Supporting Information

Rec. Nat. Prod. 17:2 (2023) 329-334

Chemical Constituents of Tectus maximus Koch, 1844

Nguyen Trong Dan¹, Le Thi Giang², Cu Nguyen Dinh¹, Truong Ba Hai¹, Nguyen Dang Hoi¹, Vu Thi Loan,¹ Dan Thi Thuy Hang,³ Nguyen Xuan Nhiem,^{3,4} Bui Huu Tai,^{3,4} and Phan Van Kiem^{3,4*}

¹Vietnam - Russia Tropical Center, 63 Nguyen Van Huyen, Cau Giay, Hanoi, Vietnam

²Thai Nguyen University of Medicine and Pharmacy, 284 Luong Ngoc Quyen Street, Thai Nguyen City, Vietnam

³Institute of Marine Biochemistry, Vietnam Academy of Science and Technology (VAST), 18 Hoang Quoc Viet, Cau Giay, Hanoi, Vietnam

⁴Graduate University of Science and Technology, VAST, 18 Hoang Quoc Viet, Cau Giay, Hanoi, Vietnam

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Figure S2: ¹H NMR spectrum of compound 1 in DMSO-*d*₆



Figure S4: ¹³C NMR spectrum of compound 1 in DMSO-*d*₆



Figure S5: HSQC spectrum of compound 1 in DMSO-*d*₆





Figure S7: ¹H-¹H COSY spectrum of compound **1** in DMSO-*d*₆



Figure S8: NOESY spectrum of compound 1 in DMSO-*d*₆



	Formula (M)	Score (MFG ∇	Mass	Mass (MFG)	m/z (Calc)	Diff (ppm)	DBE	m/z
►	C12 H14 N4 O6	99.43	310.0909	310.0913	311.0986	1.32	8	311.0982
	C11 H18 O10	97.16	310.0909	310.09	311.0973	-2.99	3	311.0982
	C13 H10 N8 O2	90.64	310.0909	310.0927	311.0999	5.64	13	311.0982

Figure S9: HRESIMS spectrum of compound 2



Figure S10: ¹H NMR spectrum of compound 2 in DMSO-*d*₆



Figure S11: Extended ¹H NMR spectrum of compound 2 in DMSO-*d*₆



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Figure S12: ¹³C NMR spectrum of compound 2 in DMSO-*d*₆

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Figure S13: HSQC spectrum of compound 2 in DMSO-*d*₆



Figure S14: HMBC spectrum of compound 2 in DMSO-d₆

Figure S15: ¹H-¹H COSY spectrum of compound 2 in DMSO-*d*₆



Figure S16: NOESY spectrum of compound 2 in DMSO-*d*₆



Figure S17: ¹H NMR spectrum of compound 3 in DMSO-*d*₆

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Figure S18: ¹³C NMR spectrum of compound **3** in DMSO- d_6



Figure S19: ¹H NMR spectrum of compound 4 in DMSO-*d*₆



Figure S20: ¹³C NMR spectrum of compound **4** in DMSO- d_6



Figure S21: HRESIMS spectrum of compound 5



Figure S22: ¹H NMR spectrum of compound **5** in DMSO- d_6



Figure S23: ¹³C NMR spectrum of compound **5** in DMSO- d_6



Figure S24: HRESIMS spectrum of compound 6



Figure S25: ¹H NMR spectrum of compound **6** in DMSO- d_6



Figure S26: Extend ¹H NMR spectrum of compound **6** in DMSO- d_6



Figure S27: ¹³C NMR spectrum of compound **6** in DMSO- d_6



Figure S29: ¹H NMR spectrum of compound **7** in DMSO- d_6



Figure S30: Extended ¹H NMR spectrum of compound 7 in DMSO- d_6



Figure S31: ¹³C NMR spectrum of compound **7** in DMSO- d_6



Figure S32: HRESIMS spectrum of compound 8



Figure S33: ¹H NMR spectrum of compound 8 in DMSO-*d*₆



Figure S34: Extended ¹H NMR spectrum of compound **8** in DMSO- d_6



Figure S35: ¹³C NMR spectrum of compound **8** in DMSO- d_6



Figure S37: ¹H NMR spectrum of compound 9 in CDCl₃



Figure S38: Extend ¹H NMR spectrum of compound 9 in CDCl₃



Figure S39: ¹H NMR spectrum of compound 9 in CDCl₃



Figure S40: ¹H NMR spectrum of compound 9 in CDCl₃



Figure S41: ¹H NMR spectrum of compound 9 in CDCl₃



Figure S42: Extended ¹H NMR spectrum of compound 9 in CDCl₃



Figure S43: ¹H-¹H COSY spectrum of compound 9 in CDCl₃



Figure S44: ¹H NMRspectrum of compound 10 in CDCl₃



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Deoxyinosine (**3**): A dark yellow solid. ¹H NMR (DMSO- d_6 , 600 MHz) δ (ppm): 8.04 (1H, s, H-2), 8.28 (1H, s, H-8), 6.30 (1H, dd, J = 6.6, 6.0 Hz, H-1'), 2.29 (1H, ddd, J = 10.4, 6.0, 2.4 Hz, H_a-2'), 2.62 (1H, ddd, J = 10.4, 8.4, 5.4 Hz, H_b-2'), 4.38 (1H, ddd, J = 5.4, 2.4, 2.4 Hz, H-3'), 3.86 (1H, m, H-4'), 3.58 (1H, dd, 12.0, 4.2 Hz, H_a-5'), 3.51 (1H, dd, 12.0, 3.6 Hz, H_b-5'). ¹³C NMR (DMSO- d_6 , 150 MHz) δ (ppm) data shown in Table S1.

Inosine (4): A dark yellow solid. ¹H NMR (DMSO- d_6 , 600 MHz) δ (ppm), 8.06 (1H, s, H-2), 8.33 (1H, s, H-8), 5.87 (1H, d, J = 6.0 Hz, H-1'), 4.48 (1H, dd, J = 6.0, 4.8 Hz, H-2'), 4.13 (1H, dd, J = 4.8, 3.6 Hz, H-3'), 3.94 (1H, m, H-4'), 3.65 (1H, dd, 12.0, 3.6 Hz, H_a-5'), 3.54 (1H, dd, 12.0, 3.6 Hz, H_b-5'). ¹³C NMR (DMSO- d_6 , 150 MHz) δ (ppm) data shown in Table S1.

Adenosine (5): A dark yellow solid. HRESIMS m/z 252.1089 [M+H]⁺ (calcd. for [C₁₀H₁₃N₅O₃]⁻: 252.1091, $\Delta = -0.8$ ppm); m/z 274.0901 [M+Na]⁺ (calcd. for [C₁₀H₁₂N₅O₃Na]⁻: 274.0911, $\Delta = -3.7$ ppm); ¹H NMR (DMSO- d_6 , 600 MHz) δ (ppm): 8.12 (1H, s, H-2), 8.33 (1H, s, H-8), 6.35 (2H, s, NH₂), 6.34 (1H, dd, J =6.6, 6.0 Hz, H-1'), 2.25 (1H, ddd, J = 13.2, 6.0, 2.4 Hz, H_a-2'), 2.72 (1H, ddd, J = 13.2, 8.0, 6.0 Hz, H_b-2'), 4.41 (1H, ddd, J = 8.0, 6.0, 6.0 Hz, H-3'), 3.88 (1H, m, H-4'), 3.62 (1H, dd, 12.0, 6.5 Hz, H_a-5'), 3.52 (1H, dd, 12.0, 5.0 Hz, H_b-5'). ¹³C NMR (DMSO- d_6 , 150 MHz) δ (ppm) data shown in Table S1.

Deoxyadenosine (**6**):A dark yellow solid. HRESIMS m/z 268.1040 [M+H]⁺ (calcd. for [C₁₀H₁₃N₅O₄]⁻: 268.1040, $\Delta = 0.0$ ppm); m/z 290.0855 [M+Na]⁺ (calcd. for [C₁₀H₁₂N₅O₄Na]⁻: 290.0860, $\Delta = -1.9$ ppm); ¹H NMR (DMSO- d_6 , 600 MHz) δ (ppm): 8.13 (1H, s, H-2), 8.34 (1H, s, H-8), 7.33 (2H, s, NH₂), 5.87 (1H, dd, J = 6.6, 6.0 Hz, H-1'), 4.61 (1H, ddd, J = 6.6, 6.0, 5.4 Hz, H-2'), 4.14 (1H, ddd, J = 6.0, 6.0, 3.0 Hz, H-3'), 3.96 (1H, m, H-4'), 3.67 (1H, ddd, 12.0, 6.0, 4.2 Hz, H_a-5'), 3.55 (1H, ddd, 12.0, 6.0, 3.6 Hz, H_b-5'), 5.42 (1H, d, J = 6.0 Hz, 5'-O<u>H</u>). ¹³C NMR (DMSO- d_6 , 150 MHz) δ (ppm) data data shown in Table S1.

Deoxyuridine (7):A dark yellow solid. HRESIMS m/z 229.0818 [M+H]⁺ (calcd. for [C₉H₁₃N₂O₅]⁺: 229.0819, $\Delta = -0.4$ ppm); m/z 251.0633 [M+Na]⁺ (calcd. for [C₉H₁₂N₂O₅Na]⁺: 251.0638, $\Delta = -2.0$ ppm); ¹H NMR (DMSO- d_6 , 600 MHz) δ (ppm): 5.61 (1H, d, J = 7.8 Hz, H-5), 7.82 (1H, d, J = 7.8 Hz, H-4), 6.15 (1H, dd, J = 6.6, 6.0 Hz, H-1'), 2.07 (2H, m, H-2'), 4.22 (1H, m, H-3'), 3.76 (1H, m, H-4'), 3.52 (1H, dd, 12.0, 3.6 Hz, H_a-5'), 3.56 (1H, dd, 12.0, 3.6 Hz, H_b-5'), ¹³C NMR (DMSO- d_6 , 150 MHz) δ (ppm) data shown in Table S1.

Thymidine (8): A dark yellow solid. HRESIMS m/z 243.0976 [M+H]⁺ (calcd. for [C₁₀H₁₆N₂O₅]⁺: 243.0975, $\Delta = +0.3$ ppm); m/z 265.0796 [M+Na]⁺ (calcd. for [C C₁₀H₁₅N₂O₅Na]⁺: 265.0795, $\Delta = +0.3$ ppm); ¹H NMR © 2022 ACG Publications. All rights reserved. (DMSO- d_6 , 600 MHz) δ (ppm): 7.68 (1H, s, H-4), 1.76 (3H, s, 5-CH₃), 6.16 (1H, dd, J = 6.6, 6.0 Hz, H-1'), 2.06 (2H, m, H-2'), 4.23 (1H, m, H-3'), 3.53 (1H, dd, J = 12.0, 3.6 Hz, H-5'_a), 3.56 (1H, dd, J = 12.0, 3.6 Hz, H-5'_b). ¹³C NMR (DMSO- d_6 , 150 MHz) δ (ppm) data shown in Table S1.

Glycerol arachidonate (**9**):Colorless solid; HRESIMS m/z 396.3108 $[M+NH_4]^+$, calcd for C₂₃H₄₂NO₄: 396.3108, Δ =0; *m/z* 401.2670 [M+Na]+, calcd. for C₂₃H₃₈O₄Na: 401.2662, Δ =+2.0 ppm. ¹H NMR (CDCl₃, 600 MHz) δ (ppm), ¹³C NMR (CDCl₃, 150 MHz) δ (ppm) data shown in Table S2.

Arachidonic acid (10):Colorless solid. ¹H NMR (CDCl₃, 600 MHz) δ (ppm), ¹³C NMR (CDCl₃, 150 MHz) δ (ppm) data shown in Table S2.

Dog	3		4		5		6		7		8	
POS.	$\delta_{C}{}^{a}$	δ_C	$\delta_{C}{}^{b}$	δ_C	$\delta_{C}{}^{c}$	δ_C	${\delta_C}^d$	δ_C	${\delta_C}^e$	δ_C	δ_{C}^{f}	δ_C
2	145.8	145.9	145.8	145.9	152.4	152.4	152.4	152.4	151.96	150.9	150.6	150.6
4	147.8	147.8	148.2	148.2	148.9	148.9	149.0	149.1	141.43	140.4	136.2	136.1
5	124.4	124.4	124.5	124.4	119.3	119.3	119.3	119.4	102.68	101.8	109.4	109.3
6	156.6	156.7	156.6	156.6	156.1	156.1	156.2	156.2	163.57	163.8	163.8	163.9
8	138.5	138.4	138.7	138.7	139.5	139.5	139.9	139.9				
1′	83.5	83.6	87.6	87.5	83.9	83.9	87.9	87.9	86.03	84.1	83.8	83.7
2'	39.5	39.4	74.1	74.1	39.4	39.4	73.4	73.4	39.34	39.1	39.5	39.4
3'	70.6	70.6	70.3	70.3	71.0	71.0	70.6	70.7	71.05	70.4	70.5	70.4
4′	87.9	87.9	85.6	85.6	88.0	88.0	85.9	85.9	87.26	87.4	87.3	87.2
5'	61.6	61.6	61.3	61.3	61.9	61.9	61.6	61.7	61.81	61.3	61.5	61.3

Table S1: ¹³C NMR data for compounds 1-8 in DMSO-d₆ and reference compounds

a: δ_C of deoxyinosine in DMSO-d₆[1]

b: δ_C of inosine in DMSO-d₆[2] c: δ_C of deoxyadenosine in DMSO-d₆[3] d: δ_C of adenosine in DMSO-d₆[3]

e: δ_C of deoxyuridine in CDCl₃[4]

f: δ_C of thymidine in DMSO-d₆[5]

Pos.	9 (glyc	cerol arachidonate)	10 (arachidonic acid)				
	$\delta_{\rm C}$	$\delta_{\rm H}$ (mult, J in Hz)	Pos.	$\delta_{ m C}{}^{ m a}$	$\delta_{ m C}$	$\delta_{\rm H}$ (mult, J in Hz)	
1'	174.1, C	-	1	180.23	nd	_	
2'	33.5, CH ₂	2.37 (t, 7.2)	2	33.43, CH ₂	33.5, CH ₂	2.37 (t, 7.2)	
3'	24.8, CH_2	1.72 (m)	3	24.52, CH ₂	24.6, CH ₂	1.72 (m)	
4'	$26.5, CH_2$	2.11 (q, 7.2)	4	26.50, CH ₂	26.5, CH ₂	2.11 (q, 7.2)	
5'	128.3, CH	5.32 - 5.42	5	129.08, CH	129.1, CH	5.32 - 5.42	
6'	129.1, CH	5.32 - 5.42	6	128.70,CH	128.3,CH	5.32 - 5.42	
7',10',13'	25.6, CH ₂	2.82 (m)	7	25.66, CH ₂	25.6, CH ₂	2.82 (m)	
8', 9', 11',	128.8,	5.32 - 5.42	8, 9,	128.21,	128.8,		
12'	128.7,			128.06, CH	128.6, CH		
	128.1,					5.32 - 5.42	
	127.9, CH						
14'	127.6, CH	5.32 - 5.42	10	25.66	25.6, CH ₂	2.82 (m)	
15'	130.6, CH	5.32 - 5.42	11,	128.52,	128.2,	5.32 - 5.42	
			12	127.85, CH	127.9, CH		
16′	27.2, CH ₂	2.06 (q, 7.2)	13	25.66	25.6, CH ₂	2.82 (m)	
17'	29.3, CH ₂	1.30 (m)	14	127.61, CH	127.6, CH	5.32 - 5.42	
18′	31.5, CH ₂	1.28 (m)	15	130.34, CH	130.5, CH	5.32 - 5.42	
19′	22.6, CH_2	1.30 (m)	16	27.25, CH ₂	27.2, CH ₂	2.06 (q, 7.2)	
20'	14.1, CH ₃	0.89 (t, 7.2)	17	29.39, CH ₂	29.3, CH ₂	1.30 (m)	
1	65.3, CH ₂	4.16 (dd, 12.0, 6.0)	18	31.57, CH ₂	31.5, CH ₂	1.28 (m)	
		4.20 (dd, 12.0, 4.8)					
2	70.3, CH	3.92 (m)	19	22.63, CH ₂	22.6, CH ₂	1.30 (m)	
3	63.4, CH ₂	3.60 (dd, 12.0, 5.4)	20	14.05, CH ₃	14.1, CH ₃	0.89 (t, 7.2)	
		3.70 (dd. 12.0, 2.4)					

Table S2: NMR spectroscopic data for 9 and 10 (in CDCl₃) and reference compounds

The signals without multiples are overlapped, nd: none detected.

Glycerol arachidonate [6]: NMR ¹H (400 MHz, CDCl₃): 0.89 (3H, t, *J* = 6,8 Hz), 1.30 (8H, m), 1.70 (2H, m), 2.09 (4H, m), 2.37 (2H, t, J = 7,8 Hz), 2.81 (6H, m), 3.60 (1H, dd, *J* = 11.7, 5.8 Hz), 3.70 (1H, dd, *J* = 11.7, 3.9 Hz), 3.93 (1H, m), 4.15 (1H, dd, *J* = 11.7, 6.8 Hz), 4.21 (1H, dd, *J* = 11.7, 4.4 Hz), 5.38 (8H, m)

a: δ_C of anachidonic acid in CDCl₃[7]

S2: Cytotoxicity Assay

Human lung carcinoma (SK-LU-1) and human hepatocellular carcinoma (HepG2) cell lines were kindly provided by the Milan University, Italy and Long Island University, USA. The cells were maintained and cultured in DMEM supplemented with FBS, trypsin-EDTA, L-glutamine, sodium piruvat, NaHCO₃, and penicillin/streptomycin at 37°C in a humidified atmosphere of 5% CO2. Cytotoxic effects of compounds were determined using Sulforhodamine B (SRB) assay as previously described by Skehan et al. In brief, the cells were incubated with/without compounds for 72h in 96-well culture plate. After incubation, cells were stained with sulforhodamine B and measured optical density (OD) at 540 nm [8]. Difference of OD between samples and vehicle well during experiments indicated cells situation induced by compounds. Results are expressed as percentage of cells death in comparison with vehicle well. Ellipticine was used as a positive control throughout experiments.

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