

Stereoselective synthesis of C1-C24 fragment of antanapeptin-A

Srinivas Avula*, Sunitha Malladi and Rajesh Kumar Gaddam

Department of Chemistry, Vaagdevi Degree & PG College, Kishanpura, Warangal,
Telangana, India 506001

(Received October 13, 2015; Revised January 5, 2016; Accepted February 1, 2016)

Abstract: The stereoselective synthesis of the (C1-C24) fragment of Antanapeptin-A is described. The required stereochemistry of β -hydroxy- α -methyl acid unit was accomplished through Aldol reaction using Evans' chiral auxiliary followed by the installation of terminal alkyne with Ohira-Bestmann reagent.

Keywords: Antanapeptin A; Aldol Reaction; Stereo selective synthesis; Peptide; Evans' auxiliary. © 2016 ACG Publications. All rights reserved.

1. Introduction

Cyclic depsipeptides¹ have at least one ester linkage formed by replacing one or more amino acid(s) by a hydroxy acid resulting in at least one ester bond in the core ring structure. Some cyclic depsipeptides are cyclized between the C-terminal carboxyl and the side chain of a Thr or Ser residue in the chain. They are secondary metabolites of fungi, plants and some are originated from the marine environment. Cyclo depsipeptides show an interesting spectrum of biological activity such as immunosuppressant, antibiotic, antifungal, anti-inflammatory and anticancer effects.² Since the discovery of the didemnins, this class of natural products continue to stimulate active research in synthetic and medicinal chemistry, as well as in clinical oncology and cell biology.³ Members of this new class of potential drugs may serve as lead compounds in the drug development process for pharmacologically more potent and toxicologically safe derivatives.⁴⁻⁸ Some of these natural products and (semi) synthetic derivatives have already been evaluated in clinical trials. Antanapeptin A (**1**) belongs to a family of natural cyclic depsipeptides, isolated from marine sponges, marine cyanobacterium *Lyngbya majuscula* collected from Antany Mora, Madagascar along with few other natural products.⁹ The structure was elucidated by interpretation of extensive 1D and 2D NMR spectroscopic data. Structurally this compound **1** is cyclic depsipeptide consisting of four α -amino acid residues, proline, *N*-methyl-Phenyl alanine, valine, *N*-methyl-isoleucine and α,β -hydroxy acid unit and one non-amino acid unit, 2-hydroxyisovelaric acid, with different degrees of unsaturation at the terminal end of each molecule. The absolute configurations of **1**, determined by Marfey's analysis, included L-*N*-Me-Ile, L-*N*-Me-Phe, L-Pro, and L-Val. However, the absolute configuration of 2-hydroxyisovelaric acid was not established. Further, the initial biological activity screening of antanapeptins proved that they are inactive in brine shrimp toxicity, sodium channel modulation and antimicrobial bioassays. Due to the paucity of compounds further evaluation remains uncertain. Therefore, the synthesis of antapeptins is necessary for the confirmation of structure along with absolute stereochemistry and further biological screening. Till date, there is no synthesis reported in the literature for this molecule. In continuation of our work on the synthesis of biological active molecules,¹⁰ we have developed a strategy for the synthesis C1-C24 fragment of Antanapeptin A, which is also part of the other molecules, Antanapeptins B, C & D.⁹

* Corresponding author: E-Mail: asvas1978@gmail.com

2. Results and Discussion

From the retrosynthetic outlook (Figure 1), the desired molecule was visualised to be obtained from the key intermediate **2** which in turn could be derived from **3** through oxidation followed by Ohira–Bestmann reaction. Compound **3** was envisioned to be constructed by coupling compound **4** with amine derived from compound **5**. The stereocentres in compound **4** could be obtained by utilizing lithiated aldol-reaction followed by reductive etherification. The stereochemistry in compound **5** was achieved from natural amino acids *L*-Phenyl alanine and *L*-valine.

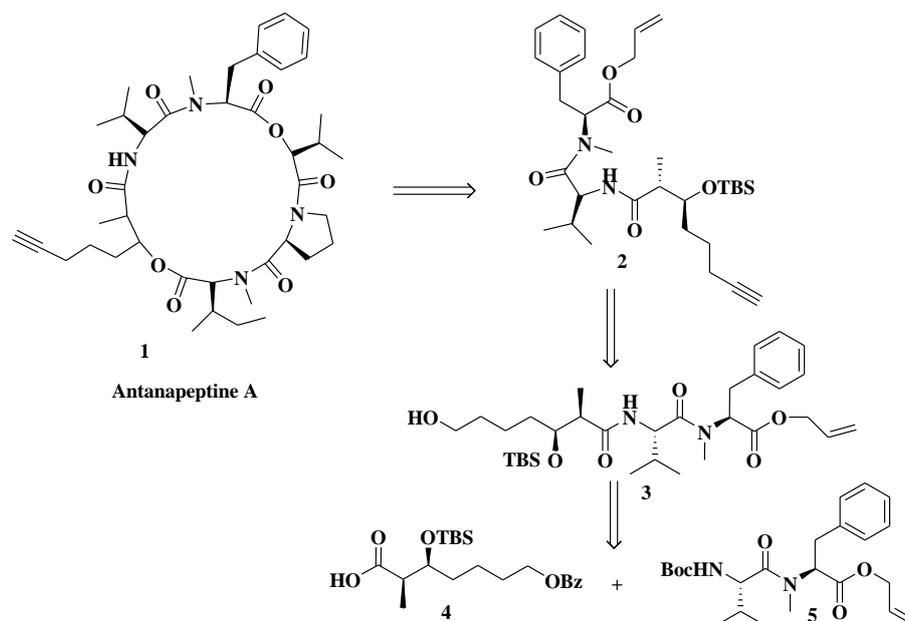


Figure 1. Retrosynthetic analysis of C1 – C24 fragment of Antanapeptin A

Preparation of acid 4: The synthesis began with Evans' aldol reaction of known aldehyde **7** using di-*n*-butylboron enolate of (*R*)-4-benzyl-3-propionyloxazolidin-2-one **6** to furnish the syn-product **8** in 83% yield (Figure 2). Exposure of **8** to TBSCl/Imidazole in CH₂Cl₂ at room temperature for 18 hours gave the TBS-ether **9** in 87% yield. Compound **9** was then treated with sodium borohydride in THF/pH 7 buffer at room temperature for the reductive removal of auxiliary to provide alcohol **10**. The primary hydroxyl group of compound **10** was oxidized to carboxylic acid using BAIB (bis acetoxy iodo benzene)/TEMPO (2,2,6,6-tetramethyl-1-piperidinyloxy free radical) in CH₂Cl₂/pH 7 buffer to obtain the acid fragment **4** in 89% yield.

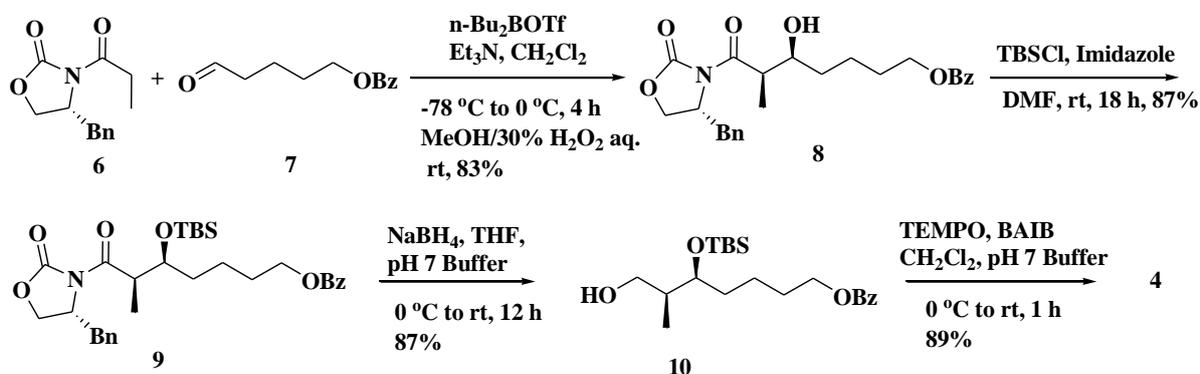


Figure 2. Synthesis of acid fragment **4**

Preparation of compound 5: The synthesis of compound **5** was commenced by carrying out the preparation of the known *N*-(*tert*-butoxycarbonyl)-*N*-methyl-*L*-Phenyl alanine **11** from *L*-Phenyl alanine using the literature procedure.²⁰ Esterification of acid **11** with allyl bromide was cleanly achieved by using K_2CO_3 in DMSO solvent at room temperature, to get the ester **12** in 80% yield. The required amide **5** was prepared from **12** by two transformations. Primarily, treating with TFA in CH_2Cl_2 to get secondary amine in good yields and the crude amine was used as such for the next step without further purification. Later, coupling of the free amine with (*tert*-butoxycarbonyl)-*L*-valine by using HATU/HOAT in CH_2Cl_2 at room temperature for 8 h to acquire the desired fragment **5** in 84% yield (Figure 3).²¹⁻²²

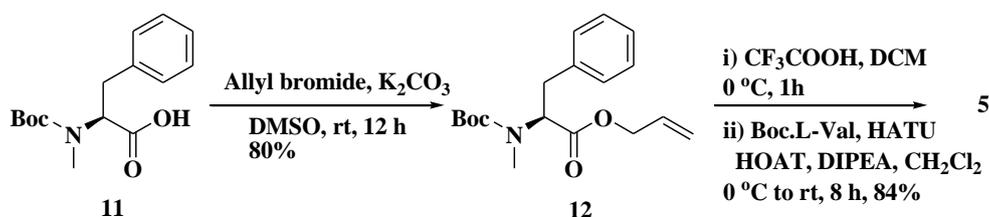


Figure 3. Synthesis of fragment **5**

Construction of C1 – C8 segment from 4 and 5: With successful completion of the desired fragments **4** and **5**, the attention was turned to couple them to give the diamide **13**. For that, initially the Boc protection of compound **5** was deprotected by using TFA in CH_2Cl_2 at 0 °C and then coupled with compound **4** under HATU/HOAT conditions at room temperature, to obtain the required product **13** in 85% yield. The resulting compound **13** was hydrolyzed with potassium carbonate in methanol to give the primary alcohol **3** in 75% yield which upon oxidation with TPAP/NMO to aldehyde followed by the addition of Ohira-Bestmann reagent gave the targeted terminal alkyne product **2** in 92% yield (Figure 4).

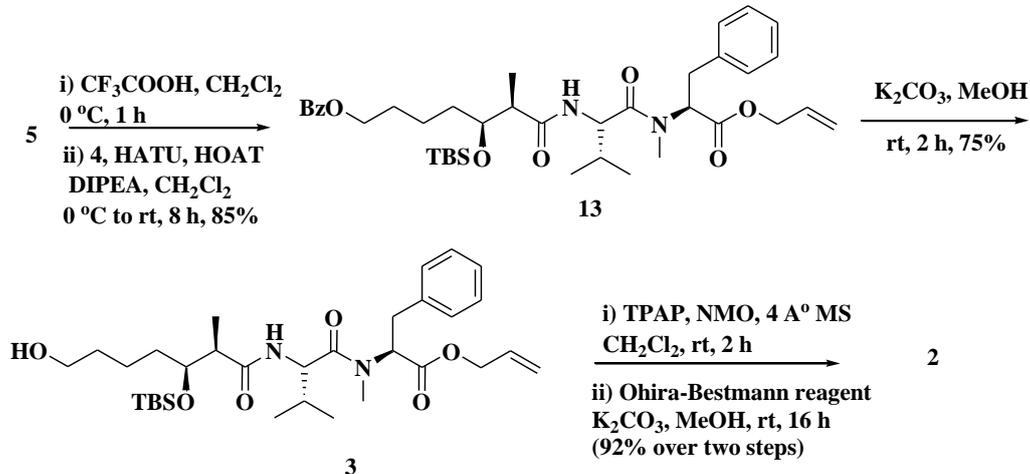


Figure 4. Construction of C1-C8 building block

3. Conclusions

In conclusion, a practical and stereoselective synthesis of C1-C24 fragment of Antanapeptin A having four stereo centers, two amide linkages, one ester linkage and with differential protective groups to allow further extensions was demonstrated, using Evans' Aldol reaction, TEMPO mediated oxidation, Ohira-Bestmann homologation as key transformations. Further investigation towards the total synthesis of Antanapeptin A are in progress.

4. Experimental Section

NMR spectra were recorded in CDCl₃ on brucker AM – 300 (300 MHz) spectrometer at ambient temperature. Chemical shifts are reported in ppm relative to TMS as internal standard and coupling constants are reported in Hz. FTIR spectra were recorded on a Nicolet FT-IR 400 spectrometer in KBr or as neat. Optical rotations were measured on an Perkin – Elmer 141 polarimeter by using a 2 mL cell with a path length of 1 dm with CHCl₃ or CDCl₃ as solvent. Low-resolution mass spectra were obtained on VG 70–70H or LC/MSD trap SL spectrometer operating at 70 eV using direct inlet system. High resolution mass spectra (HRMS) were recorded on an Agilent Technologies 6510 Q-TOF spectrometer. Technical-grade EtOAc and hexanes used for column chromatography were distilled before use. All the reagents and solvents were of reagent grade and used without further purification unless otherwise stated.

(5S,6R)-7-((R)-4-Benzyl-2-oxooxazolidin-3-yl)-5-hydroxy-6-methyl-7-oxoheptylbenzoate (**8**): To a stirred solution of acyl oxazolidinone **6**²⁰ (1.04 g 4.45 mmol) in CH₂Cl₂ (10 mL) was added drop wise *n*-Bu₂BOTf (1.0 M in CH₂Cl₂, 4.67 mL, 4.67 mmol) and stirred for 10 min. *i*-Pr₂NEt (0.93 mL, 5.34 mmol) was then added drop wise and the reaction was stirred at the same temperature for 1h. The mixture was cooled to -78 °C before a solution of aldehyde **7** (1.02 g, 4.95 mmol) in CH₂Cl₂ (15 mL) was added drop wise via cannula. Stirring was continued at -78 °C for 3 h before gradually warming to 0 °C. The reaction mixture was stirred for additional 3 h at 0 °C and then quenched by the addition of 0.1M pH 7 phosphate buffer (7.5 mL) followed by MeOH (10 mL) at 0 °C. After stirring for 5 min, a solution of 30% aqueous H₂O₂ (7.5 mL) in MeOH (15 mL) was added drop wise and stirred at the same temperature for 1h before being concentrated under reduced pressure. The residue was diluted with Et₂O, the phases were separated and the aqueous phase extracted with Et₂O. The combined organic phase was washed with brine, dried over Na₂SO₄, filtered and concentrated under reduced pressure. Flash chromatography over silica gel (35% ethyl acetate in pet. ether) gave **8** (1.63 g, 83%) as a viscous, color less oil. IR (CHCl₃): ν 3250, 2936, 1780, 1710, 1453, 1386, 1277, 1214, 1146, 763, 506 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 8.10 – 7.98 (m, 2H), 7.55 (t, *J* = 7.42 Hz, 1H), 7.43 (t, *J* = 7.5 Hz), 7.39 – 7.24 (m, 3H), 7.20 (d, *J* = 5.6 Hz, 2H), 4.71 (ddt, *J* = 10.4, 6.9, 3.3 Hz, 1H), 4.33 (t, *J* = 6.5 Hz, 2H), 4.28 – 4.13 (m, 2H), 4.04 – 3.93 (m, 1H), 3.77 (qd, *J* = 7.0, 2.7 Hz, 1H), 3.25 (dd, *J* = 13.4, 3.2 Hz, 1H), 2.95 (s, 1H), 2.79 (dd, *J* = 13.4, 9.4 Hz, 1H), 1.87- 1.42 (m, 6H), 1.27 (d, *J* = 7.1 Hz, 3H); ¹³C NMR (75 MHz, CDCl₃): δ 178.0, 166.8, 153.3, 135.2, 133.1, 130.0, 129.0, 128.0, 127.3, 71.6, 66.2, 65.1, 55.1, 42.2, 38.2, 33.0, 28.5, 23.0, 10.4; HRMS (ESI): *m/z* calculated for C₂₅H₃₀O₆N: [M+H]⁺ 440.2067, found 440.2044; [α]_D²⁵ = +29.0 (*c* 1.62, CDCl₃).

(5S,6R)-7-((R)-4-Benzyl-2-oxooxazolidin-3-yl)-5-(tert-butyl dimethylsilyloxy)-6-methyl-7-oxoheptyl benzoate (**9**): To a stirred solution of **8** (1.60 g, 3.64 mmol) in DMF (15 mL) were added imidazole (0.8 g, 12.3 mmol) and TBSCl (*tert*-butylchlorodimethylsilane) (0.95 g, 7.4 mmol). After 18 h at 25 °C, the reaction mixture was added to 20 % CH₂Cl₂: hexane (100 mL) and successively washed with 10% aq. NaHSO₃ (25 mL) and water (2 x 25 mL). The organic layer was dried over Na₂SO₄, filtered, concentrated in vacuo and distilled to yield **9** (1.74 g, 87%) as colorless oil. IR (CHCl₃): ν 3349, 3380, 3088, 2971, 2934, 1699, 1740, 1472, 1452, 1391, 1368, 1311, 1256, 1152, 992, 933, 771, 666, 560 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 8.10 – 7.98 (m, 2H), 7.55 (t, *J* = 7.4 Hz, 1H), 7.43 (t, *J* = 7.7 Hz, 2H), 7.38 – 7.28 (m, 3H), 7.20 (d, *J* = 7.0 Hz, 2H), 4.6 (ddt, *J* = 10.2, 6.4, 3.1 Hz, 1H), 4.33 (t, *J* = 6.5 Hz, 2H), 4.12 – 4.27 (m, 2H), 4.02 (q, *J* = 5.4, 5.6 Hz, 1H), 3.88 (qd, *J* = 6.8, 1.6 Hz, 1H), 3.30 (dd, *J* = 13.2, 3.0 Hz, 1H), 2.78 (dd, *J* = 13.2, 9.6 Hz, 1H), 1.70 (m, 2H), 1.62 – 1.42 (m, 4H), 1.22 (d, *J* = 6.8 Hz, 3H), 0.90 (s, 9H), 0.20 (s, 6H); ¹³C NMR (75 MHz, CDCl₃): δ 175.2, 166.3, 153.0, 135.0, 133.1, 129.3, 129.1, 128.1, 127.6, 72.6, 65.7, 64.7, 56.3, 43.2, 35.0, 29.2, 26.3, 22.0, 19.0, 12.3, - 4.5; HRMS (ESI): *m/z* calculated for C₃₁H₄₇O₆N₂Si: [M+Na]⁺ 571.3197, found 571.3174; [α]_D²⁵ = +10.53 (*c* 1.33, CDCl₃).

(5S,6S)-5-(tert-Butyldimethylsilyloxy)-7-hydroxy-6-methylheptylbenzoate (**10**): To a stirred solution of **9** (1.70 g, 3.07 mmol) in THF (70 mL) at 0 °C was added a solution of NaBH₄ (0.58 g, 15.3 mmol) in pH 7 buffer (18.5 mL). The resulting solution was stirred for 10 min at 0 °C before being allowed to gradually warm to room temperature and continued stirring for overnight. The reaction was quenched

by the addition of sat. aq. NH_4Cl (20 mL) and stirred at room temperature for 1h. The separated aqueous phase was extracted with EtOAc (2 x 25 mL). The combined organic phase was washed with brine, dried over anhydrous Na_2SO_4 , filtered and concentrated under reduced pressure. Flash chromatography over silica gel (15% ethyl acetate in pet. ether) gave **10** (1.01 g, 87%) as a colorless oil. IR (CHCl_3): ν 3436, 2954, 2930, 2857, 1721, 1459, 1275, 1113, 1033, 836, 773, 712 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3): δ 8.02 – 8.10 (m, 2H), 7.55 (t, $J = 7.3$ Hz, 1H), 7.43 (t, $J = 7.9$ Hz, 2H), 4.70 (s, 1H), 4.33 (t, $J = 6.7$ Hz, 2H), 3.79 – 3.75 (m, 1H), 3.70 (t, $J = 9.0$ Hz, 2H), 3.56 – 3.48 (m, 1H), 1.97 (m, 1H), 1.56 – 1.32 (m, 6H), 0.90 (s, 9H), 0.82 (d, $J = 7.8$ Hz, 3H), 0.20 (s, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ 166.7, 132.8, 130.1, 129.3, 128.4, 77.4, 65.5, 64.8, 39.4, 32.0, 28.7, 25.9, 22.7, 17.9, 12.1, -4.6; HRMS (ESI): m/z calculated for $\text{C}_{21}\text{H}_{37}\text{O}_4\text{Si}$: $[\text{M}+\text{H}]^+$ 381.24556, found 381.24414.

(2*R*,3*S*)-7-(Benzoyloxy)-3-(*tert*-butyldimethylsilyloxy)-2-methylheptanoic acid (**4**): To a stirred solution of **10** (1.0 g, 2.62 mmol) in (5 mL) CH_2Cl_2 at 0 °C were added BAIB (3 g, 9.31 mmol), catalytic amount of TEMPO in pH 7 buffer (3 mL). The resulting solution was stirred for 10 min at 0 °C before being allowed to gradually warm to room temperature with stirring for 1h. The reaction was quenched by the addition of sat. aq. NH_4Cl (20 mL) and stirred at room temperature for 1h. The separated aqueous phase was extracted with EtOAc (2 x 25 mL). The combined organic phase was washed with brine, dried over anhydrous Na_2SO_4 filtered and concentrated under reduced pressure. Flash chromatography over silica gel (30% ethyl acetate in pet ether) gave **4** (0.92 g, 89%) as colorless oil. IR (CHCl_3): ν 2931, 2857, 1715, 1459, 1386, 1274, 1220, 1110, 1069, 1026, 936, 836, 773, 711, 675 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3): δ 8.10 – 7.98 (m, 2H), 7.50 (t, $J = 7.4$ Hz, 1H), 7.43 (t, $J = 7.5$ Hz, 2H), 4.33 (t, $J = 6.7$ Hz, 2H), 3.99 (q, $J = 6.0, 5.2$ Hz, 1H), 2.66 – 2.56 (m, 1H), 1.72 – 1.30 (m, 6H), 1.22 (d, $J = 6.8$ Hz, 3H), 0.90 (s, 9H), 0.20 (s, 6H); ^{13}C NMR (75 MHz, CDCl_3): δ 165.6, 134.2, 130.0, 129.9, 128.4, 73.5, 65.3, 44.5, 33.5, 29.2, 26.2, 22.2, 18.4, 11.5, -5.4; HRMS (ESI): m/z calculated for $\text{C}_{21}\text{H}_{34}\text{O}_5\text{Si}$: $[\text{M}+\text{H}]^+$ 393.4556, found 392.4414.

(*S*)-allyl 2-(*tert*-butoxycarbonyl(methyl)amino)-3-phenylpropanoate (**12**): Na_2CO_3 (2.55 g, 24.3 mmol) and Boc_2O (3.94 g, 18.2 mmol) were added to a solution of *L*-Phenylalanine (2.1 g, 12.3 mmol), in H_2O (20 mL) and THF (5 mL) at 0 °C. After the reaction mixture had been stirred at room temperature for 12h, it was neutralized with HCl (10%) until pH 2 has been reached. The mixture was then extracted with EtOAc (50 mL x 3), washed with brine, dried over Na_2SO_4 . Concentration gave the crude *N*-Boc-phenylalanine (3.1 g, 100%). NaH (60 % in mineral oil, 2.45 g, 61.3 mmol) was added in portions to a solution of *N*-Boc-isoleucine (3.1 g, 11.69 mmol) and MeI (6.05 mL) in THF (50 mL) at 0 °C. After the reaction mixture had been stirred at room temperature for 36 h, it was poured in to saturated NH_4Cl solution (250 mL), extracted with EtOAc (3 x 150 mL) and dried over Na_2SO_4 . Concentration gave *N*-methyl-*N*-Boc-phenylalanine (3 g, 92%). K_2CO_3 (3.08 g, 22.3 mmol) and allyl bromide (1.4 mL, 16.9 mmol) were added to a solution of *N*-methyl-*N*-Boc-isoleucine (3 g, 10.75 mmol) in DMSO (40 mL). After the mixture had been stirred at room temperature for 12h, it was portioned between EtOAc (75 mL) and brine (75 mL). The organic phase was separated and aqueous phase was extracted with EtOAc (2 x 100 mL). The combined organic phase was dried over Na_2SO_4 and concentrated. Flash chromatography gave **12** (3.2 g, 93.2%). IR (CHCl_3): ν 3065, 2936, 2880, 1741, 1701, 1602, 1650, 1480, 1456, 1439, 1393, 1367, 1313, 1255, 1183, 1145, 1047, 990, 931, 871, 773 cm^{-1} ; ^1H NMR (300 MHz, CDCl_3): δ 7.37 – 7.34 (m, 3H), 7.23 (d, $J = 8.2$ Hz, 1H), 7.21 (d, $J = 8.3$ Hz, 1H), 5.97 – 5.84 (m, 1H), 5.34 – 5.20 (m, 1H), 4.62 – 4.60 (m, 2H), 4.55 (d, $J = 11.1$ Hz, 1H), 4.36 (t, $J = 11.9$ Hz, 1H), 3.29-3.13 (m, 2H), 2.81 (s, 3H), 2.78* (s, 3H), 1.45 (m, 9H); ^{13}C NMR (75 MHz, CDCl_3): δ 172.3, 156.4, 135.2, 132.1, 127.6, 125.8, 118.4, 80.3, 65.6, 59.6, 34.5, 30.8, 28.7; HRMS (ESI): m/z calculated for $\text{C}_{18}\text{H}_{26}\text{O}_4\text{N}$: $[\text{M}+\text{H}]^+$ 319.1784, found 319.1566. $[\alpha]_{\text{D}}^{25} = -34.7$ (c 1.1, CHCl_3). * denotes the rotamer peak.

(*S*)-allyl 2-((*S*)-2-(*tert*-butoxycarbonylamino)-*N*,3-dimethylbutanamido)-3-phenylpropanoate (**5**): A solution of (*tert*-butoxycarbonyl)-*L*-valine acid (0.35 g, 1.22 mmol), HATU (0.24 g, 1.8 mmol) and HOAT (0.34 g, 1.8 mmol) in CH_2Cl_2 (5 mL) was stirred at 0 °C under N_2 atmosphere for 15 min, treated sequentially with salt [prepared from **12** (0.395 g, 1.24 mmol) in dry CH_2Cl_2 (1 mL) at 0 °C on treatment with CF_3COOH (0.1 mL)] and DIPEA (0.6 mL, 3.6 mmol) and stirred for 8 h. The reaction mixture was quenched with aq. satd. NH_4Cl solution (10 mL). After 10 min, it was diluted with CHCl_3

(2 x 10 mL) and washed with water (10 mL), NaHCO₃ solution (10 mL) and brine (10 mL). The organic layers were dried over Na₂SO₄, evaporated and the residue was purified by column chromatography (60-120 mesh Silica gel, 35% ethyl acetate in pet. ether) to afford **5** (0.46 g, 89%) as a colorless syrup. IR (CHCl₃): ν 3333, 3060, 2945, 2862, 1742, 1706, 1658, 1602, 1462, 1439, 1367, 1294, 1178, 1001, 938, 876, 774 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 7.37–7.34 (m, 3H), 7.23 (d, J = 8.2 Hz, 1H), 7.21 (d, J = 8.3 Hz, 1H), 5.40–5.41 (m, 1H), 5.05 (d, J = 2.2 Hz, 2H), 4.40 (q, J = 2.6 Hz, 1H), 3.70 (s, 3H), 3.51–3.55 (m, 2H), 3.09 (s, 2H), 2.10–1.89 (m, 2H), 1.50 (s, 9H), 1.00–0.80 (m, 7H); ¹³C NMR (75 MHz, CDCl₃): δ 173.5, 171.4, 136.4, 132.6, 128.2, 125.4, 127.7, 118.6, 79.8, 60.4, 55.3, 52.2, 31.4, 29.8, 19.8; HRMS (ESI): m/z calculated for C₂₃H₃₄N₂O₅Na: [M+Na]⁺ 441.2562, found 441.2458.

(5*S*,6*R*,9*R*,12*S*)-allyl-5-(4-(benzyloxy)butyl)-12-benzyl-9-isopropyl-2,2,3,3,6,11-hexamethyl-7,10-dioxo-4-oxa-8,11-diaza-3-silatridecan-13-oate (**13**): A solution of acid **5** (0.51 g, 1.29 mmol), HATU (0.24 g, 1.8 mmol) and HOAT (0.34 g, 1.8 mmol) in CH₂Cl₂ (5 mL) was stirred at 0 °C under N₂ atmosphere for 15 min, treated sequentially with TFA salt [prepared from **5** (0.45 g, 1.2 mmol) in dry CH₂Cl₂ (1 mL) at 0 °C on treatment with CF₃COOH (0.1 mL)] and DIPEA (0.6 mL, 3.6 mmol) and stirred for 8 h. The reaction mixture was quenched with aq. satd. NH₄Cl solution (10 mL). After 10 min, it was diluted with CHCl₃ (2 x 10 mL) and washed with water (10 mL), NaHCO₃ solution (10 mL) and brine (10 mL). The organic layers were dried over Na₂SO₄, evaporated and the residue purified by column chromatography (60-120 mesh Silica gel, 45% ethyl acetate in pet. ether) to afford **13** (0.725 g, 85%) as a colorless syrup. IR (CHCl₃): ν 3342, 3333, 3233, 3060, 2926, 2915, 2854, 2840, 1701, 1667, 1651, 1609, 1460, 1448, 1380, 1220, 1172, 1110, 1069, 1026, 1003, 936, 836, 773, 771, 711, 676, 667 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 8.00 (d, J = 7.5 Hz, 2H), 7.55 (d, J = 8.3 Hz, 1H), 7.43 (t, J = 7.5 Hz, 2H), 7.36–7.32 (m, 3H), 7.23 (d, J = 8.2 Hz, 1H), 7.21 (d, J = 8.3 Hz, 1H), 5.96–5.79 (m, 1H), 5.39–5.18 (m, 2H), 5.11–5.01 (m, 1H), 4.69–4.66 (m, 2H), 4.65–4.54 (m, 2H), 4.34–4.27 (m, 3H), 3.82–3.68 (m, 3H), 3.10–3.05 (m, 2H), 2.06–1.89 (m, 1H), 1.36–1.19 (m, 4H), 1.20–1.05 (m, 1H), 1.01–0.78 (m, 19H), 0.10–0.03 (m, 6H); ¹³C NMR (75 MHz, CDCl₃): δ 175.1, 171.2, 165.9, 136.5, 133.5, 132.5, 130.0, 129.4, 128.6, 125.9, 118.2, 73.5, 65.9, 56.7, 54.1, 49.2, 32.1, 30.1, 29.4, 26.2, 19.2, 16.4, 14.4, -5.4. HRMS (ESI): m/z calculated for C₃₉H₅₈N₂O₇SiNa: [M+Na]⁺ 717.4556, found 717.4214.

(5*S*,6*R*,9*R*,12*S*)-allyl-12-benzyl-5-(4-hydroxybutyl)-9-isopropyl-2,2,3,3,6,11-hexamethyl-7,10-dioxo-4-oxa-8,11-diaza-3-silatridecan-13-oate(**3**): To a stirred solution of **13** (0.70 g, 1.06 mmol) in MeOH (5 mL) was added K₂CO₃ (0.045 g, 0.33 mmol). The reaction was stirred at room temperature until complete by TLC (2h). The mixture was then diluted with CH₂Cl₂ (25 mL) and washed with H₂O (5mL). The separated aqueous phase was extracted with CH₂Cl₂ (3 x 10 mL) and the combined organic phase dried over Na₂SO₄, filtered, evaporated and the residue was purified by column chromatography (60-120 mesh Silica gel, 52% ethyl acetate in pet ether) to afford **3** (0.44 g, 75%) as a colorless syrup. IR (CHCl₃): ν 3640, 3300, 3233, 3100, 2956, 2926, 2925, 2854, 2840, 1701, 1667, 1651, 1460, 1448, 1380, 1220, 1172, 1110, 1069, 1026, 1003, 936, 836, 773, 771, 711, 676, 667 cm⁻¹; ¹H NMR (300 MHz, CDCl₃): δ 7.34–7.32 (m, 3H), 7.22 (d, J = 8.2 Hz, 1H), 7.20 (d, J = 8.3 Hz, 1H), 5.94–5.81 (m, 1H), 5.36–5.28 (m, 2H), 5.11–5.01 (m, 1H), 4.65–4.62 (m, 2H), 4.60–4.54 (m, 2H), 4.45 (brs, 1H), 4.31–4.26 (m, 3H), 3.82–3.68 (m, 3H), 3.09–3.05 (m, 2H), 2.05–1.89 (m, 1H), 1.32–1.19 (m, 4H), 1.19–1.07 (m, 1H), 1.05–0.83 (m, 19H), 0.10–0.03 (m, 6H); ¹³C NMR (75 MHz, CDCl₃): δ 175.4, 174.3, 171.2, 136.6, 132.5, 128.6, 118.2, 74.5, 66.0, 65.3, 62.8, 58.2, 54.1, 49.2, 38.0, 34.2, 32.1, 26.2, 25.1, 19.2, 16.4, 14.4, -5.4; HRMS (ESI): m/z calculated for C₃₂H₅₄N₂O₆SiNa: [M+Na]⁺ 613.3914, found 613.3812.

(5*S*,6*R*,9*R*,12*S*)-allyl-12-benzyl-9-isopropyl-2,2,3,3,6,11-hexamethyl-7,10-dioxo-5-(pent-4-ynyl)-4-oxa-8,11-diaza-3-silatridecan-13-oate(**2**): To a stirred solution of **3** (0.42 g, 0.755 mmol) and powdered 4 Å molecular sieves (0.65 g) in CH₂Cl₂ (15 mL) were subsequently added 4-methylmorpholine-*N*-Oxide (0.23 g, 1.94 mmol) and tetrapropylammonium perruthenate (0.023 g, 0.065 mmol) and the mixture was stirred at room temperature for 2 h. The reaction mixture was filtered through a pad of silica gel and the filtrate was concentrated under reduced pressure to give aldehyde as a colorless oil. The residue was placed under high vacuum for 2 h before being used in the

subsequent reaction without purification. Stirred the aldehyde, K_2CO_3 and the Ohira-Bestmann reagent (0.26 g, 1.32 mmol) for 16 h. The mixture was then diluted with CH_2Cl_2 (25 mL) and washed with H_2O (5 mL). The separated aqueous phase was extracted with CH_2Cl_2 (3 x 10 mL) and the combined organic phase dried over Na_2SO_4 , filtered and evaporated and the residue was purified by column chromatography (60-120 mesh Silica gel, 5% ethyl acetate in pet ether) to afford **2** (0.35 g, 92%) as yellow syrup. 1H NMR (300 MHz, $CDCl_3$): δ 7.36–7.34 (m, 3H), 7.26 (d, $J = 8.2$ Hz, 1H), 7.25 (d, $J = 8.3$ Hz, 1H), 5.94–5.81 (m, 1H), 5.36–5.28 (m, 2H), 5.11–5.01 (m, 1H), 4.65–4.62 (m, 2H), 4.60–4.54 (m, 2H), 4.31–4.26 (m, 3H), 3.82–3.68 (m, 3H), 2.20 (m, 1H), 2.05–1.89 (m, 1H), 1.32–1.19 (m, 4H), 1.19–1.07 (m, 1H), 1.05–0.83 (m, 19H), 0.10–0.03 (m, 6H); ^{13}C NMR (75 MHz, $CDCl_3$): δ 172.1, 132.3, 136.8, 128.5, 127.8, 125.9, 118.5, 83.9, 71.5, 68.2, 65.9, 56.5, 52.1, 48.2, 38.1, 34.7, 32.1, 31.4, 26.2, 18.7, 16.4, -5.4; HRMS (ESI): m/z calculated for $C_{33}H_{52}N_2O_5SiNa$: $[M+Na]^+$ 607.3140, found 607.3120.

5. Acknowledgements

A. Srinivas thankful to CSIR, New Delhi for Research Associate fellowship, Director, Indian Institute of Chemical Technology (IICT), Hyderabad for providing research facilities and Dr. G.V. M. Sharma, Chief Scientist, IICT, Hyderabad for valuable guidance.

6. References

- [1] Lemmens-Gruber, R.; Kamyar, M. R.; Dornetshuber, R. Cyclodepsipeptides - Potential drugs and lead compounds in the drug development process. *Curr. Med. Chem.* **2009**, *16*, 1122-1137.
- [2] Sarabia, F.; Chammaa, S.; Ruiz, A. S.; Ortiz, L. M.; Herrera, F. J. Chemistry and biology of cyclic depsipeptides of medicinal and biological interest. *Curr. Med. Chem.* **2004**, *11*, 1309-1332.
- [3] Vera, M. D.; Joullie, M. M. Natural products as probes of cell biology: 20 years of didemnin research. *Med. Res. Revs.* **2002**, *22*, 102-145.
- [4] Yurek-George, A.; Cecil, A. R.; Mo, A. H.; Wen, S.; Rogers, H.; Habens, F.; Maeda, S.; Yoshida, M.; Packham, G.; Ganesan, A. The first biologically active synthetic analogues of FK228, the depsipeptide histone deacetylase inhibitor. *J. Med. Chem.* **2007**, *50*, 5720-5726.
- [5] Adrio, J.; Cuevas, C.; I. Manzanares, I.; Joullie, M. M. Total synthesis and biological evaluation of tamandarin B analogues. *J. Org. Chem.* **2007**, *72*, 5129-5138.
- [6] Liang, B.; Richard, D. J.; Portonovo, P. S.; Joullie, M. M. Total Syntheses and Biological Investigations of Tamandarins A and B and Tamandarin A Analogs. *J. Am. Chem. Soc.* **2001**, *123*, 4469-4474.
- [7] Pan, P. S.; McGuire, K. L.; McAlpine, S. R. Identification of Sansalvamide a analog potent against pancreatic cancer cell lines. *Bio. Org. Med. Chem. Letts.* **2007**, *17*, 5072-5077.
- [8] Otrubova, K.; Lushington, G.; Velde, D. V.; McGuire, K. L.; McAlpine, S. R. Comprehensive study of sansalvamide A derivatives and their Structure-Activity Relationships against drug-resistant colon cancer cell lines. *J. Med. Chem.* **2008**, *51*, 530-544.
- [9] Nogle, L. M.; Gerwick, W. H. Isolation of Four New Cyclic Depsipeptides, Antanapeptins A-D, and Dolastatin 16 from a Madagascan Collection of *Lyngbya majuscula*. *J. Nat. Prod.* **2002**, *63*, 21-24.
- [10] Srinivas, A.; Nagaraj, A.; Reddy, C. S. Synthesis and *in vitro* study of methylene-bis-tetrahydro[1,3]thiazolo[4,5-c]isoxazoles as potential nematicidal agents. *Eur. J. Med. Chem.* **2010**, *45*, 2353-2355
- [11] Reddy, C. S.; Srinivas, A.; Sunitha, M.; Nagaraj, A. Design and synthesis of Novel methylene-bis-fused pyrazoles as biologically active molecules. *J. Heterocycl. Chem.* **2010**, *47*, 1301-1309.
- [12] Reddy, C. S.; Nagaraj, A.; Srinivas, A.; Reddy, G. P. $ZrCl_4$ catalyzed efficient one-pot synthesis of novel methylene-bis- β -amino/ methylene-bis- β -acetamido ketones. *Indian J. Chem. B.* **2010**, *49B*, 617-622.
- [13] Srinivas, A.; Reddy, C. S.; Nagaraj, A. Synthesis, nematicidal and antimicrobial properties of bis-[4-methoxy-3-[3-(4-fluorophenyl)-6-(4-methylphenyl)-2(aryl)-tetrahydro-2H-pyrazolo[3,4-d]thiazol-5-yl]phenyl]methanes. *Chem. Pharm. Bull.* **2009**, *57*, 685-693.
- [14] Reddy, C. S.; Srinivas, A.; Nagaraj, A.; Synthesis and *in vitro* study of a new class of methylene-bis-4,6-diarylbenzo[d]isoxazoles as potential antifungal agents. *J. Heterocycl. Chem.* **2009**, *46*, 497-502.
- [15] Reddy, C. S.; Nagaraj, A.; Srinivas, A.; Reddy, G. P. $ZrOCl_2 \cdot 8H_2O$ catalyzed Bayer condensation: A facile and efficient synthesis of triarylmethanes under solvent-free condition *Indian J. of Chem B.* **2009**, *48B*, 248-254.
- [16] Reddy, C. S.; Srinivas, A.; Nagaraj, A.; Synthesis and biological evaluation of novel methylene-bis-thiazolidinone derivatives as potential Nematicidal agents. *J. Heterocycl. Chem.* **2008**, *45*, 999-1003.

- [17] Reddy, C. S.; Srinivas, A.; Nagaraj, A. Synthesis of some novel Methylene-bis-pyrimidinyl-spiro-4-thiazolidinones as biologically potent agents. *J.Heterocycl.Chem.* **2008**, *45*, 1121-1125.
- [18] Reddy, C. S.; Reddy, G. P.; Nagaraj, A.; Srinivas, A. Synthesis and Biological study of novel methylene-bis-benzofuranyl-[1,5]-benzothiazepines. *Org. Commun.* **2008**, *1*, 84-94.
- [19] Reddy, C. S.; Srinivas, A.; Nagaraj, A. Synthesis and nematicidal activity of 2-(1*H*-benzo[*d*]imidazole-2-ylmethyl)-4-aryl-1-thia-4-azaspiro [4,5]decan-3-one. *Indian J. of Chem B.* **2008**, *47B*, 787-791.
- [20] Gage, J. R.; Evans, D. A. Diastereoselective aldol condensation using a chiral oxazolidinone auxiliary: (2*s**,3*s**)-3-hydroxy-3-phenyl-2-methylpropanoic acid. *Org. Syn.* **1990**, *68*, 83- 91.
- [21] Ilankumaran, P.; Verkade, J. G. P(RNCH₂CH₂)₃N: Efficient Catalysts for Transesterifications, Acylations, and Deacylations. *J. Org. Chem.* **1999**, *64*, 3086.
- [22] Waldmann, H.; Kunz, H. Allylester als selektiv abspaltbare Carboxyschutzgruppen in der Peptid- und N-Glycopeptidsynthese. *Lie. Ann. Chem.* **1983**, 1712-1725.

ACG
publications

© 2016 ACG Publications