

Radical Scavenging Effects of Different *Veronica* Species

Ummuhan Şebnem Harput^{1*}, Yasin Genç¹, Newaj Khan² and

İclal Saracoglu¹

¹Department of Pharmacognosy, Faculty of Pharmacy, Hacettepe University, 06100, Ankara, Türkiye

²The School of Pharmacy, University of London, 29-39 Brunswick Square, London, United Kingdom

(Received July 15, 2010; Revised September 3, 2010, Accepted September 10, 2010)

Abstract: It is well known that the excessive production of reactive oxygen species is hazardous for living organisms and damages major cellular constituents such as DNA, lipid and protein. To find new products reducing free radical damage is very important researches in recent pharmaceutical investigations. Considering this information, fourteen *Veronica* species are decided to research in the view point of their antioxidant capacity and the chemical content. Water extracts of the plants were tested for their radical scavenging activity against 2,2-diphenyl-1-picryl hydrazyl (DPPH), superoxide (SO) and nitric oxide (NO) radicals spectroscopically. Dose dependent radical scavenging activity was observed and the results were found to be comparable to that of ascorbic acid, quercetin and BHA which are known antioxidative compounds. In addition, gallic acid equivalent total phenolic contents of the plants were also determined using Folin-Ciocalteu reagent. The most significant scavenging activity was found for *V. chamaedrys* against SO radical (IC₅₀ 113.40 µg/ml) and *V. officinalis* against DPPH and NO radicals (IC₅₀ 40.93 µg/ml, 570.33 µg/ml, respectively).

Keywords: *Veronica*; Plantaginaceae; radical scavenging effect; DPPH; Nitric oxide; Superoxide.

1. Introduction

The genus *Veronica* L. (Plantaginaceae) is a large genus of annual and perennial plants located in Central and Southern Europe and Turkey. It is represented by 79 species, 26 of which are endemic in Turkish Flora [1]. The phytochemistry of the genus has been studied extensively with many species surveyed for their iridoid [2, 3] and flavonoid constituents [4,5]. Earlier investigations on *Veronica* species were resulted in the isolation of mainly iridoid glucosides, especially benzoic and cinnamic acid esters of catalpol, some phenylethanoid and flavonoid glycosides [3, 6-10]. Isolated flavonoids were found flavone glycosides, often with additional hydroxyl substitution at C-6 or C-8 of the A-ring. Acylation of the sugars is another characteristic feature of some of the glycosides [4, 6]. A recent systematic study of *Veronica* based on sequence data from the internal transcribed spacer region of nuclear ribosomal DNA has renewed interest in the taxonomy of this genus [11] and it is transferred from the Schrophulariaceae to Plantaginaceae family [10,12].

* Corresponding author: E-Mail: sharput@hacettepe.edu.tr

In addition to the chemotaxonomic and phytochemical importance of the genus, *Veronica* species were also become prominent from the view point of their traditional usage and biological activities. While *V. beccabunga* and *V. officinalis* were used as diuretic and for wound healing [13], *V. anagallis-aquatica* was used for the treatment of rheumatic pain as a poultice [14] in Anatolia. The usage of *V. hederifolia* in cough and influenza, and *V. polita* as expectorant and antiscorbutic were also found in traditional Turkish medicine [15]. In addition, anticancer usages of different *Veronica* species were determined in different countries [14, 16].

Previously, we have performed a lot of phytochemical and biological studies on the genus *Veronica* [7,8,17,18]. *V. persica*, *V. polita*, *V. hederifolia*, *V. anagallis-aquatica*, *V. cymbalaria* and *V. serpyllifolia* were studied phytochemically. We have isolated different iridoid, phenylethanoid and flavonoid glycosides from these species. However we have not studied their biological activity comparatively. Our previous researches and recent phylogenetic and pharmacological studies were turned us to focus on the researches on Plantaginaceae family from the view point of phytochemistry and biological activity. As a part of this ongoing research project, it is planned to test radical scavenging activity of *V. peduncularis*, *V. baranetzki*, *V. officinalis*, *V. orientalis*, *V. persica*, *V. polita*, *V. cuneifolia* subsp. *cuneifolia*, *V. cymbalaria*, *V. hederifolia*, *V. anagallis-aquatica*, *V. fuhsii*, *V. chamaedrys*, Unknown *Veronica* and *V. serpyllifolia* against 2,2-diphenyl-1-picryl hydrazyl (DPPH), superoxide (SO) and nitric oxide (NO) radicals spectroscopically. Their total phenolic contents were also determined using Folin-Ciocalteu reagent.

Extensive studies with different model systems and biological materials have clearly shown that reactive free radicals are able to produce chemical modifications and damage to proteins, lipids, carbohydrates and nucleotides. Many diseases and degenerative processes can be associated with the overproduction of reactive oxygen species (ROS) including inflammation, brain ischemia, mutagenesis, dementia and physiological aging. In addition, it is well known the increased amount of free radicals in wounded and inflamed tissues, cancerous cells and organs [19, 20, 21]. For these reasons, several methods have been developed to measure antioxidant activity of the herbal *extracts in vitro* and here we have tested nitric oxide (NO), superoxide (SO) and 2,2-diphenyl-1-picrylhydrazil (DPPH) radical scavenging activity of the aqueous extract of selected species together with their gallic acid equivalent total phenolic content. There are no reported studies about these species relating to these properties to any biological activity.

2. Materials and Methods

2.1. Plant Material

Tested *Veronica* species were collected from different places in Turkey. A voucher specimen has been deposited in the Herbarium of the Faculty of Pharmacy, Hacettepe University, Ankara, Turkey. Their locations and herbarium numbers were given in Table 1.

2.2. General

DPPH, nitro blue tetrazolium (NBT), sodium nitroprusside, Folin–Ciocalteu reagent, gallic acid, ascorbic acid were obtained from Sigma-Aldrich Chem Co (St. Louis, MO). 3-*t*-butyl-4-hydroxyanizole (BHA) was purchased from Nacalai Tesque Co. (Kyoto, Japan). Sulfanilamide and naphthylethylenediamine dihydrochloride were obtained from Merck Co. (Darmstadt, Germany).

Table 1. Information about the plant species

Plant Material	Location	Herbarium No:
<i>V. peduncularis</i> Bieb.	Trabzon-Maçka	HUEF 09008
<i>V. baranetzki</i> Bordz.	Trabzon-Maçka	HUEF 09007
<i>V. orientalis</i> Miller	Kars-Göle	HUEF 90026
<i>V. polita</i> Fries	Ankara	HUEF 99013
<i>V. persica</i> Poirlet	Urfa	HUEF 99015
<i>V. cymbalaria</i> Bodard	Urfa	HUEF 99131
<i>V. hederifolia</i> L.	Urfa	HUEF 99016
<i>V. cuneifolia</i> subsp. <i>cuneifolia</i> D. Don	Antalya-Akseki	HUEF 06006
<i>V. fuhsii</i> Freyn et.Sint.	Bolu-Abant	HUEF 09327
<i>V. chamedrys</i> L.	Bolu-Abant	HUEF 09326
<i>V. serpyllifolia</i> L.	Bolu-Abant	HUEF 09328
Unknown <i>Veronica</i>	Bolu-Abant	-

2.3. Preparation of herbal extracts

The air-dried aerial parts of the plants were extracted with MeOH at 40°C for three times. The combined MeOH extracts were evaporated under vacuum to give crude MeOH extracts. MeOH extract was dissolved in water and partitioned with petroleum ether to remove chlorophylls. Water extracts were tested for the bioactivity and phytochemistry studies.

2.4. DPPH radical scavenging effect

The DPPH radical scavenging effect of water extracts was assessed by the decoloration of methanol solution of 2,2-diphenyl-1-picrylhydrazyl (DPPH) spectroscopically; BHA, quercetin and ascorbic acid were used as standard compounds. MeOH solution (100 µl) of the extract at various concentrations was added to DPPH/MeOH (80 µg/ml) solution. The reaction mixture was shaken vigorously and the absorbance of remaining DPPH was measured at 520 nm after 30 min. The radical scavenging activity was determined by comparing the absorbance with that of blank (100%) containing only DPPH and solvent. All the analyses were done in 3 replicates [22, 7, 9, 23].

2.5. Superoxide radical scavenging activity by alkaline DMSO method

Superoxide radical scavenging activity of the extracts were determined by alkaline DMSO method according to the method of Elizabeth and Rao (1990) with slight modification. In this method, the concentration of superoxide in alkaline DMSO system corresponds to the concentration of oxygen dissolved in DMSO. In tightly stoppered vessels, superoxide radical is stable more than 24 h; however in open vessels rapidly decreases its concentration. Briefly, superoxide radical was generated in non-enzymatic system. To the reaction mixture containing 10 µL of NBT (1 mg/mL solution in DMSO) and 30 µL of the extract or standard compounds were dissolved in DMSO, 100 µL of alkaline DMSO (1 mL DMSO containing, 5 mM NaOH in 0.1 mL water) was added to give a final volume of 140 µL and the absorbance was measured at 560 nm using microplate reader. The decrease in the absorbance at 560 nm with antioxidants indicated the consumption of generated superoxide [24-27].

2.6. NO scavenging activity

In order to determine NO radical scavenging activity of extracts, 60 µl of serial diluted sample were added into a 96-well flat-bottomed plate. Following this, 60 µl of 10 mM sodium nitroprusside, dissolved in phosphate buffered saline (PBS), were added to each well and the plate was incubated under light at room temperature for 150 min. Finally, an equal volume of Griess reagent (1% sulfanilamide, 0.1% naphthylethylenediamine dihydrochloride, 2.5% H₃PO₄) was added into each well in order to measure the nitrite content. After chromofore was formed at room temperature in 10 min, absorbance at 577 nm was measured in a microplate reader [7, 28, 29].

2.7. Estimation of total phenol content

Antioxidant compounds generally contain phenolic group(s) and the amount of phenolic compounds in the extract was estimated by using Folin–Ciocalteu reagent. Briefly, 10 µL sample or standard (50-500 mg/L gallic acid) plus 150 µL diluted Folin–Ciocalteu reagent (1:4 reagent:water) was placed in each well of a 96-well plate, and incubated at room temperature for 3 min. Following addition of 50 µL sodium carbonate (2:3 saturated sodium carbonate: water) and a further incubation of 2 h at room temperature, absorbance was read at 725 nm. The total phenolic content was expressed as gallic acid equivalents (GAE) in milligrams per gram extract. All tests were conducted in triplicate [30, 31].

3. Results and Discussion

Involvement of free radical mediated cell damage in many different diseases and wide usage area of the *Veronica* species has led us to determine radical scavenging activity of fourteen *Veronica* species collected from different location in Turkey. Radical scavenging activity of the aqueous extracts was screened against DPPH, NO and SO radicals. All the tested extracts were found to show dose dependent DPPH radical scavenging ability from the concentration of 50 µg/mL. While they show strong radical scavenging activity against DPPH radical, their activity is not stronger than that of standard compounds BHA and ascorbic acid. The highest IC₅₀ value was found for *V. officinalis* as 40.93 µg/mL. *V. cuneifolia* subsp. *cuneifolia*, *V. serpyllifolia*, *V. unknown*, *V. chamaedrys*, *V. baranetzki* and *V. peduncularis* showed also high IC₅₀ lower than 100 µg/mL (Table 2).

Nitric oxide (NO) is a small molecule that contains one unpaired electron, generated in biological system by nitric oxide synthase. Nitric oxide is an abundant reactive radical that acts as an important oxidative biological signaling molecule in a variety of physiological process, such as blood pressure regulation, defence mechanism against pathogens, smooth muscle relaxation and immune regulation. However overproduction of NO may occur when the generation of nitrogen species exceeds the system's ability to neutralize and eliminate them. Increased NO concentration may lead to nitrosylation reactions that can alter the structure of protein and inhibit their normal function. When super oxide reacts with NO, it produces more oxidatively active molecules peroxy nitrite ion (ONOO⁻), which causes DNA fragmentation and lipid peroxidation [26, 32]. NO scavenging effect of the aqueous extracts determined using Griess reagent. As seen on the Table 2, tested extracts were also showed dose dependent NO radical scavenging activity. While their IC₅₀ values were very high comparing to DPPH radical scavenging activity, their effects were found comparable to that of known antioxidants BHA, quercetin and ascorbic acid (AA). *V. officinalis* showed highest NO scavenging

capacity (IC₅₀ 570.33 µg/mL) and *V. chamaedrys* and *V. serpyllifolia* showed also high IC₅₀ values comparing to other tested extracts (IC₅₀: 651.98 and 697.09 µg/mL, Table 2).

Table 2. DPPH, NO and SO radical scavenging activity and total phenolic contents of the *Veronica* species.

Tested species	IC ₅₀ (µg/ml)			Gallic acid equivalent total phenolic content (mg/g dry extract)
	DPPH	NO	SO	
<i>V. orientalis</i>	103.63	824.24	198.91	127.64
<i>V. peduncularis</i>	54.19	770.01	186.31	139.92
<i>V. officinalis</i>	40.93	570.33	223.44	200.20
<i>V. baranetzki</i>	99.04	< 900	330.40	83.15
<i>V. chamaedrys</i>	57.31	651.98	113.39	79.50
<i>V. unknown</i>	63.03	727.84	< 900	98.50
<i>V. fuhsii</i>	117.99	< 900	697.05	46.52
<i>V. serpyllifolia</i>	53.96	697.09	475.23	78.11
<i>V. cuneifolia</i>	76.09	-	214.33	91.02
<i>V. persica</i>	154.49	811.43	142.81	82.04
<i>V. polita</i>	161.61	700.10	143.91	63.75
<i>V. hederifolia</i>	274.1	< 900	540.54	40.90
<i>V. anagallis-aquatica</i>	267.83	< 900	393.09	50.87
<i>V. cymbalaria</i>	390.09	< 900	588.89	-
BHA	> 10	306.64	502.47	
AA	> 10	< 900	13.36	
Quercetin	> 10	658.85	136.6	

On the other hand, NBT assay was carried out to test whether the extracts scavenge superoxide anions or not. Alkaline DMSO, used as a superoxide generating system, reacts with NBT to give colored diformazan. SO radical scavenging activity of the extracts was observed from the concentration of 25 µg/mL and *V. chamaedrys* showed the highest scavenging activity with IC₅₀ 113.39 µg/mL. When the results compared with the activity of standard compounds, only AA showed higher activity, quercetin (IC₅₀ 136.6 µg/mL) and BHA (IC₅₀ 502.47 µg/mL) showed lower scavenging activity than *V. chamaedrys* (Table 2). Since superoxide anion is the most common free radicals *in vivo*, a precursor for other ROS such as hydrogen peroxide, hydroxyl radical and singlet oxygen that have the potential reactivity with biological molecules and inducing tissue damage, the concentration of superoxide anions is very important under conditions of oxidative stress.

Table 3. Isolated iridoid glycosides from tested species (1. *V. cuneifolia*, 2. *V. cymbalaria*, 3. *V. persica*, 4. *V. hederifolia*, 5. *V. polita*, 6. *V. anagallis-aquatica* 7. *V. fuhsii*, 8. *V. serpyllifolia*)

	1	2	3	4	5	6	7	8
Aucubin [1]	+	+	+	+	+			
Catalpol [2]	+	+						+
Veronicoside [3]	+	+	+	+	+	+		+
Catalposide [4]	+	+	+	+	+	+		+
Verproside [5]	+	+	+				+	+
Amphicoside [6]	+	+	+	+	+			
6-O-veratroyl catalposide [7]		+	+					+
6-O-izovanilloil katalpol [8]		+						
Verminoside [9]	+	+	+	+	+	+	+	
8-epiloganin [10]	+	+			+			
6'-O-benzoyl-8-epiloganic acid [11]							+	
6'-O-p-hydroxybenzoyl-8-epiloganic acid [12]							+	
6'-O-benzoyl-gardoside [13]							+	
Urphoside A [14]				+				
Pikuroside [15]				+				

Concerning DPPH, NO and SO radical scavenging activity of the tested extracts, DPPH radicals found to be the most scavenged radicals. Radical scavenging activity of the extracts indicated the presence of high phenolic contents of the extract. It is obvious that the total phenolic content measured by the Folin–Ciocalteu procedure does not give a full picture of the quality or quantity of the phenolic constituents in the extracts. However, correlation between the radical scavenging activity and the total phenolic contents of the different extracts was shown in a lot of different study [29]. Therefore, total polyphenols in the extracts were determined by Folin-Ciocalteu reagent. This assay has been extensively used to measure the total phenolics in plant materials for many years and based on electron transfer reaction and actually measures a sample's reducing capacity. Therefore, it is accepted as a routine assay for rough estimation of the antioxidant capacity of herbal samples. Here, total phenolic content of the aqueous extracts was expressed as gallic acid equivalent in mg/g dry extract. As shown in Table 2, highest phenolic content was determined for *V. officinalis*, 202.20 mg/g dry extract. Since *V. officinalis* has been found to show strongest DPPH and NO radical scavenging activity, direct correlation between the high phenolic content of the plant and its radical scavenging activity indicated that radical scavenging activity of *V. officinalis* was arising from phenolic content of the plant. On the other hand, radical scavenging activity of tested species was found close to each other against NO and SO radicals. Well known high ester-iridoid and phenylethanoid contents for *Veronica* species, and above results show us to the importance of phenolic constituents for the

scavenging activity [7,8,18]. In addition, since highest phenolic content was found for *V. officinalis* together with its strongest DPPH and NO scavenging capacity, bioactivity guided isolation studies will be performed on this species to find the responsible compounds. In our previous studies on genus *Veronica*, *V. cuneifolia* subsp. *cuneifolia*, *V. cymbalaria*, *V. persica*, *V. hederifolia*, *V. polita*, *V. anagallis-aquatica* and *V. fuhsii* were studied phytochemically. As seen on Table 3, their iridoid contents are very close to each other with minor differences. We have mostly focused on iridoid contents of the plants up to now because of their chemotaxonomic importance. In future studies, it should be investigated for phenolic contents of the *Veronica* species together with their iridoid contents. Our phytochemical and biological researches on the member species of Plantaginaceae family are still continuing and further researches are needed to clarify chemotaxonomic and biologic significance of these species

Acknowledgments

This study was financially assisted by The Scientific and Technological Research Council of Turkey (TUBITAK Project No: 108T518).

References

- [1] Davis PH (1978) Flora of Turkey and the East Aegean Islands, Vol. 6. pp 325-326 University Press, Edinburgh.
- [2] S.R. Jensen, D.C. Albach, T. Ohno and R.J. Grayer (2005). *Veronica*: Iridoids and cornoside as chemosystematic markers, *Biochem. Syst. Ecol.* **33**, 1031-1047.
- [3] R.M. Taskova, D. Peev, and N. Handjieva, (2002). Iridoid glucosides of the genus *Veronica* s.l. and their systematic significance, *Pl. Syst. Evol.* **231**, 1-17.
- [4] R.J. Grayer-Barkmeijer (1978). Flavonoids in *Parahebe* and *Veronica*: a chemosystematic study, *Biochem. Syst. Ecol.* **6**, 131-137.
- [5] D.C. Albach, R.J. Grayer, S.R. Jensen, F. Ozgokce and N.C. Veitch (2003). Acylated flavone glycosides from *Veronica*, *Phytochemistry* **64**, 1295-1301.
- [6] V.M. Chari, R.J. Grayer-Barkmeijer, J.B. Harborne and B.G.O. Sterdahl (1981). An acylated allose-containing 8-hydroxyflavone glycoside from *Veronica filiformis*, *Phytochemistry* **20**, 1977-1979.
- [7] U.S. Harput, I. Saracoglu, M. Inoue and Y. Ogihara (2002). Antiinflammatory and cytotoxic activities of five *Veronica* species, *Biol. Pharm. Bull.* **25**, 483-486.
- [8] U.S. Harput, I. Saracoglu, A. Nagatsu, Y. Ogihara (2002). Iridoid glucosides from *Veronica hederifolia*, *Chem. Pharm. Bull.* **50**, 1106-1108.
- [9] Saracoglu, I., Harput, U.S., Inoue, M. and Ogihara, Y. (2002). New phenylethanoid glycosides from *Veronica pectinata* var. *glandulosa* and their free radical scavenging activities. *Chem. Pharm. Bull.*, **50**, 665-668.
- [10] R.M. Taskova, C.H. Gottfredsen and S.R. Jensen (2006). Chemotaxonomy of Veroniceae and its allies in the Plantaginaceae, *Phytochemistry* **67**, 286 - 301.
- [11] D.C. Albach and M.W. Chase (2001). Paraphyly of *Veronica* (Veroniceae; Scrophulariaceae): evidence from the internal transcribed spacer (ITS) sequences of nuclear ribosomal DNA, *J. Plant Res.* **114**, 9-18.
- [12] R.M. Taskova, C.H. Gottfredsen and S.R. Jensen (2005). Chemotaxonomic markers in Digitalideae (Plantaginaceae), *Phytochemistry* **66**, 1440 - 1447.
- [13] T. Baytop (1999). *Türkiye'de Bitkiler ile Tedavi (Geçmişte ve Bugün) (2. Baskı)*. Nobel Tıp Kitabevleri, İstanbul.
- [14] T. Fujita, E. Sezik, M. Tabata, E. Yesilada, G. Honda, Y. Takeda, T. Tanaka and Y. Takaishi (1995). Traditional medicine in Turkey VII. Folk medicine in Middle and West Black Sea Regions, *Economic Botany* **49**, 406-422.
- [15] L. Tomassini, D. Brkic, M. Serafini and M. Nicoletti (1995). Constituents of *Veronica hederifolia* and *Veronica polita*, *Fitoterapia* **66** (4), 382.
- [16] J.G. Graham, M.L. Quinn, D.S. Fabricant and N.R. Farnsworth (2000). Plants used against cancer-an extension of the work of Jonathan Hartwell, *J Ethnopharmacol.* **73**, 347-377.

- [17] U.S. Harput, A. Nagatsu, Y. Ogihara and I. Saracoglu (2003). Iridoid glucosides from *Veronica pectinata* var. *glandulosa*, *Z. Naturforsch.* **58c**, 481-484.
- [18] I. Saracoglu, M. Varel, US Harput and A. Nagatsu (2004) Acylated Flavonoids and Phenol Glycosides *Veronica thymoides* subsp. *pseudocinerea*. *Phytochemistry*, **65**, 2379-2385.
- [19] E.A. Asongalem, H.S. Foyet, S. Ekobo, T. Dimo and P. Kamtchouing (2004). Antiinflammatory, lack of central analgesia and antipyretic properties of *Acanthus montanus* (Ness) T. Anderson, *J. Ethnopharmacol* **95**, 63-68.
- [20] F.T. Slater (1984). Free-radical mechanism in tissue injury, *J.Biochem.* **222**,1-15.
- [21] M.S. Willis and F.H. Wians (2003). The role of nutrition in preventing prostate cancer: a review of the proposed mechanism of action of various dietary substances, *Clinica Chimica Acta* **330**, 57-83.
- [22] T. Hatano, R. Edamatsu, M. Hiramatsu, A. Mori, Y. Fujita, T. Yasuhara, T. Yoshida and T. Okuda (1989). Effects of the interaction of tannins with co-existing substances. VI. Effects of tannins and related polyphenols on superoxide anion radical and on 1,1-diphenyl 2-picrylhydrazyl radical, *Chem. Pharm. Bull.* **37**, 2016–2021.
- [23] E.P. Kostadinova, K.I. Alipieva, T. Kokubun, R.M. Taskova and N.V. Handjieva (2007). Phenylethanoids, iridoids and a spirostanol saponin from *Veronica turrilliana*, *Phytochemistry* **68**, 1321–1326.
- [24] K. Elizabeth and M.N.A. Rao (1990). Oxygen radical scavenging activity of curcumin, *Int. J. Pharm.* **58**, 237–240.
- [25] R. Srinivasan, M.J.N. Chandrasekar, M.J. Nanjan and B. Suresh (2007). Antioxidant activity of *Caesalpinia digyna* root, *J. Ethnopharmacol.* **113**, 284–291.
- [26] B. S. Reddy, R. K. K. Reddy, B. P. Reddy, S. Ramakrishna and P. V. Diwan. (2008) Potential in vitro antioxidant and protective effects of *Soymida febrifuga* on ethanol induced oxidative damage in HepG2 cells, *Food Chem. Tox.*, **46**, 3429-3442.
- [27] P. Klatt and S. Lamas (2000) Regulation of protein function by S-glutathiolation in response to oxidative and nitrosative stress. *European Journal of Biochemistry*, **267**, 4928–4944.
- [28] K. Hensley, S. Mou and Q.N. Pye (2003). Nitrite Determination by Colorimetric and Fluorometric Griess Diazotization Assays *Methods in Pharmacology and Toxicology: Methods in Biological Oxidative Stress* Edited by: K. Hensley and R. A. Floyd pp 185-193 © Humana Press Inc., Totowa, NJ.
- [29] P. Tsai, T.H. Tsai, C. Yu and S.C. Ho (2007). Comparison of NO-scavenging and NO-suppressing activities of different herbal teas with those of Green Tea, *Food Chem.* **103**, 181–187.
- [30] M. Dominguez, A. Nieto, J.C. Marin, A.S. Keck, E. Jeffery and C.L. Cespedes (2005). Antioxidants activities of extracts from *Barkleyanthus salicifolius* (Asteraceae) and *Penstemon gentianoides* (Scrophulariaceae), *J. Agric. Food Chem.* **53**, 5889–5895.
- [31] C.L. Cespedes, M. El-Hafidi, N. Pavon, J. Alarcon (2008). Antioxidant and cardioprotective activities of phenolic extracts from fruits of Chilean Blackberry *Aristotelia chilensis* (Elaeocarpaceae), Maqui”, *Food Chem.* **107**, 820-829.
- [32] Valko, M., Leibfritz, D., Moncola, J., Cronin, M.T.D., Mazura, M., Telser, J. (2007). Free radicals and antioxidants in normal physiological functions and human disease. *Int. J. Biochem. Cell Biol.* **39**, 44–84.