

Rec. Nat. Prod. 14:3 (2020) 190-195

records of natural products

Diterpenoids and Sesquiterpenoids from Syzygium fluviatile

Dingli Zhang¹, Yikao Hu¹, Li Wang², Shengxiong Huang², Yan Zhao¹, Mengjia Li¹, Feng Li¹ and Yong Zhao¹,*

¹ College of Chemistry and Chemical Engineering, Yunnan Normal University, Kunming 650500,

China

² State Key Laboratory of Phytochemistry and Plant Resources in West China, Kunming Institute of Botany, Chinese Academy of Sciences, Kunming 650201, China

(Received August 30, 2019; Revised October 26, 2019; October 27, 2019)

Abstract: A new monocyclic diterpenoid, cassipouryl formate, (1) together with three known diterpenoids (2-4) and five known sesquiterpenoids (5-9) were isolated from the twigs and leaves of *Syzygium fluviatile*. The new isolate was elucidated by various spectroscopic technologies. This was the first example of the chemical constituents from this species and the first report on the diterpenoids from genus *Syzygium*.

Keywords: *Syzygium fluviatile;* Myrtaceae; diterpenoids; sesquiterpenoids.. © 2020 ACG Publications. All rights reserved.

1. Introduction

The genus *Syzygium* is one of the largest groups of the family Myrtaceae with more than 500 species distributed in the tropic and subtropical regions of the world [1]. Many of them have been used as an important crude drug for the treatment of diabetes in Southeast Asian nations [2]. Previous phytochemical research on *Syzygium* demonstrated that flavonoids, hydrolysable tannins, phenylpropanoids, chromone derivatives, phloroglucinol derivatives, and triterpenes were their major constituents [1, 3-7]. *Syzygium fluviatile* (Hemsl.) Merr. et Perry, is a shrub which distributed in Guangdong and Guangxi Province in China [8]. As far as we know, there has not report on the chemical constituents of this plant to date. In our interest in the discovery of bioactive metabolites from *Syzygium* plants, one new monocyclic diterpenoid, (1) together with three known diterpenoids (2–4) and five known sesquiterpenoids (5–9) (Figure 1) were isolated from the twigs and leaves of this species.

The article was published by ACG Publications

http://www.acgpubs.org/journal/records-of-natural-products May-June 2020 EISSN:1307-6167 DOI: http://doi.org/10.25135/rnp.158.19.08.1393

^{*} Corresponding author: <u>zhaooy@126.com;</u> Phone: 86-871-65941087 Fax: 86-871-65941088

2. Materials and Methods

2.1. Plant Material

The twigs and leaves of *Syzygium fluviatile* (Hemsl.) Merr. et Perry were collected in November 2016 from Xishuang Banna Tropical Botanical Garden, P. R. China, and were authenticated by Prof. Yu Chen at Kunming Institute of Botany, Chinese Academy of Science. Plant specimens (CEYNNU 20161102) were deposited at College of Chemistry and Chemical Engineering, Yunnan Normal University.

2.2. Extraction and Isolation

The air-dried and powdered twigs and leaves of *S. fluviatile* (5.0 kg) were extracted with 70% acetone (8 L × 3) at room temperature, then evaporated under reduced pressure to provide a residue. the residue was divided with petroleum ether (4 × 1 L), EtOAc (4 × 1 L), and n-BuOH (4 × 1 L), sequentially into three parts. The EtOAc extract (120 g) was subjected to silica gel column chromatography (CC) using a gradient system of petroleum ether/acetone (v/v, 50:0 to 0:1) to afford ten fractions (Fr A–J).

Fraction A (0.6 g) was further chromatographed over silica gel CC using a gradient elution of petroleum ether/ethyl acetate (v/v, 20:1–5:1) to afford three subfractions Fr A1–A4, then fraction A2 (70 mg) was isolated by Sephadex LH-20 column (CHCl₃/MeOH, 1:1), followed by semipreparative HPLC (MeOH/H₂O 95:5) to give compounds 4 (1.5 mg, $t_{\rm R} = 16$ min) and 7 (2.5 mg, $t_{\rm R} = 22$ min). The Sephadex LH-20 column (CHCl₃/MeOH, 3:2) chromatography for fraction D (2.1 g), followed by silica gel CC with petroleum ether/ethyl acetate (v/v, 20:1) give four subfractions Fr D1-D4. The purification of fraction D3 (0.5 g) with Sephadex LH-20 column (CHCl₃/MeOH, 1:1) and semipreparative HPLC (MeOH/H₂O 93:7) to give compounds 1 (1.0 mg, $t_{\rm R}$ = 15 min) and 2 (1.5 mg, $t_{\rm R} = 12$ min). Fraction E (4.0 g) was decolorized by MCI gel (MeOH/H₂O 95:5), then treated with silica gel CC using petroleum ether/ethyl acetate (v/v, 5:1) to provide three subfractions Fr E1–E3. Fraction E2 (0.7 g) afforded compounds 6 (1.6 mg, $t_R = 11 \text{ min}$), 8 (1.5 mg, $t_R = 15 \text{ min}$), and 9 (1.2 mg, $t_R = 17$ min) by Sephadex LH-20 chromatography (CHCl₃/MeOH, 3:2) and semipreparative HPLC (MeOH/H₂O 92:8). Fraction I (2.9 g) was decolorized in the same way, followed by chromatograhy over silica gel CC with petroleum ether/ethyl acetate (v/v, 10:1) to yield five subfractions Fr I1–I5. Sephadex LH-20 column (CHCl₃/MeOH, 3:2) separation of fraction I4 (0.3 g) along with the semipreparative HPLC (MeOH/H₂O 90:10) led to compounds 3 (1.5 mg, $t_R = 20$ min) and 5 (1.0 mg, $t_{\rm R} = 15$ min).

3. Results and Discussion

Compound **1**, a colorless gum (CH₃OH), of which molecular formula was determined as C₂₁H₃₈O₂ by positive HR-EI-MS (m/z 322.2878 [M]⁺, calc. 322.2872), requiring 3 degrees of unsaturation. Strong UV absorption at 197 nm implied it had isolated chromophores. IR spectrum disclosed the existence of ester carbonyl (1731 cm⁻¹) and olefinic bond (1639 cm⁻¹). One olefinic methine at $\delta_{\rm H}$ 5.36 (1H, td, J = 7.2, 0.8 Hz), one oxymethylene at $\delta_{\rm H}$ 4.68 (2H, d, J = 7.1 Hz), one formyl group at $\delta_{\rm H}$ 8.07 (1H, s), two tertiary methyl at $\delta_{\rm H}$ 0.85 (3H, d, J = 6.6 Hz) and $\delta_{\rm H}$ 0.84 (3H, d, J = 6.6 Hz), three quaternary methyls ($\delta_{\rm H}$ 0.86, s; 0.87, s; and 1.71, s) including one bearing sp^2 one were observed in the ¹H NMR. The ¹³C NMR spectra demonstrated 21 carbon signals covering a formyl carbon ($\delta_{\rm C}$ 161.1), two olefinic ones ($\delta_{\rm C}$ 143.8, and 117.3), five methyls ($\delta_{\rm C}$ 22.7, 22.6, 19.8, 19.7, and 16.4), nine methylenes ($\delta_{\rm C}$ 24.8–60.8) including one oxidized, three sp^3 methines ($\delta_{\rm C}$ 27.9, 32.7 and 32.8), and one sp^3 quaternary carbon. Considering of 3 degrees of unsaturation these evidences unambiguously disclosed that it was a formate of monocyclic diterpenoid. All the NMR data of **1** were closely similar to those of the known compound **2**, cassipourol [9], except for the only difference of a formyloxy at C-15 in **1** instead of a hydroxyl in cassipourol. This hypothesis was supported by the HMBC cross peaks (as Figure 2) between H-1' ($\delta_{\rm H}$ 8.07) and C-15 ($\delta_{\rm C}$ 60.8), and between H-15 ($\delta_{\rm H}$ 4.68) and C-13 ($\delta_{\rm C}$ 143.8), C-14 ($\delta_{\rm C}$ 117.3) and C-1' ($\delta_{\rm C}$ 161.1).

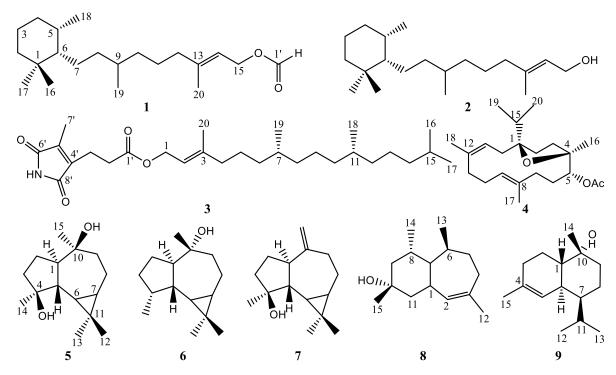


Figure 1. The chemical structures of compounds 1-9

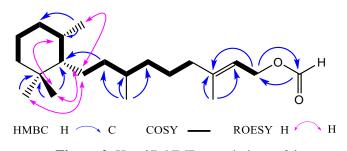


Figure 2. Key 2D NMR correlations of 1

Further HMBC correlations combing with the ¹H-¹H COSY correlations (Figure 2) of H-2/H-3/H-4/H-5/H-6, of H-7/H-8/H-9/H-10/H-11/H-12 and of H-14/H-15 confirmed the above deduction. The stereochemistry of compound **1** was presumed to be as shown (as Figure 2) on the basis of the ROESY cross peaks from H-7 ($\delta_{\rm H}$ 1.34) to H-17 ($\delta_{\rm H}$ 0.86) and H-18 ($\delta_{\rm H}$ 0.84) and from H-6 ($\delta_{\rm H}$ 1.07) and H-5 ($\delta_{\rm H}$ 1.54) to H-16 ($\delta_{\rm H}$ 0.87). Additionally, their almost identical chemical shifts of C-5–C-7 and C-18 also supported this deduction. Detailed examination of the ¹³C NMR data of C-12–C-15 and C-20 suggested that compound **1** had the same *E*-configuration of the double bond at C-13 as cassipourol [9], which was confirmed by the ROESY connections of CH₃-20 ($\delta_{\rm H}$ 1.71) and H-15 ($\delta_{\rm H}$ 4.68), and of H-12 ($\delta_{\rm H}$ 2.01) and H-14 ($\delta_{\rm H}$ 5.36). Consequently, the structure of **1** was established as cassipouryl formate. The biosynthetic pathway of diterpenoids **1–4** was discussed as the reference [10] (Figure 3).

	1			1	
No	$\delta_{ m H}$	$\delta_{ m c}$	No	$\delta_{ m H}$	$\delta_{ m C}$
1		36.6	12	2.01 m, 1.14 m	39.8
2	1.14 m	39.4	13		143.8
3	1.38 m	24.8	14	5.36 td (7.2,	117.3
4	1.26 m	37.3	15	4.68 d (7.2)	60.8
5	1.54 m	27.9	16	0.87 s	22.7
6	1.07 m	32.8	17	0.86 s	22.6
7	1.20 m, 1.34	24.5	18	0.84 d (6.6)	19.7
8	1.38 m	37.3	19	0.85 d (6.6)	19.8
9	1.38 m	32.7	20	1.71 s	16.4
10	1.26	37.4	1'	8.07 s	161.1
11	1.38 m, 1.45	25.0			
1 12					

Table 1. ¹H NMR and ¹³C NMR data for compounds 1-4 (δ in ppm)^a.

^{a 1}H NMR and ¹³C NMR Data for 1 in CDCl₃ at 600 MHz and 150 MHz, respectively.

The known compounds were assigned as cassipourol (2) [9], 4',5'-dehydrodiodictyonema (3) [11], acetyl incensole (4) [12], aromadendrane- 4β , 10β -diol (5) [13], (-)-globulol (6) [14], spathulenol (7) [15], 3β , 6β , 8α , 10β -tetramethylwiddrane-2(3)-en- 10α -ol (8) [16], and α -cadinol (9) [17] according to their NMR and MS data as well as the comparison with the reported values.

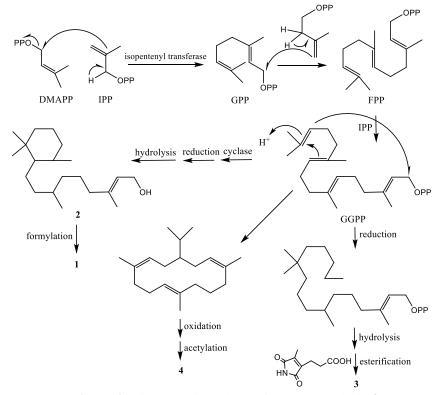


Figure 3. Biosynthetic pathway for compounds 1-4.

In summary, one new monocyclic diterpenoid, together with three known diterpenoids and five sesquiterpenoids were isolated from the twigs and leaves of *S. fluviatile*. This was the first examples for the chemical constituents of *S. fluviatile* and the first report on the diterpenoids of genus *Syzygium*.

Terpenoids from Syzygium fluviatile

This discovery provided new evidences not only for the chemical diversity of the genus *Syzygium*, but also for the chemotaxonomic makers of *S. fluviatile*.

Acknowledgments

This work was financially supported by the National Natural Science Foundation of China (No. 31560098).

Supporting Information

Supporting Information accompanies this paper on <u>http://www.acgpubs.org/journal/records-of-natural-products</u>

ORCID 💿

Dingli Zhang: <u>0000-0002-9073-6761</u> Yikao Hu: <u>0000-0002-3616-8556</u> Li Wang: <u>0000-0002-7004-4725</u> Shengxiong Huang: <u>0000-0002-3616-8556</u> Yan Zhao: <u>0000-0002-1638-7751</u> Mengjia Li: <u>0000-0002-7454-3767</u> Feng Li: <u>0000-0002-3943-5058</u> Yong Zhao: <u>0000-0002-3996-2480</u>

References

- [1] L. W. Tian, M. Xu, D. Wang, H. T. Zhu, C. R. Yang and Y. J. Zhang (2011). Phenolic constituents from the leaves of *Syzygium forrestii* Merr. and Perry, *Biochem. Syst. Ecol.* **39**, 156-158.
- [2] G. Q. Li, Y. B. Zhang, P. Wu, N. H. Chen, Z. N. Wu, L. Yang, R. X. Qiu, G. C. Wang and Y. L. Li (2015). New phloroglucinol derivatives from the fruit tree *Syzygium jambos* and their cytotoxic and antioxidant activities, *J. Agric. Food Chem.* **63**, 10257-10262.
- [3] T. Manaharan, D. Appleton, H. M. Cheng and U. D. Palanisamy (2012). Flavonoids isolated from *Syzygium aqueum* leaf extract as potential antihyperglycaemic agents, *Food Chem.* **132**, 1802-1807.
- [4] F. F. Liu, T. Yuan, W. Liu, H. Ma, N. P. Seeram, Y. Y. Li, L. Xu, Y. Mu, X. S. Huang and Y. L. Li (2017). Phloroglucinol derivatives with protein tyrosine phosphatase 1B inhibitory activities from *Eu -genia jambolana* seeds, *J. Nat. Prod.* 80, 544-550
- [5] Y. K. Hu, L. Wang, Y. Y. Li, M. J. Li, W. Xu, Y. Zhao, F. Li and Y. Zhao (2018). Five new triterpenoids from *Syzygium samarangense* (Bl.) Meer. et Perry, *Phytochemistry Lett.* **25**, 147-151.
- [6] S. H. Xu, W. Xu, L. Wang, Y. K. Hu, J. P. Liu, Y. Zhao, M. J. Li, F. Li, S. X. Huang and Y. Zhao (2018). New phloroglucinol derivatives with protein tyrosine phosphatase 1B (PTP1B) inhibitory activities from *Syzygium austroyunnanense*, *Fitoterapia* 131, 141-145.
- [7] T. Manaharan, D. Appleton, H.M. Cheng and U.D. Palanisamy (2012). Flavonoids isolated from *Syzygium aqueum* leaf extract as potential antihyperglycaemic agents, *Food Chem.* **132**, 1802-1807.
- [8] J. Chen (1984). Flora of China. Vol 53 (1). Beijing: Science Press, pp. 72.
- [9] V. S. P. Chaturvedula, A. Norris, J. S. Miller, F. Ratovoson, R. Andriantsiferana, V. E. Rasamison and D. G. I. Kingston (2006). Cytotoxic diterpenes from *Cassipourea madagascariensis* from the Madagascar rainforest, J. Nat. Prod. 69, 287-289.
- [10] N. Shimizua, D. Sakataa, E. A. Schmelzb, N. Moric and Y. Kuwaharad (2017). Biosynthetic pathway of aliphatic formates via a Baeyer–Villiger oxidation in mechanism present in astigmatid mites, *PNAS*, **114**, 2616-2621.
- [11] P. Yang, D. Q. Liu, T. J. Liang, J. Li, H. Y. Zhang, A. H. Liu, Y. W. Guo and S. C. Mao (2015). Bioactive constituents from the green alga *Caulerpa racemosa, Bioorg. Med. Chem.* 23, 38-45.
- [12] F. Wang, Z. L. Li, T. Liu and H. M. Hua (2009). Cembrane diterpenes in olibanum, *China J. Chin. Mater. Med.* 34, 2477-2480.
- [13] I. C. Moreira, J. H. G. Lago, M. C. M. Young and N. F. Roque (2003). Antifungal aromadendrane sesquiterpenoids from the leaves of *Xylopia brasiliensis, J. Brazil. Chem. Soc.* **14**, 828-813.

- [14] J. Wang, J. J. Xu, W. Qiao and S. A. Tang (2016). Chemical constituents from fruits of *Eucalyptus globulus, Chin. Tradit. Herb. Drugs* 47, 4336-4339.
- [15] P. Du, Z. R. Hu-Yang, W. C. Zhang and G. B. Xie (2017). Study on chemical constituents of *Uvaria* grandiflora, China Med. Herald. 14, 20-22.
- [16] T. Muhammad, S. Deny and F. R. Mohamad (2013). A new sesquiterpene from *Knema patentinervia*, *Chem. Nat. Compd.* **48**, 985-987.
- [17] M. L. D. Miranda, F. R. Garcez, A. R. Abot and W. S. Garcez (2014). Sesquiterpenes and other contituents from leaves of *Pterodon pubescens* Benth (Leguminosae), *Quím. Nova* **37**, 473-476.



2020 ACG Publications

195