

**Phytotoxic and Antimicrobial Metabolites from
Paraphaeosphaeria sp. QTYC11 Isolated from the Gut of
Pantala flavescens Larvae**

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Abstract: A new lunatoic acid C (**1**) along with eight known compounds were isolated from *Paraphaeosphaeria* sp. QTYC11 residing in the gut of *Pantala flavescens* larvae. They were determined on the basis of extensive spectroscopic analysis and by comparison of the corresponding data reported previously. Compounds **1** showed good phytotoxic activities, extremely potent antifungal activities and good antibacterial activities respectively.

Keywords: *Paraphaeosphaeria* sp.; natural products; antifungal activity; antibacterial activity; phytotoxic activity. © 2015 ACG Publications. All rights reserved.

1. Introduction

Microorganisms keep interaction between different insect species help shape the biological complexity and diversity of the planet. The class insects have 900 000 known and 2-30 million predicted species that occupied a diverse range of ecological niches, often facilitated by their endosymbionts [1]. Such genetically tractable microorganisms undergo coevolution with the insect by producing small molecules, supporting nutrient or protection [2]. Irrespective of their role in the complicated systems, microbes associated with the insect can be regarded as a huge reservoir for pharmaceutically agents [3,4]. In our previous study, two microorganisms *Daldinia eschscholzii* and *Pestalotiopsis* sp. HC02 have been isolated from mantis (*Tenodera aridifolia*) and grasshopper (*Chondracris rosee*), respectively. Further chemical isolation revealed several new metabolites, which had potential to be used as immunosuppressants and herbicides [5,6]. With the interest to search for new bioactive metabolites from the insect gut microbiota, we isolated one endosymbiont and identified

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it as *Paraphaeosphaeria* sp. QTYC11, which was isolated from the gut of *Pantala flavescens* larvae. The ethyl acetate extract from the culture filtrate of the QTYC11 exhibited good phytotoxic activity against the radical growth of *Echinochloa crusgalli* and *Amaranthus retroflexus*, and antifungal activities to several plant pathogens as well as antibacterial activities against a panel of food spoiling microorganisms. Subsequently, one lunatoic acid C (**1**) together with 8 known biosynthetic relatives was isolated and identified from the ethyl acetate crude extract. Herein we reported the details of the isolation, structure elucidation, and biological activities of all the tested metabolites are presented.

2. Materials and Methods

2.1. Microorganism Material

According to the methods detailed elsewhere [7], the strain QTYC11 was isolated from the gut of *P. flavescens* larvae collected from the river near by the Zhejiang Normal University (29°00'17.37"N, 119°29'54.84"E), PR China, in August 2012. The title fungus QTYC11 was identified according to the morphological characteristics and 5.8S rDNA sequence to those of standard record. Preliminary identification of QTYC11 sequences were compared with those available in GenBank by using BLAST to determine its phylogenetic affiliation. Phylogenetic analysis was performed using the MEGA v5.0 after multiple alignments of data by CLUSTAL-X software [8]. Phylogenetic tree based on the neighbor-joining analysis was constructed by the model of (NJ) Kimura-2-parameter and gamma distribution [9]. Bootstrap analysis was used to evaluate the tree topology of the neighbor-joining data by performing 1,000 replicates. The phylogenetic tree (Fig. 1) inferred from the ribosomal DNA ITS sequences indicated that the QTYC11 closely related to *Paraphaeosphaeria* sp. DQ092509 with the ITS sequence similarity of 99.7%. Therefore, QTYC11 was identified as *Paraphaeosphaeria* sp. The fungal strain QTYC11 was deposited in the China Center for Type Culture Collection as CCTCC M 2013435 and the GenBank accession number was KJ469553.

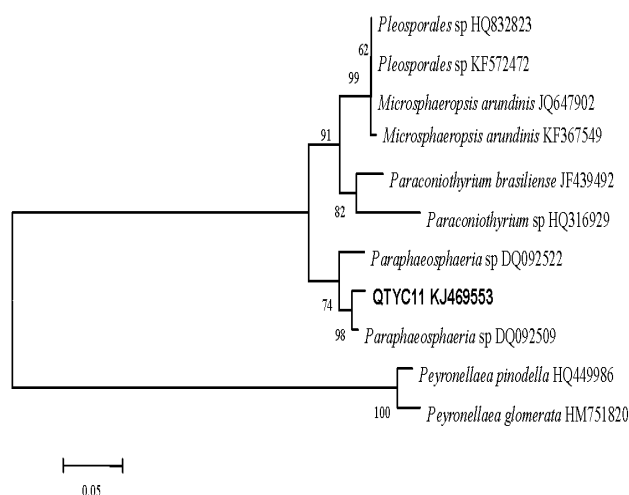


Figure 1. Phylogenetic tree of the QTYC11 based on 5.8S and ITS regions sequences

2.2. Fermentation and Isolation

The fresh mycelium grown on Malt Extracts Agar medium at 28°C for 4 d was inoculated into 250 mL Erlenmeyer flask containing 100 mL of ME medium (20 g of malt extract, 20 g of sucrose, 1 g of peptone, 1 L of distilled H₂O). After 3 d of incubation at 28°C on rotary shakers at 180

rpm, 15 mL of culture liquid were transferred as seed into each 1000 mL Erlenmeyer flask containing 350 mL of ME medium, and cultivation was carried out at 28°C for 7 d.

The filtrate of the cultural broth (50 L) was extracted with ethyl acetate (6 × 50 L). Evaporation of menstruum *in vacuo* gave a yellow oily residue (32.6 g), which was subjected to column chromatography over silica-gel (SiO₂; 200-300 mesh; Qingdao Marine Chemical Ltd, Qingdao, China) eluting with CH₂Cl₂/MeOH mixtures of a stably growing polarity (100:0, 100:1, 100:2, 100:4, 100:8 v/v) to afford five parts (F1, 3.2 g; F2, 4.6 g; F3, 8.9 g; F4, 2.3 g; F5, 2.3 g). Compound **5** (1530 mg) was crystallized from the MeOH solution of subfraction F1. The fraction of F2 was purified by Sephadex LH-20 using a CH₂Cl₂/MeOH mixture (1:1) as the eluent to give the compound **9** (9 mg). Fraction (F3) was further fractionated over silica gel using CH₂Cl₂/MeOH (100:2) to give five subfractions (R1-R5), compound **1** (1356 mg), compound **4** (10.6 mg) and compound **6** (30.3 mg) were crystallized from the MeOH solution of subfraction R1, R3, R4 respectively. R2 was further chromatographed over silica gel (CH₂Cl₂/MeOH, 100:0-100:2) to give compound **7** (15 mg), **8** (6 mg). F4 (CH₂Cl₂/MeOH, 100:8) was repeatedly purified on Sephadex LH-20 (MeOH) and detected by TLC to yield compound **3** (56 mg). The remaining F5 was purified by Sephadex LH-20 to give the compound **2** (13 mg).

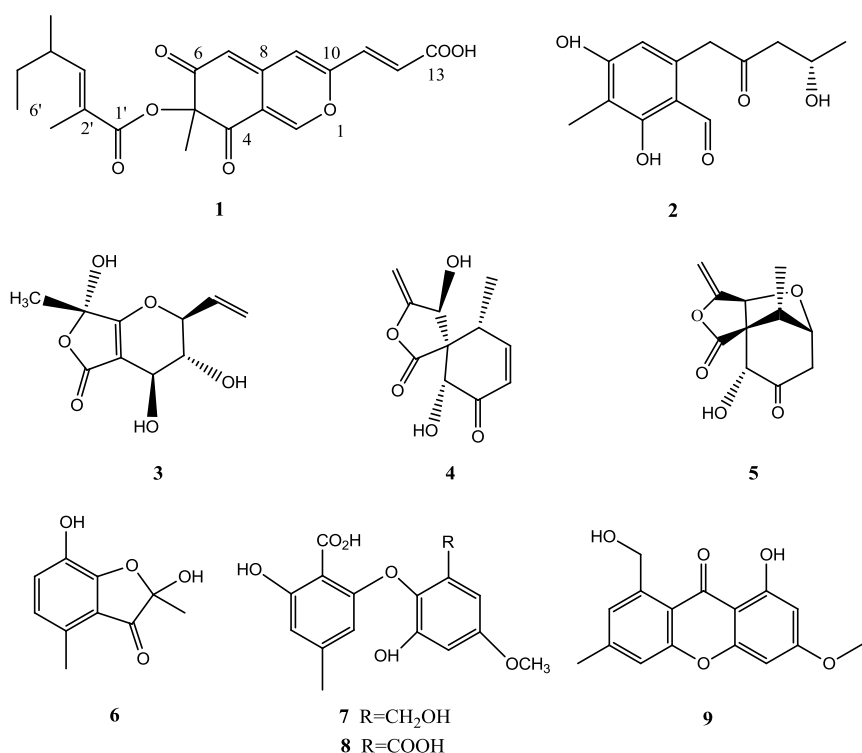


Figure 2. Structures of Compounds **1-9**

3. Results and Discussion

3.1. Structure elucidation

Compound **1** was obtained as orange powder and its molecular formula C₂₁H₂₂O₇ was deduced from HR-ESI-MS (409.1259 [M+Na]⁺, calculated for, 409.1258) and ¹³C NMR spectra. The ¹H NMR (Table 1) of **1** indicated the presence of four methyls (δ_H 0.88, 1.02, 1.60 and 1.87), one methylene (δ_H 1.40), one methine (δ_H 2.44), six olefinic methines (δ_H 5.78, 6.54, 6.55, 6.73, 7.23 and 7.94). The ¹³C NMR spectra (Table 1) of **1** showed 21 carbon signals in conjunction with DEPT spectrum indicated

the presence of four methyls, one methylenes, six olefinic methines, four quaternary olefinic carbons, one oxygenated quaternary carbon, one ester carbonyl and two enone carbonyls. The bicyclic bis-enone structure was established from the two and three-bond HMBC correlations of H-2 to C-3 (δ_C 114.9), C-4 (δ_C 193.4), C-8 (δ_C 140.8), and C-10 (δ_C 152.5); H-7 to C-3 (δ_C 114.9), C-5 (δ_C 84.0); H-9 to C-3, C-7 (δ_C 110.8), C-10, and C-11 (δ_C 135.2); H-11 to C-9, C-10, and C-13 (δ_C 169.5); H-12 to C-10 and C-13; and H₃-14 (δ_H 1.60) to C-5 and both enone carbonyls C-4 and C-6 (δ_C 192.9). The acyl moiety of **1** was clarified as follows: a triplet methyl at δ_H 0.88 ($J=7.4$ Hz), one doublet methyls at δ_H 1.02 ($J=6.6$ Hz), 1.19 ($J=7$ Hz), one single methyls δ_H 1.87 and the ^{13}C NMR spectra indicated the presence of two olefinic carbons, one ester carbonyl and HMBC correlations of H-3' to C-1' (δ_C 167.7) and C-2' (δ_C 124.9); H-4' to C-2', C-3' (δ_C 151.0), C-6' (δ_C 11.9) and C-8' (δ_C 19.4); H-5' to C-4' (δ_C 35.1); H-6' to C-4' and C-5' (δ_C 29.5); H-7' to C-1', C-2', C-3'; H-8' to C-3' thus establishing the structure as a derivative of lunatoic acid A [10].

The known compounds isolated from the *Paraphaeosphaeria* sp. QTYC11 were identified as 2,4-Dihydroxy-6-((R)-4-hydroxy-2-oxopentyl)-3-methylbenzaldehyde (**2**) [11], massarilactone D (**3**) [12] 6-epi-5'-hydroxymycosporulon (**4**) [13], massarilactone E (**5**) [14] enalin A (**6**) [15], barceloneic acid A (**7**), barceloneic acid B (**8**) [16] and 1-Hydroxy-8-(hydroxymethyl)-3-methoxy-6-methyl-9H-xanthen-9-one (**9**) [17] by comparison of their spectroscopic data with literature values.

Table 1. 1H NMR, ^{13}C NMR and HMBC data for compound **1** (at 600 MHz in acetone- d_6 , δ in ppm, J in Hz).

| Position | δ_C | δ_H , mult, J in Hz | HMBC |
|----------|------------|--------------------------------|------------------|
| 2 | 153.4 | 7.94 (1H, s) | C-3, 4, 8, 10 |
| 3 | 114.9 | | |
| 4 | 193.4 | | |
| 5 | 84.0 | | |
| 6 | 192.9 | | |
| 7 | 110.8 | 5.78 (1H, s) | C-3, 5 |
| 8 | 140.8 | | |
| 9 | 116.6 | 6.55 (1H, s) | C-3, 7, 10, 11 |
| 10 | 152.5 | | |
| 11 | 135.2 | 7.23 (1H, d, $J=15.6$) | C-9, 10, 13 |
| 12 | 123.6 | 6.54 (1H, d, $J=15.6$) | C-10, 13 |
| 13 | 169.5 | | |
| 14 | 21.8 | 1.60 (3H, s) | C-4, 5, 6 |
| 1' | 167.7 | | |
| 2' | 124.9 | | |
| 3' | 151.0 | 6.73 (1H dd, $J=10.1, J=1.2$) | C-1', 2' |
| 4' | 35.1 | 2.44 (1H, m) | C-2', 3', 6', 8' |
| 5' | 29.5 | 1.40 (2H, m) | C-4', 5' |
| 6' | 11.9 | 0.88 (3H, t, $J=7.3$) | C-4', 5' |
| 7' | 12.4 | 1.87 (3H, s) | C-1', 2', 3' |
| 8' | 19.4 | 1.02 (3H, d, $J=6.6$) | C-3' |

3.2. Phytotoxic and Antimicrobial activity

The phytogrowth inhibitory activity was evaluated on seedling of *A. retroflexus* and *E. crusgalli* using a Petri-dish bioassay, which was performed as described [18]. Antimicrobial assays were performed as described [19-21].

Compounds **1** and **6** showed good activities against the radical growth of *A. retroflexus* with the IC_{50} values of 10.2 and 27.6 $\mu g/mL$ respectively. The metabolites **1** and **4** had good phytotoxic activities against the radical growth of *E. crusgalli* with IC_{50} values of 5.9 and 16.0 $\mu g/mL$ respectively, and compound **1** showed moderate post-emergence activity for *E. crusgalli* with 46.0% decrease in dry weight. The ingredient **1** showed extremely potent antifungal activities against the *Valsa mali* ($IC_{50} = 3.3 \mu g/mL$), *Gibberella sanbinetti* ($IC_{50} = 1.7 \mu g/mL$) and *Fusarium oxysporum* f. sp. *cucumerinum* ($IC_{50} = 4.2 \mu g/mL$), which were comparable to those of referenced cycloheximide with IC_{50} values of 0.3, 3.3 and 4.9 $\mu g/mL$, respectively. The compound **1** also showed good antibacterial activities against *Staphylococcus aureus* and *Bacillus subtilis* with the inhibition zone of

17.1 and 28.3 mm, respectively. Based on this result, a patent had been applied in China (number of the patent application 201310729506.1).

Interestingly, lunatoic acid A was first reported with strong antifungal activity but bacterial growth was not affected at all [22]. Compound **1** as a derivative of lunatoic acid A was different from lunatoic acid A by an extra double bond on the fatty acid moiety of the molecule. However, lunatoic acid C exhibited moderate antibacterial activities against *S. aureus* and *B. subtilis*. It was obvious that the extra double bond on the fatty acid moiety of the molecule contribute to the antibacterial activities of family lunatoic acid. Whether the incorporation of an extra double bond on the fatty acid moiety into natural pharmacophore could improve antibacterial activity of medicines needed deep research.

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Supporting Information

Supporting Information accompanies this paper on <http://www.acgpubs.org/RNP>

References

- [1] J.M. Crawford and J. Clardy (2011). Bacterial symbionts and natural products, *Chem. Commun.* **47**, 7559-7566.
- [2] J. C. Brownlie and K. N. Johnson (2009). Symbiont-mediated protection in insect hosts, *Trends Microbiol.* **17**, 348-354.
- [3] D. C. Oh, M. Poulsen, C. R. Currie and J. Clardy (2009). Dentigerumycin: a bacterial mediator of an ant-fungus symbiosis, *Nat. Chem. Biol.* **5**, 391-393.
- [4] D. R. Colman, E. C. Toolson and C. D. Takacs-Vesbach (2012). Do diet and taxonomy influence insect gut bacterial communities, *Mol. Ecol.* **21**, 5124-5137.
- [5] Y. L. Zhang, J. Zhang, N. Jiang and R. X. Tan (2011). Immunosuppressive polyketides from mantis-associated *Daldinia eschscholzii*, *J. Am. Chem. Soc.* **133**, 5931-5940.
- [6] Y. L. Zhang, H. M. Ge, F. Li and R. X. Tan (2008). New phytotoxic metabolites from *Pestalotiopsis* sp. HC02, a fungus residing in *Chondracris roseae* gut, *Chem. Biodivers.* **5**, 2402-2407.
- [7] S. Li, M.W. Shao, Y.H. Lu, L.C. Kong, D.H. Jiang, and Y. L. Zhang (2014). Phytotoxic and antibacterial metabolites from *Fusarium proliferatum* ZS07 isolated from the gut of long-horned grasshoppers, *J. Agric. Food Chem.* **62**, 8997-9001.
- [8] C.J. Zheng, L.L. Xu, Y.Y. Li, T. Han, Q.Y. Zhang, Q.L. Ming and L.P. Qin (2013). Cytotoxic metabolites from the cultures of endophytic fungi from *Panax ginseng*, *Appl. Microbiol. Biot* **97**, 7617-7625.
- [9] P. Bhadury, H. Bik, J.D. Lamshead, M.C. Austen, G.R. Smerdon and A.D. Roger (2011). Molecular diversity of fungal phylotypes co-amplified alongside nematodes from coastal and deep-sea marine environments, *PloS one* **6**, e26445.
- [10] M. Nukina and S. Marumo (1977). Lunatoic acid A and B, aversion factor and its related metabolite of *Cochliobolus lunata*, *Tetrahedron Lett.* **18**, 2603-2606.
- [11] Y. M. Lee, H. Li, J. Hong, and J. H. Jung (2010). Bioactive metabolites from the sponge-derived fungus *Aspergillus versicolor*, *Arch. Pharm. Res.* **33**, 231-235.
- [12] I. Kock, K. Krohn, H. Egold, S. Draeger, B. Schulz and J.Rheinheimer (2007). New massarilactones, Massarigenin E, and coniothyrenol, isolated from the endophytic fungus *Coniothyrium* sp. from *Carpobrotus edulis*, *Eur. J. Org. Chem.* **2007**, 2186-2190.
- [13] A. Fukami, Y. Taniguchi, T. Nakamura, M. C. Rho, K. Kawaguchi, M. Hayashi and S. Omura (1999). New Members of the Macrocephelides from *Microsphaeropsis* sp. FO-5050 IV, *J. Antibiot.* **52**, 501-504.
- [14] Krohn, K.; Zia, U.; Hussain, H.; Florke, U.; Schulz, B.; S. Draeger and Kurtan, T (2007). Massarilactones E-G, new metabolites from the endophytic fungus *Coniothyrium* sp., associated with the plant *Artimisia maritime*, *Chirality* **19**, 464-470.

- [15] Y. Lin, X. Wu, Z. Deng, J. Wang, S. Zhou, L. Vrijmoed and E. Jones (2002). The metabolites of the mangrove fungus *Verruculina enalia* No. 2606 from a salt lake in the Bahamas, *Phytochemistry* **59**, 469-471.
- [16] H. Jayasuriya, R. G. Ball, D. L. Zink, J. L. Smith, M. A. Goetz, R. G. Jenkins, M. Nallin-Omstead, K. C. Silverman and G. F. Bills (1995). Barceloneic acid A, a new farnesyl-protein transferase inhibitor from a *Phoma* species, *J. Nat. Prod.* **58**, 986-991.
- [17] J.H. Pan, J.J. Deng, Y.G. Chen, J.P. Gao, Y.C. Lin, Z.G. She and Gu, Y.C. (2010). New lactone and xanthone derivatives produced by a mangrove endophytic fungus *Phoma* sp. SK3RW1M from the South China Sea, *Helv. Chem. Acta.* **93**, 1369-1374.
- [18] Y. L. Zhang, L. C. Kong, D. H. Jiang, C. P. Yin, Q. M. Cai, Q. Chen, and J. Y. Zheng (2011). Phytotoxic and antifungal metabolites from *Curvularia* sp. FH01 isolated from the gut of *Atractomorpha sinensis*, *Bioresour. Technol.* **102**, 3575-3577.
- [19] M.M. Wagenaar and J. Clardy (2001). Dicerandrols, New Antibiotic and Cytotoxic Dimers produced by the fungus *Phomopsis longicolla* isolated from an endangered Mint, *J. Nat. Prod.* **64**, 1006-1009.
- [20] M.Y. Yoon, Y.S. Kim, , S.Y. Ryu, G.J. Choi, Y.H. Choi, K.S. Jang, and J.C. Kim (2011). *In vitro* and *in vivo* antifungal activities of decursin and decursinol angelate isolated from *Angelica gigas* against *Magnaporthe oryzae*, the causal agent of rice blast, *Pestic. Biochem. and Phys.* **101**, 118-124.
- [21] H. Hussain, K. Krohn, S. Draeger, K. Meier and B. Schulz (2009). Bioactive chemical constituents of a sterile endophytic fungus from *Melilotus dentatus*, *Rec. Nat. Prod.* **3**, 114-117.
- [22] S. Marumo, M. Nukina, S. Kondo, and K. Tomiyama (1982). Lunatoic acid A, a morphogenic substance inducing chlamydospore-like cells in some fungi, *Agric. Biol Chem.* **46**, 2399-2401.

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