

Rec. Nat. Prod. 7:2 (2013) 137-140

records of natural products

Antiproliferative Activity and Constituents of *Aspidosperma*macrocarpon (Apocynaceae) Leaves

Geanderson Bannwart¹, Cecília M. A. de Oliveira², Lucilia Kato², Cleuza C. da Silva¹, Ana Lúcia T. G. Ruiz³, João E. de Carvalho³ and Silvana M. O. Santin^{1*}

(Received May 5, 2012; Revised May 22, 2012; Accepted January 17, 2013)

Abstract: Aspidosperma macrocarpon belongs to the family Apocynaceae and is endemic to Americas and mainly found from Mexico to Argentina. It is known in Brazil as "guatambu" or "peroba". Crude extracts and their fractions from leaves were assayed against human cancer cells lines: glioma (U251), melanoma (UACC-62), mammary (MCF-7), ovarian expressing the multidrug resistance phenotype (NCI-ADR/RES), lung (NCI-H460), prostate (PC-3), kidney (786-0), ovarian (OVCAR-3), colon (HT-29) and leukemia (K-562). The crude extract (EAM), hexane (HA) and chloroform (CA) fractions were the most active fractions against K-562 with GI50 values low than 1 μ g/mL. Also, CA was moderate active against OVCAR-3 and NCI-ADR/RES cells lines. This phytochemical study allowed to identify the known kopsanone, kopsinine, ursolic acid, rutin, 5-*O*-caffeoylquinic acid, and 3,5-*O*-dicaffeoylquinic acid. The kopsanone was also evaluated against human cancer cell lines and showed activity to the U251 and K-562 cell lines, with GI50 values of 20.6 μ g/mL and 8.7 μ g/mL, respectively.

Keywords: Aspidosperma macrocarpon; antiproliferative activity; alkaloids.

1. Plant Source

As part of our continuing search for antiproliferative agents derived from natural herbals resources, and assessment of the efficacy crude drugs used by traditional communities in Brazil, extract and fractions from leaves of *Aspidosperma macrocarpon* were assayed against ten human cancer cell lines. We report here the bioactivity of these extracts, fractions and the alkaloid kopsanone as well as the isolation of secondary metabolites.

Leaves of *Aspidosperma macrocarpon* were collected on May 2010 in the municipality of Goiânia (Goiás, Brazil), and identified by Dr. Heleno Dias Ferreira. A voucher specimen (45524) was deposited in the Herbarium of the Instituto de Biologia, Universidade Federal de Goiás (ICB/UFG).

_

¹Departamento de Química, Universidade Estadual de Maringá, Av. Colombo 5790, 87020-900, Maringá, Paraná, Brazil

²Instituto de Química, Universidade Federal de Goiás, Campus Samambaia; P.O. Box 131,74001-970, Goiânia, Goiás, Brazil

³Centro Pluridisciplinar de Pesquisa Químicas, Biológicas e Agrícolas, Universidade Estadual de Campinas, P.O. Box 6171, 13081-970, Campinas, São Paulo, Brazil.

^{*} Corresponding author: E-Mail: smoliveira@uem.br

2. Previous Studies

The alkaloids (-)-vincadifformine, ervinceine, kopsanone, kopsinine, kopsanol and 18-epikopsanol have been isolated from seeds and stem bark of *Aspidosperma macrocarpon* [2-3].

3. Present Study

Dried powdered leaves (738.9 g) of *Aspidosperma macrocarpon* were exhaustively extracted by maceration with methanol at room temperature. Evaporation of the solvent yielded the crude extract (**EAM** 139.0 g).

Part of the crude extract (14.2 g) was re-dissolved in MeOH- H_2O 1:1 (50 mL) and HCl 0.1 M (100 mL). The acid solution was partitioned with CHCl₃ (5x25 mL) yielded a chloroform fraction (1.35 g). The acid solution fraction was basified with NH₄OH (pH 9.0) and partitioned with CHCl₃ (3 x 70 mL) yielding alkaloid fraction (**AF**, 0.57 g). The alkaloid fraction (0.47 g) was fractionated on silica gel column eluted with hexane: CHCl₃ (10-100%), CHCl₃ and CHCl₃: MeOH (10-100%), resulting in 55 sub-fractions. The sub-fractions AM-14 and AM-15 (hexane: CHCl₃ 4:6) yielded kopsanone (45.8 mg, **1**) and kopsinine (9,4 mg, **2**).

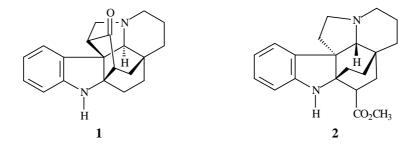


Figure 1. Structures of alkaloids from leaves of A. macrocarpon.

Part of the crude extract (11.5 g) was dissolved in MeOH-H₂O 1:1 and partitioned with different solvents to obtain the hexane (**HA**, 1.30 g), chloroform (**CA**, 3.45 g), ethyl acetate (**EA**, 1.58 g) and hydromethanolic (**HMA**, 3.40 g) fractions. The resulting fractions were subjected to conventional purification procedures and leading to isolation of ursolic acid, 5-O-caffeoylquinic acid, rutin and 3,5-O-dicaffeoylquinic acid.

Cell lines and culture medium: The in vitro antiproliferative activity of the extracts and fractions was assessed using ten different human cancer cell lines: U251 (glioma), UACC-62 (melanoma), MCF-7 (mammary), NCI-ADR/RES (ovarian expressing the multidrug resistance phenotype), NCI-H460 (lung), PC-3 (prostate), 786-0 (kidney), OVCAR-3 (ovarian), HT-29 (colon) K-562 (leukemia) and normal cell line HaCaT (human keratinocyte) were kindly provided by Frederick Cancer Research & Development Center, National Cancer Institute, Frederick, MA, USA. The chemotherapeutic agent doxorubicin was used as a positive control. Stock cultures were grown in a medium containing 5 mL of RPMI 1640 (GIBCO BRL, Life Technologies) and supplemented with 5% of foetal bovine serum. Gentamicine (50 μg/mL) was added to the experimental cultures.

Cytotoxicity assay: Cells in 96 well-plates (100 μ L cells/well) were exposed to varying concentrations of samples in DMSO (0.25; 2.5; 25 and 250 μ g/mL) and 5% CO₂ in air for 48 h at 37 °C. The final concentration of DMSO did not affect the cell viability. A 50% trichloroacetic acid solution was added and incubated with the cells for 30 min at 4 °C. After washing and drying, the degree of cell proliferation was determined by spectrophotometric quantification (540 nm) of the cellular protein content, using the sulforhodamine B assay. Doxorubicin (DOX; 0,025-25 μ g/mL) was used as positive control. Three measurements were obtained at the beginning of incubation (time zero, T_0) and 48h post-incubation for sample-free (C) and tested (T) cells. Cell proliferation was determined according to the equation $100x[(T-T_0)/C-T_0]$, for $T_0 < T \le C$, and $100x[(T-T_0)/T_0]$ for $T \le T_0$ and a concentration-response curve for each cell line was plotted using software Origin 7.5 (OriginLab Corporation) [4].

Using the concentration-response curve for each cell line, GI_{50} (concentration causing 50% growth inhibition) was determined by means of non-linear regression analysis, using software Origin 7.5 (Origin Corporation). The average activity (mean of log GI_{50}) of the extracts tested was also determined using MSExcel software [5]. (**Table 1**)

 $\textbf{Table 1}. \ \text{Antiproliferative activity } (GI_{50}, \mu g.mL^{\text{--}1}) \ \text{of leaves crude extract, fractions and kopsanone of }$

Aspidosperma macrocarpon on different cancer cell lines.

_11	U251	UACC-	MCF-	NCI-	786-	NCI-	PC-3	OVCAR-	HT-	K562	HaCaT	Mean
		62	7	ADR/RES	0	H460		3	29			log
												GI_{50}
Doxo	0,027	0,025	<0,025	0,15	0,051	<0,025	0,12	0,26	0,10	0,049	0,053	<-1,3 P
EAM	67,0	20,4	26,0	20,0	51,2	38,4	61,8	23,5	130,0	0,51	1,5	1,3 W
AF	26,6	27,3	28,6	13,6	29,1	37,1	40,9	9,7	89,8	2,6	<0,25	<1,0 M
HA	24,7	23,3	9,2	8,8	25,1	25,8	25,5	22,7	27,2	0,78	2,9	1,1 M
CA	13,3	38,7	7,6	1,6	3,1	22,0	7,5	1,2	69,4	0,36	9,1	0,8 M
EA	48,5	38,1	49,7	11,5	31,7	83,1	120,6	25,0	88,0	3,0	8,5	1,5 W
HMA	>250	>250	55,6	56,1	>250	>250	>250	147,9	159,6	40,6	>250	>2,2 I
kopsanone	20,6	38,9	54,7	56,0	66,4	96,7	79,4	47,2	56,0	8,7	60,9	1,7 I

Cell lines: U251 (glioma); UACC-62 (melanoma); MCF-7 (mammary); NCI-ADR/RES (ovarian expressing the multidrug resistance phenotype); 786-0 (kidney); NCI-H460 (lung); PC-3 (prostate); OVCAR-3 (ovarian); HT-29 (colon); K562 (leukemia); HaCat (human keratinocyte, normal cell).

NCI's critera: W, weak activity: $1.5 \ge log~GI_{50} > 1.10$; M, moderate activity: $1.1 \ge log~GI_{50} > 0$; P, potent activity: $log~GI_{50} < 0$. (Foucher et al, 2008).

The crude methanolic extract from leaves (**EAM**) exhibited a high inhibitory effect on cell growth, and was very effective against the UACC-62, MCF-7, NCI-ADR/RES and K-562 cell lines, with GI_{50} values of 20.4, 26.0, 20.0 and 0.51 µg/mL, respectively (Table 1). This crude extract was further separated into five fractions, alkaloid fraction (**AF**), hexane (**HA**), chloroform (**CA**), ethyl acetate (**EA**) and hydromethanolic (**HMA**) fractions, which were assayed against the same cells. **AF** fraction presented high activity in most of the cell lines tested. The highest antiproliferative activities of fraction was found against the NCI-ADR/RES, OVCAR-3 and K-562 cells, with GI_{50} values of 13.6, 9.7 and 2.6 µg/mL, respectively. A significant amount of the alkaloid kopsanone (**1**, 45.8 mg) was isolated from **AF**, along with kopsinine (**2**). These compounds were identified by analysis of their spectroscopic data and comparison with available literature data [6-7]. The alkaloid **1** also was assessed and showed high selectivity and activity against the U251 and K-562 cell lines, with GI_{50} values of 20.6 µg/mL and 8.7 µg/mL, respectively.

HA fraction showed potent activity against MCF-7, NCI-ADR/RES and K-562 cell lines, with GI₅₀ values of 9.2, 8.8 and 0.78 μg.mL⁻¹, respectively. Toward most cell lines, GI₅₀ values of the CA were lower than those of HA. This fraction showed highest activity against MCF-7, NCI-ADR/RES, 786-0, PC-3, OVCAR-3 and K562 cell lines, with GI₅₀ values of 7.6, 1.6, 3.1, 7.5, 1.2 and 0.36 μg/mL, respectively. Phytochemical investigation showed the presence of an expressive amount of the ursolic acid triterpene in this fraction, identified by comparison of their spectroscopic data with those previously published [8-9]. In the literature, the ursolic acid is described as very active against a broad range of cancer cell lines [10]. The NCI-ADR/RES, OVCAR-3 and K562 were more sensitive to EA fraction than the other cells, with GI₅₀ values of 11.5, 25,0 and 3.0 μg/mL, respectively. Studies of EA fraction allowed identify the 5-*O*-caffeoylquinic acid, 3,5-*O*-dicaffeoylquinic acid and rutin, also identified by their spectroscopic data compared with available literature [11-12]. The chlorogenic acids and rutin have been reported as antioxidant and anti-inflammatory agents [13-14] and, as antioxidants, can inhibit carcinogenesis. Recently, the dicaffeoylquinic derivatives have been reported as potent antiproliferative agents against breast carcinoma (MCF-7), myeloid leukemia (HL-60), histiocytic lymphoma (U937), leukemia (HL-60), colon cancer (DLD-1) cell lines [11].

In this study we focused the evaluation of the extracts, fractions and the indol alkaloid from leaves of A. macrocarpon against a representative set of human cancer cell lines. The bio-guided

fractionation of the active crude extract showed that the hexane (HA), chloroform (CA) fractions were the most active fractions against the K-562 with GI_{50} values low than 1 μ g/mL. CA fraction showed moderate activity against OVCAR-3 and NCI-ADR/RES cells line. The phytochemical study allowed to identify the known kopsanone, kopsinine, ursolic acid, rutin, 5-O-caffeoylquinic acid, and 3,5-O-dicaffeoylquinic acid, identified by NMR data. The kopsanone was assayed against the U251 and K-562 cell lines, with GI_{50} values of 20.6 μ g/mL and 8.7 μ g/mL, respectively. Our results are in agreement with previous evidences which have shown that *Aspidosperma* species are likely sources of useful substances for the development of new drugs.

References

- [1] M. M. Pereira (2007). Alcalóides indólicos isolados de espécies do gênero *Aspidosperma* (Apocynaceae), *Quim. Nova.* **30**, 970-983.
- [2] A. C. Mitaine, K. Mesbash, C. Petermann, S. Arrazola, C. Moretti and M. Zeches-Hanrot (1996). Alkaloides from *Aspidosperma* species from Bolívia, *Planta Med.* **62**, 458-461.
- [3] M. Ferreira Filho, B. Gilbert, M. Kitagawa, L. A. Paes Leme and L. J. Durham (1966). Four heptacyclic alkalois from *Aspidosperma* species, *J. Chem. Soc.* (C), 1260-1266.
- [4] A. Monks, D. Scudeiro, P. Skehan, R. Shoemaker, K. Paull, D. Vistica, C. Hose, J. Langley, P. Cronise, A. Vaigro-Wolff, M. Gray-Goodrich, H. Campbell, J. Mayo and M. Boyd (1991). Feasibility of a high-flux anticancer drug screen using a diverse panel of cultured human tumor cell lines, *J. Natl. Cancer*. **183,** 757-766.
- [5] G. Foucher, G. M. Cragg, P. Pillay, N. Kolesnikova, V. J. Maharaj and J. Senabe (2008). *In vitro* anticancer screening of South African plants, *J. Ethnopharmacol.* **119**, 455-461.
- [6] X. Z. Feng (1984). Kopsoffine: A new dimeric indole alkaloid of pleiomutine type from *Kopsia officinalis*, *J. Nat. Prod.* **47**, 117-122.
- [7] M. E. Kuehne and P. J. Seaton (1985). Studies in biomimetic alkaloid syntheses. 13. Total syntheses of racemic aspidofractine, pleiocarpine, pleiocarpinine, kopsinine, *N*-methylkopsanone and kopsanone, *J. Org. Chem.* **50**, 4790-4796.
- [8] G. F. Lemes and P. H. Ferri (2011). Constituintes químicos de *Hyptidendron canum* (Pohl ex Benth.) R. Harley (Lamiaceae), *Quim. Nova.* **34,** 39-42.
- [9] S. B. Mahato and A. P. Kundu (1994). ¹³C NMR Spectra of pentacyclic triterpenoids–a complication and some salient features, *Phytochemistry*. **37**, 1517-1573.
- [10] J. Liu (1995). Pharmacology of oleanoic acid and ursolic acid, J Ethnopharmacol. 49, 57-68.
- [11] A. R. Santos, M. P. Barros, M. H. Sarragiotto and S. M. O. Santin (2004). Constituintes polares das folhas de *Machaonia brasiliensis* (Rubiaceae), *Quim. Nova.* **27**, 525-527.
- [12] N. Didry, V. Seidel, L. Dubreuil, F. Tillequin and F. Bailleaul (1999). Isolation and antibacterial activity of phenylpropanoid derivatives from *Ballota nigra*, *J Ethnopharmacol*. **67**, 197-202.
- [13] S. Puangpraphant, M. A. Berhow, K. Vermillion, G. Potts and E. G. Mejia (2011). Dicaffeoylquinic acids in yerba mate (*Ilex paraguariensis* St. Hilaire) inhibit NF-kB nucleous translocation in macrophages and induce apoptosis by activating caspases-8 and -3 in human colon cancer cells. *Mol. Nutr. Food Res.* 55, 1509-1522.
- [14] R. J. Nijveldt, E. V. Nood, D. E. V. Hoom, P. G. Boelens, K. V. Norren and P. A. V. Leeuwen (2001). Flavonoids: a review of probable mechanisms of action and potential applications, *Am. J. Clin. Nutr.* **74**, 418-425.



© 2013 Reproduction is free for scientific studies