 1383-1386.
- [5] Chapman, E.; Ding, S.; Schultz, P.G.; Wong, C.H. A potent and highly selective sulfotransferase inhibitor. *J. Am. Chem. Soc.* **2002**, *124*, 14524-1425.
- [6] Altmann, E.; Cowan-Jacob, S.W.; Missbach, M. Novel purine nitrile derived inhibitors of the cysteine protease cathepsin K. *J. Med. Chem.* **2004**, *47*, 5833-5836.
- [7] Rosemeyer, H. The chemodiversity of purine as a constituent of natural products. *Chem. Biodivers.* 2004, *1*, 361–401.
- [8] Schmidt, A.P.; Lara, D.R.; Souza, D.O. Proposal of a guanine-based purinergic system in the mammalian central nervous system. *Pharmacol. Ther.* **2007**, *116*, 401–416.
- [9] Nelson, J.; Leonard, J.; Achmatowicz, S.; Loeppky, R.; Carraway, K.; Grimm, W. A.H.; Szwezkowska, A.; Hamzi, H.Q.; Skoog, F. Development of cytokinin activity by rearrangement of 1-substituted adenines

to 6-substituted aminopurines: inactivation by N^6 , 1-cyclization. *Proc. Natl. Acad. Sci.* U.S.A. **1966**, *56*, 709-716.

- [10] Davies, P.J. Plant Hormones, Biosynthesis, Signal Transduction, Action. *Kluwer Academic Publishers*: Dordrecht, **2004**, pp 1–8.
- [11] Fathi-Afshar, R.; Allen, T.M. Biologically active metabolites from agelas mauritiana. *Can. J. Chem.* **1988**, *66*, 45-50.
- [12] Brathe, A.; Andresen, G.; Gundersen, L.L.; Malterud, K.E.; Rise, F. Antioxidant activity of synthetic cytokinin analogues: 6-alkynyl- and 6-alkenylpurines as novel 15-Lipoxygenase inhibitors. *Bioorg. Med. Chem.* **2002**, *10*, 1581-1586.
- [13] Andersen, G.; Gundersen, L. L.; Nissen Meyer, J.; Rise, F.; Spilsberg, B. Cytotoxic and antibacterial activity of 2-oxopurine derivatives. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 567-569.
- [14] Kim, B.Y.; Ahn, J.B.; Lee, H.W.; Kang, S.K.; Lee, J.H.; Shin, J.S.; Ahn, S.K.; Hong, C.I.; Yoon, S.S. Synthesis and biological activity of novel substituted pyridines and purines containing 2,4thiazolidinedione. *Eur. J. Med. Chem.* 2004, *39*, 433-447.
- [15] Sidwell, R.W.; Huffman, J.H.; Khare, G.P.; Allen, L.B.; Witkowski, J.T.; Robins, R.K. Broad-spectrum antiviral activity of virazole: 1-beta-*D*-ribofuranosyl-1,2,4-triazole-3-carboxamide. *Science*, **1972**, *177*, 705-706.
- [16] Jin, G.; Wu, C.C.N.; Tawatao, R.I.; Chan, M.; Carson, D.A.; Cottam, H.B. Synthesis and immunostimulatory activity of 8-substituted amino 9-benzyladenines as potent Toll-like receptor 7 agonists. *Bioorg. Med. Chem. Lett.* **2006**, *16*, 4559-4563.
- [17] Bordon-Pallier, F.; Jullian, N.; Ferrari, P.; Girard, A.M.; Bocquel, M.T.; Biton, J.; Bouquin, N.; Haesslein, J.L. Inhibitors of Civ1 kinase belonging to 6-aminoaromatic-2-cyclohexyldiamino purine series as potent anti-fungal compounds. *Biochim. Biophys. Acta*. 2004, *1697*, 211-223.
- [18] Patel, P.R.; Ramalingan, C.; Park, Y.T. Synthesis and antimicrobial evaluation of guanylsulfonamides. *Bioorg. Med. Chem. Lett.* **2007**, 17, 6610-6614.
- [19] Petri, W.A. Good man& Gilman's The PharmacologicalBasis of Therapeutics, 10th ed.; J.G. Hardman, L.E. Limbird, A. G. Gilman, Eds.; McGraw-Hill: New York, **2001**; pp 1171-1179.
- [20] Berger, K.; Petersen, B.; Pfaue, B.H. Persistence of drugs occurring in liquid manure in the food chain. *Arch. Lebensmittelhyg.* **1986**, *37*, 85-108.
- [21] Thornber, C.W. Isosterism and molecular modification in drug design. Chem. Soc Rev. 1979, 8, 563-580.
- [22] Ogden, R.C.; Flexner, C.W. Protease inhibitors in AIDS therapy; New York, U.S.A: *Marcel Dekker*; 2001.
- [23] Nishimori, I.; Vullo, D.; Innocenti, A.; Scozzafava, A.; Mastrolorenz, A.; Supuran, C.T. Carbonic anhydrase inhibitors: Inhibition of the transmembrane isozyme XIV with sulfonamides. *Bioorg. Med. Chem.Lett.* **2005**, *15*, 3828-3833.
- [24] Li, J.J.; Anderson, D.; Burton, E.G.; Cogburn, J.N.; Collins, J.T.; Garland, D.J.; Gregory, S.A.; Huang, H.C.; Isakson, P.C.; Koboldt, C.M.; Logusch, E.W.; Norton, M.B.; Perkins, W.E.; Reinhard, E.J.; Seibert, K.; Veenhuizem, A.W.; Zang, Y.; Reitz, D.B. 1,2-Diarylcyclopentenes as selective cyclooxygenase-2 inhibitors and orally active anti-inflammatory agents. *J. Med. Chem.*, **1995**, 38, 4570-4578.
- [25] Boyd, A.E. Sulfonylurea receptors, ion channels, and fruit flies. *Diabetes*. 1988, 37, 847-850.
- [26] Claudiu, T.S.; Angela, C.; Andrea, S. Protease inhibitors of the sulfonamide type: anticancer, antiinflammatory, and antiviral agents. *Med. Res. Rev.* **2003**, *23*, 535-538.
- [27] Folkmann, M.; Lund, F.G. Acyloxymethyl carbonochloridates. New intermediates in prodrug synthesis. *Synthesis*. **1990**, 1159-1166.
- [28] Warrass, R.; Weismuller, K.H.; Jung, G. Cyclic oligocarbamates. Tetrahedron Lett. 1998, 39, 2715-2716.
- [29] Graca, M.; do Rosario, V.E.; Jim, I.; Luis, C.; Rui, M. A carbamate-based approach to primaquine prodrugs: Antimalarial activity, chemical stability and enzymatic activation. *Bioorg. Med. Chem.* 2012, 20, 886-892.
- [30] Narasaiah, T.; Subba Rao, D.; Rasheed, S.; Madhava, G.; Srinivasulu, D.; Brahma Naidu, P.; Naga Raju, C. Synthesis of novel carbamate, sulfonamide analogues of (2'-(1*H*-tetrazol-5- yl)-biphenyl-4-yl)methanamine and their antibacterial, antifungal activities. *Der Pharmacia Lettre*, **2012**, *4*, 854-856.
- [31] Lakshmi Reddy, S.V.; Thaslim Basha, SK.; Naresh, K.; Naga Raju, C. Synthesis and biological evaluation of carbamate and sulfonamide derivatives of carvedilol. *Der Chemica Sinica*. **2013**, *4*, 127-132.
- [32] Subramanyam, C.H.; Subba Rao, D.; Naga Raju, C.; Adam, S.; Durga Srinivasa Murthy, S. New *N* linked phosphonamidate derivatives of 6-chloropurine: Synthesis and evaluation of antimicrobial and antioxidant activities. *Phosphorus, Sulfur Silicon Relat. Elem.* **2014**, *189*, 1-14.
- [33] Cruickshank, R.; Duguid, J.P.; Marmion, B.P.; Swain, R.H.A. Medicinal Microbiology, 12th ed.; *Churchill Livingstone*: New York, 1975.
- [34] Collins, A.H. Microbiological Methods, 2nd ed.; *Butterworth*: London, 1976.

- [35] Bonjar Shahidi, G.H. Evaluation of antibacterial properties of Iranian medicinal plants against Micrococcus luteus, Serratiamarcescens, Klebsiella pneumonia and Bordetellabronchoseptica. *Asian J. Sci.* **2004**, *3*, 82-86.
- [36] National committee for clinical laboratory standards, *Methods for dilution, antimicrobial susceptibility tests for bacteria that grow aerobically*, 5thed: Wayne, P.A., **2000**.
- [37] Bauer, A.W.; Kirby, M.M.; Sherris, J.C.; Truck, M. Antibiotic susceptibility testing by a standardized single disc method. *Am. J. Clin. Pathol.***1966**, *45*, 493-496.



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