

records of natural products

An Update on Phytochemistry and Biological Activities of

Cinnamomum

Meiting Wu ^(D), Zhiyi Lin ^(D)², Bingbing Huang ^(D)¹, Kuo Xu ^(D)⁴, Shuangquan Zou ^(D)², Lin Ni ^(D)^{1,3*} and Yuxi Chen ^(D)^{1,3}

¹ College of Plant Protection, Fujian Agriculture and Forestry University, Fuzhou, Fujian 350002, P. R. China

² College of Forestry, Fujian Agriculture and Forestry University, Fuzhou, Fujian 350002, P. R.

China

³ Key Laboratory of Biopesticide and Chemical Biology, Ministry of Education, Fujian Agriculture and Forestry University, Fuzhou, Fujian 350002, P. R. China

⁴ Tobacco Research Institute of Chinese Academy of Agricultural Sciences, Qingdao, Shandong

266101, P. R. China

(Received April 26, 2021; Revised May 25, 2021; Accepted May 26, 2021)

Abstract: The genus Cinnamomum belongs to the family *Lauraceae* and contains about 250 species. Cinnamomum plants have great economic value and have been widely used in the pharmaceutical, chemical, food and cosmetic industries. A great deal of research on the chemical constituents and their various biological activities has been conducted on only 20 species of Cinnamomum. We have already summarized the chemical structures and bioactivities of terpenoids from Cinnamomum. Herein, we give an update on other types of compounds and their biological activities. According to the findings, 380 chemical compounds obtained from Cinnamomum, including lignans, butanolides, flavonoids, phenylpropanoids, alkaloids and other compounds are summarized, and their corresponding unique chemical structures and significant biological activities are introduced in this paper.

Keywords: *Cinnamomum*; phytochemistry; immunomodulatory; anti-inflammatory; antioxidant activity. © 2021 ACG Publications. All rights reserved.

^{*}Corresponding author: E-Mail: <u>nilin_fjau@126.com</u> (L. Ni)

The article was published by ACG Publications
http://www.acgpubs.org/journal/records-of-natural-products January-February 2022 EISSN:1307-6167
DOI: http://doi.org/10.25135/rnp.262.21.04.2053
http://doi.org/10.25135/rnp.262.21.04.2053
http://doi.org/10.25135/rnp.262.21.04.2053

http://doi.org/10.25135/rnp.262.21.04.2053

1. Introduction

The family Lauraceae contains about 45 genera and 2500 species that are economically important in the pharmaceutical, chemical, food and cosmetic industries. As one of the largest genera of Lauraceae, *Cinnamomum* comprises approximately 250 species, which is represented by evergreen trees and shrubs. *Cinnamomum* plants are mainly distributed in tropical and subtropical Asia, Australia and the Pacific islands [1]. There are about 46 species in China, which are endemic in the southern religions, with the most species in Yunnan province, followed by Guangdong and Sichuan [2].

Cinnamomum species have been used as important sources of traditional medicine, timber, edible fruits, spices, and perfumes in China for a long history [3]. Some *Cinnamomum* species, such as *C. cassia, C. zeylanicum, C. tamala* and *C. wilsonii*, are famous herbs that have a long history of being used as medicine. Cinnamomi cortex, which is obtained from some *Cinnamomum* species, has been used for treating cardiovascular, chronic gastrointestinal and inflammatory diseases [4, 5]. The extracts from *Cinnamomum* plants have been reported to show various biological activities, including immunomodulatory, anti-inflammatory, anticancer and other activities; moreover, many studies have also been done on the activity of monomer compounds [6-10] However, the relationship between the activities of the extracts from *Cinnamomum* and those of the monomer compounds has not been fully elucidated Therefore, this paper aims to reveal the activity relationship between extracts and compounds from the genus, which may provide a theoretical basis for the discovery of active ingredients from *Cinnamomum* and better utilization of *Cinnamomum* plants.

To date, the research on bioactive constituents from *Cinnamomum* is a research focus in China and many experiments have been done. There have been over 500 compounds isolated from *Cinnamomum* with various pharmaceutical activities. Many constituents have been confirmed to be effective in *in vivo* experiments or even clinically used in treatment for various diseases, such as cinnamaldehyde, cinnamic acid, sesamin, camphor, borneol and so on. There are many lead compounds that are under research and development and many new compounds with unique structures under activity screening tests. Therefore, a summary of the active ingredients of *Cinnamomum* is necessary, which will help to explore more valuable lead compounds.

Among the *Cinnamomum* species, *C. cassia* is the most important species in the genus *Cinnamomum* and has been thoroughly studied. Around 300 constituents with many new skeletons have been found in this species. There are also many studies on phytochemistry of other species, such as *C. burmannii*, *C. camphora*, *C. kotoense* and *C. subavenium*. And the constituents obtained from *Cinnamomum* have shown a variety of biological activities, which provides a lot of bioactive ingredients for the development of new drugs.

We have previously introduced the structures of terpenoids from *Cinnamomum* and their biological activities [11]. Herein, we summarized other types of constituents from *Cinnamomum* plants in this paper, including lignans, butanolides, flavonoids, phenylpropanoids, alkaloids, and other compounds. Also, their pharmacological activities are also introduced in this review, which covers antioxidant, immunomodulatory, anti-inflammatory, anticancer and other effects. Many compounds have been proved to be potent bioactive constituents through a lot of assays and thus are promising treatment for many diseases.

2. Chemical Constituents and Their Biological Activities

A great deal of phytochemical research has been conducted on a few species. Excluding the studies that only give focus on volatile oils, we have summarized a total of 380 constituents from 17 *Cinnamonum* species, which include 82 lignans, 46 butanolides, 65 flavonoids, 76 phenylpropanoids, 19 alkaloids and 92 other compounds.

2.1. Lignans

Lignans are an important part of the secondary metabolites of *Cinnamomum* species, which have high content and abundant structural types. There are 82 lignans isolated from *Cinnamomum*

plants (Table 1 & Figure 1), including five diarylbutanes (1-5), ten arylnaphthalenes (6-15), eleven tetrahydrofurans (16-26), sixteen bis-tetrahydrofurans (27-42), sixteen benzofurans (43-58), eight 8-O-4'-neolignan (59-66), four spirodienones (67-70), two biphenyls (71-72), three norlignans (73-75), four sesquilignans (76-79), one dimer (80), and two neolignans (81-82).

Spirolignans can be rarely found in natural products. Herein, Lai *et al.* [12] separated two pairs of spirodienone neolignan racemates (**67-70**) with a rare 2-oxaspiro[4.5]deca-6,9-dien-8-one motif from *C. subavenium*. It was the first time to report spirodienone neolignans with this rare skeleton. Moreover, these compounds showed significant inhibitory effects against NO production in RAW264.7 mouse macrophages, with IC₅₀ values of 17.9, 5.6, 15.1, and 4.3 μ M, respectively. Among the four lignans, **70** exhibited strongest inhibitory effects while **67** weakest. Thus, the methoxy substituent at C-5 enhanced the inhibitory effects of the compound. In addition, **69** and **70** showed much stronger inhibitory effects than **67** and **68**, respectively. Thus, the chirality of the spirolignans significantly affected the inhibitory effects on the NO production in the RAW264.7 mouse macrophages.

The lignans obtained from *Cinnamomum* contains two glycosides (**47**, **58**), both of which were isolated from the bark of *C. cassia*, and **58** is a unique compound of *Cinnamomum* plants [2, 13]. It is noteworthy that hydroxyl groups at the 9-positions of some lignans (**3-4**, **12-13**) formed ester groups with ferulyl groups. This special structure was only found in *C. osmophloeum* [14]. Moreover, C-7, C-7', C-8 and C-8' of compound **82** isolated from *C. balansae* formed a cyclobutane, which is also a relatively rare structure [15].

Biphenyllignans are common in separation, such as magnolol analogues [16]. However, according to Liu *et al.* [17], the C-7 and C-8 positions of biphenyl lignans (**71-72**) formed peroxy bonds, which are rare in natural products. Moreover, both the compounds have not been found in other genera and have showed certain neuroprotective activities.

Sesamin (34) has high content in *Cinnamomum* plants, especially in the leaves of *Cinnamomum camphora* [7], which has showed various biological activities *in vivo* and *in vitro*. Treatment with 34 could accelerate wound healing by promoting the proliferation, adherence, migration in human umbilical vein endothelial cells. It could also promote neogenesis of granulation tissue and deposition and remodeling of the collagen matrix in a rat model [18]. According to Majdalawieh *et al* [19], the anti-hyperlipidemic activities of 34 have been proven in many *in vivo* studies. It mainly exerts the anti-hyperlipidemic effects by downregulating the activity of $\Delta 5$ desaturase, suppressing the activity of SREBP-1, and inhibiting the process of PUFA chain elongation via sesamin-dependent upregulation of PPAR α regulatory pathways. Moreover, many other *in vivo* experiments of sesamin have been conducted and 34 has the potential to treat or prevent intestinal ischemia, cardiovascular diseases, lung inflammation and many other diseases [20-22]. In a clinical trial, sesamin supplement could relieve clinical symptoms and pathological changes that were caused by inflammatory impairment in patients with rheumatoid arthritis [23].

Many lignans from different plant sources have been reported to show good neuroprotective activities and some have been used in treatment of neurodegenerative diseases [24]. Lignans **17**, **71**, **72** and **85** were tested for their neuroprotective effects against tunicamycin-induced cell death in SH-SY5Y cells. All these complounds exhibited strong neuroprotective effects with EC_{50} values ranging from 21 to 75 μ M [17].

Phytochemistry and biological activities of Cinnamomum

Table 1	 Ligna 	ns from	Cinnamomum	genus
---------	---------------------------	---------	------------	-------

No	Compounds	Plants	Rof	No	Compounds	Plante	Rof
1	secoisolariciresinol	C O	[25 28]	42	4-ketopinoresinol	1 Tanto	[28]
2	methoxysecoisolariciresinol	с,0 С	[23, 20]	43	(7S 8R)-lawsonicin	c	[25]
3	Secoisolariciresinol diferulovl ester	0	[14]	44	urolignoside	7	[29]
	9.9'-Di-O-ferulovl-(+)-5.5'-	0	[1.]		9.9'-dihydroxy-3.4-methylenedioxy-	-	[=>]
4	dimethoxy secoisolariciresinol	0	[14]	45	3'-methoxy[7-O-4'.8.5']neolignan	с	[17]
5	cinnacassoside A	с	[13]	46	(7R,8S)-ficusal	с	[17]
	(6R,7R,8R)-7a-[(β-D-						1001
6	glucopryanosyl)oxy] lyoniresinol	С	[2]	47	samwiside	с	[30]
-	(6S,7R,8R)-7a-[(β-D-	_	[2]	40	() lenteleniel C	_	[25]
/	glucopryanosyl)oxy] lyoniresinol	С	[2]	40	(+)-ieptolepisol C	с	[25]
	(6R,7S,8S)-7a-						
8	[(β-D-glucopryanosyl)oxy]	с	[2]	49	cinncassin D	с	[25]
	lyoniresinol						
9	isolariciresinol	с	[25]	50	picrasmalignan A	с	[25]
10	5-methoxy-isolariciresinol	с	[28]	51	spicatolignan B	с	[13]
11	(–)-lyoniresinol	с	[31]	52	balanophonin	с	[17]
12	(/'S,8'R,8R)-Lyoniresinol-9-O-(E)-	0	[14]	53	5-methoxybalanophonin	с	[17]
	feruloyl ester						
13	(/'S,8'R,8R)-Iyoniresinol-9,9'-di-O-	0	[14]	54	hierochin B	с	[17]
	(E)-refutioyi ester						
14	(-)-iyoimesinoi 5a-O-p-D- glucopyranoside	с	[32]	55	simulanol	с	[17]
15	schizandriside	d	[33]	56	salvinal	C	[17]
16	cinnacassin G	C C	[17]	57	herpetal	c	[17]
17	cinnacassin H	c	[17]	58	cinnacassoside B	c	[13]
	(+)-(7'R.8R.8'R)-5.5'-	·	[1,]		(-)-ervthro-(7R.8S)-guajacylglycerol-	•	[10]
18	dimethoxylariciresinol	с	[25]	59	β -O-4'-sinapoyl ether	с	[25]
10	(+)-(7'S,8R,8'R)-5,5'-		[0.5]	60	(-)-erythro-(7S,8R)-syringylglycerol-		[0.5]
19	dimethoxylariciresinol	с	[25]	60	β -O-4'-sinapyl ether	с	[25]
20	5'-methoxylariciresinol	с	[34]	61	cinncassin E	с	[25]
21	ainneasain M	0	[17]	67	(+)-threo-(7S,8S)-guaiacylglycerol-β-	0	[25]
41	chinacassin M	C	[1/]	02	coniferyl aldehyde ether	C	[23]
22	(+)-episesaminone	d	[35]	63	(+)-erythro-(7S,8R)-guaiacylglycerol-	C	[25]
	() opisesummone	u	[55]	05	β-coniferyl aldehyde ether	C	[25]
•••			50.07		1-(4-hydroxy-3-methoxyphenyl)-2-[3-		
23	dehydroxycubebin	р	[36]	64	(3-hydroxy-1-propenyl)-5-	с	[17]
					metnoxypnenoxy]-1,3-propanediol		
24	cubebin	р	[36]	65	(+)-erythro-(/R,85)-gualacyigiyceroi-	с	[25]
					1.2.3 propagatrial 1 [4 (1P 2P) 2		
					hydroxy-(4-hydroxy-3-methoxy-		
25	hinokinin	р	[36]	66	phnevl)-1-(hydroxymethyl)ethoxyl-3-	с	[37]
					methoxyphneyl		
	(7'S,8S,8'R)-4,4'-dihydroxy-				51 5		
26	3,3',5,5'-tetramethoxy-7',9-	с	[17]	67	(+)-subaveniumin A	s	[12]
	epoxylignan-9'-ol-7-one						
27	(+)-syringaresinol	b,c,k,m,r,s,t	[38-40]	68	(+)-subaveniumin B	S	[12]
28	(+)-yangambin	b,c	[28, 38]	69	(-)-subaveniumin A	S	[12]
29	clemaphenol A	k	[41]	70	(-)-subaveniumin B	S	[12]
30	cinnacassin F	с	[17]	71	Cinnacassin I	с	[17]
31	(+)-pinoresinol	b,c,d	[28, 31, 42]	72	cinnacassin J	с	[17]
			100 103		6-hydroxy-2-(4-hydroxy-3,5-		
32	(+)-medioresinol	b,c	[28, 43]	73	dimethoxy-phenyl)-3,7-dioxabicyclo-	c	[13]
22		1	F 4 0 1	- 4	[3.3.0]-octane		F 1 77
33	pinoresinol methyl ether	d	[42]	74	zhebeiresinol	c	[17]
34 25	sesamin	v,u,J,K,n,S,t	[2/, 44-4/]	15	(-)-(/K,8K,8K)-acuminatolide	a	[42]
33 24	(+)-missesamin	u,j,n	[27, 47]	/0 77	ouddienol A	c	[1/] [17]
30 27	(+)-episesaililli physiatilal	u,j 1-	[27,47] [/1]	11 79	ficuses quilignen A	C	[1/] [29]
32	pinviation	K d	[41] [42]	70 70	huddlenol C	C C	[20] [28]
30	piperitor	u	[42]	19	ouddienor C	U	[20]

Wu et.al., Rec. Nat. Prod. (2022) 16:1 1-26

39	9α-hydroxysesamin	d	[48]	80	hedyotisol A	с	[17]
40	9β -hydroxysesamin	d	[48]	81	cinnaburmanin A	b	[49]
41	l-asarinin	d	[48]	82	cinbalansan	a	[15]

a-C. balansae, b-C. burmannii, c-C. cassia, d-C. camphora, i-C. inunctum, j-C. insulari-montanum, k-C. kotoense, m-C. macrostemon, o-C. osmophloeum, p-C. parthenoxylon, q-C. philippinense, n-C. randaiense, r-C. reticulatum, s-C. subavenium, t-C. tenuifolium, u-C. trichophyllum, z-C. zeylanicum. The same as below.

The anti-inflammatory activities of the isolated lignans were evaluated on production of nitric oxide (NO) induced by lipopolysaccharide (LPS) in BV-2 microglial cells. Compounds including **19** and **61-63** showed significant inhibition activities with IC₅₀ values of 17.5, 17.6, 17.7 and 18.7 μ M, respectively. Other compounds exhibited moderate inhibitory activities, including **18**, **20**, **27**, **43**, **59**, **60** and **65**. In addition, it was noticed that 8-*O*-4'-lignans showed significant inhibition with IC₅₀ values ranging from 17.6 to 42.0 μ M. And among them, lignans with acrylaldehyde group at C-1' exhibited highest anti-inflammatory activities [25].

Compound **50** significantly inhibited NO production and suppress TNF- α and IL-6 release at three doses (10, 30 and 100 μ M) in LPS-activated macrophage RAW 264.7 cells. Furthermore, the inhibitive action of **50** was more potent than that of the positive control hydrocortisone, a commonly used anti-inflammatory drug. The substance can also inhibit the overexpression of iNOS and COX-2 and the activity of iNOS and COX-2 enzymes in the assays [26].

Three lignan esters, including compounds **4**, **12** and **13**, were tested for their cytotoxicities against HepG2, Hep3B, and Ca9-22 cancer cells. Compounds **12** and **13** have significant cytotoxicities on three cancer cell lines with EC_{50} values of less than 20 and 10 µg/mL respectively, while compound 4 showed moderate effect. The structure-activity relationships are as followed: (a) The cyclolignans (**12** and **13**) demonstrated stronger effects than the dibenzylbutane lignan (**4**) on these three cancer cell lines. (b) The lignan with two feruloyl groups (**13**) showed stronger activities than that with only one (**12**). Thus, both C-9 and C-9' feruloyl groups significantly increased the cytotoxicity of the compounds [14].

In another assay, the cytotoxicity of **34** on Hep G2 was investigated. The percentage of Hep G2 cells in the S phase decreased from 40% to 30% after treatment with 200 μ M **34** for 24 hours, showing a slight cytotoxic effects [27]. Moreover, the lignan **8** was reported to show significant inhibitory effects on ConA-induced T cell proliferation with an inhibition ratio of 80.1% at a concentration of 200 μ M, whilst at low doses of 25 and 12.5 μ M, stimulated the proliferation of T cells. Some compounds exhibited weak inhibition, including compounds **6**, **238** and **362** [2].



OCH3 OCH3 OH

Η

Η

Η

Η

Η

 $OCH_3 OH$

-OCH₂O-

-OCH₂O-

-OCH₂O-

OCH₃ OH

OCH₃ OH

Η

Η

Η

Η

Η

Η

Η

7S, 7'S

7S, 7'S

7S, 7'S

7R, 7'R

7R, 7'S

7R, 7'S

7S, 7'S

32 OCH₃ OH

34

35

36

37

38

33 OCH₃ OCH₃ H

-OCH₂O-

-OCH₂O-

-OCH₂O-

-OCH₂O-

-OCH₂O-

OHC

	R_1	R_2	R ₃	
43	OH	OCH ₃	CH ₂ OH	7S, 8R
44	OGlc	OCH ₃	CH ₂ OH	7S, 8R
45	-OCH	1 ₂ O-	CH ₂ OH	7S, 8R
46	OH	OCH ₃	СНО	7R, 8S
47	OH	OCH_3	CH ₂ OGlc-Api	7S, 8R

 R_1 R_2 R_3 51 OH Η COOH 7R, 8S 52 OH Η CHO 7R, 8S 53 OH OCH₃ CHO 7R, 8S **54** OCH₃ H CH₂OH 7S, 8R 55 OH OCH₃ CH₂OH 7S, 8R

Phytochemistry and biological activities of Cinnamomum

Figure 1. The structures of lignans from Cinnamomum

2.2. Butanolides

Butanolides are not very common in the separation of natural products. However, there have been 46 butanolides obtained from the genus *Cinnamomum* (Table 2 & Figure 2), which are important active ingredients with a variety of structure types. Furthermore, some extracts from *Cinnamomum* plants have been reported to show potent anticancer effects and butanolides may be the major active ingredients [50].

The butanolides isolated from the *Cinnamomum* include two simple γ -butyrolactones (83, 84), ten α,β -diphenyl- γ -butyrolactones (85-94), twenty-nine long-chain fatty alkyl-substituted γ -lactone (95-123) and five secobutanolides (124-128).

According to Liu *et al.* [17], the α,β -diphenyl- γ -butyrolactones (**85-94**) are a class of unique natural compounds that have only been isolated from *C. cassia* and thus could be used as potential chemotaxonomic markers for this species. Among these compounds, **85**, **88** and **90** has shown significant neuroprotective activities.

Compounds **96**, **99** and **100** were firstly obtained from *Cinnamomum kotoense* in 2006 and showed significant anti-proliferation activity [41, 51]. Isoobtusilactone A (**97**) and obtusilactone A (**98**) are common in *Cinnamomum* plants and both have been found in seven *Cinnamomum* species. Isophilippinolide A (**103**) and philippinolide A (**104**) were firstly found in the roots of *Cinnamomum philippinense* and also showed potent anticancer activities [52]. Compounds **97--110** share the same β -hydroxy- γ -methylene- α , β -unsaturated- γ -lactone skeleton.

According to the findings, many butanolides from the genus were proved to exhibit potent anticancer effects and it is illustrated as follows. It was reported that **112**, **113**, **116** and **127** can induce significant cell death in the human colorectal cancer line SW480. At a dose of 50 μ M, SubG1 levels were increased to 25.4% and 23.7% respectively, showing that **112** and **113** induced significant DNA damage. The subG1 population in cells treated with **116** and **127** was 11.0% and 9.1%, respectively. All these compounds caused DNA damage in a dose-dependent manner and at a dose of 75 μ M, SubG1

expression was increased up to 23.4%-47.2% [44]. In another assay, **106** and **116** also showed potent cytotoxicity, with SubG1 levels of 47.2 and 27.4%, respectively [53].

Subamolide A (112), only obtained from *Cinnamomum subavenium*, showed significant effects in the screening of anti-cancer activities *in vivo* [54]. The compound was demonstrated to selectively induce apoptosis in two cancerous human urothelial carcinoma cell lines (NTUB1 and T24) by triggering the mitochondria-dependent apoptotic pathways and p53 and ERK1/2 activation [55]. Compound **112** induced apoptosis in human lung cancer cells A549 and NCI-H460 resulting from triggering mitotic catastrophe with apoptosis and caused a dramatic 70% reduction in tumor size in an animal model [56].

No.	Compounds	Plants	Ref.	No.	Compounds	Plants	Ref.
83	(R)-3-hydroxybutanolide	с	[37]	106	subamolide D	S	[50]
84	3-hydroxy-4,4-dimethyl-4- butyrolactone	i	[57]	107	subamolide E	s	[50]
85	cinncassin A	с	[17]	108	Linderanolide B	S	[44, 50]
86	Cinncassin A ₁	с	[17]	109	Isolinderanolide B	k,s	[51, 53]
87	Cinncassin A ₂	с	[17]	110	isoreticulide	r	[58]
88	Cinncassin A ₃	с	[17]	111	lincomolide B	k	[40]
89	Cinncassin A ₄	c	[17]	112	subamolide A	s	[44, 50]
90	Cinncassin A ₅	с	[17]	113	subamolide B	s	[44, 50]
91	Cinncassin A ₆	с	[17]	114	philippinolide B	q	[52]
92	Cinncassin A7	c	[17]	115	tenuifolide B	t	[45]
93	cinnamomulactone	c	[17]	116	subamolide C	s	[44, 50]
94	cinnamomumolide	c	[31]	117	kotomolide B	k	[41]
95	5R-methyl-3-heptatriacontyl-2(5H)- furanone	c	[34, 59]	118	kotomolide	k	[60]
96	Cinnakotolactone	k	[51]	119	5-dodecanyl-4-hydroxy-4-methyl-2- cyclopentenone	d	[60]
97	isoobtusilactone A	d,j,k,n, r,s,t	[39, 45, 47]	120	Kotolactone A	k	[40]
98	obtusilactone A	d,j,n,k, r,s,t	[39, 45, 47]	121	kotolactone B	k	[40]
99	isokotomolide A	k	[41]	122	2-acetyl-5-dodecylfuran	k	[40]
100	kotomolide A	k	[41]	123	2-acetyl-5-methylfuran	k	[40]
101	tenuifolide A	t	[45]	124	secokotomolide A	k	[40]
102	isotenuifolide A	t	[45]	125	secokotomolide	k	[40, 60]
103	isophilippinolide A	q	[52]	126	secotenuifolide A	t	[45]
104	philippinolide A	q	[52]	127	secosubamolide	S	[44, 50]
105	isomahubanolide	d	[48]	128	secosubamolide A	S	[50]

Table 2. Butanolides from Cinnamomum genus

Compound **124** was also found to have significant cytotoxic effects on the human HeLa cell line. Compared to the vehicle control group, incubation with **124** at 0, 25, 50, and 100 μ M for 24 h induced apoptosis in sub-G1 phase at 1.4, 68.8, 75.6, and 81.8% [41]. Moreover, **126** showed cytotoxic activity against two human prostate cancer epithelial cell lines, DU145 and LNCaP, with EC₅₀ values of less than 7 μ M (equal to 3.45 μ g/mL) [45]. Inhibitory effects of **103** against the A375.S2 melanoma cell line were evaluated. Compared with untreated cells, treatment with 10, 25, 50, and 100 μ M **103** for 24 h resulted in a dose-dependent increase in the subG1 accumulation, extending from 2.34 to 3.92, 4.27, 8.79 and 14.11%, respectively [52].

Figure 2. The structures of butanolides from Cinnamomum

2.3. Flavonoids

Leaves are mostly used as a medicinal part in traditional Chinese medicine, which undoubtedly have high content of flavonoids. There have been 65 flavonoids (**129-193**) isolated from *Cinnamomum* plants (Table 3 & Figure 3), including seven simple flavones (**129-135**), thirty-one flavonols (**136-166**), one dihydroflavones (**167**), two dihydroflavonols (**168**, **169**), one chalcone (**170**), seventeen flavanes (**171-187**) and seven anthocyanidins (**188-193**). Flavonoids are representative constituents of *Cinnamomum* plants, exemplified by quercetin (**142**), kaempferol (**136**) and their glycosides. Flavonoids from *Cinnamomum* mostly share similar types with other genus, but some of them have uncommon skeletons and show many biological activities, especially antioxidant activities.

Compound **193** is a flavonol galactoside-lignan ester. The compound has a rare skeleton, in which the kaempferol moiety was connected to a diacyl moiety with a cyclobutane ring bearing two 4-hydroxyphenyl through a sugar moiety.

Many flavonoids are natural antioxidants. It was confirmed that the hydroxy group at the C-3' position of the B ring is essential for the antioxidant activity, which accounts for higher effect of rutin (146) compared to nicotiflorin (147) and isorhoifolin (133) in both concentrations of 10 and 20 μ M, respectively [61].

Compound **140**, isolated from *C. osmophloeum*, was proved to have antioxidant capacity through DPPH and NBT Assays, with the EC₅₀ values of 26.9 and 68.1 μ M, respectively [62]. Compound **155** exhibited stronger radical scavenging activity (65.21%) than **156** (17.40%) at 60 μ mol/L concentration in a DPPH assay, mostly because of the difference of coumaroyl group positions in the compounds [63]. The *in vitro* IC₅₀ values against DPPH for tiliroside (**158**) was found to be 60.40 μ g/mL and the ferric ion (Fe³⁺) reducing ability of **158** were 0.324 at the dose of 50 mg/mL [64].

Furthermore, DPPH assays were performed to evaluate the antioxidative potential of some compounds. Compound **358** exerted moderate antioxidant effect with IC₅₀ value of 75.03 μ M [65]. In another assay, compounds **44**, **144**, **146**, **188** and **320** showed free radical scavenging activities of 30.4, 60.3, 44.7, 39.3 and 77.3%, respectively, at 12.5 ppm concentration. Among these compounds, **320** showed the highest antioxidant activity in the β -carotenelinolate system [29].

In addition to antioxidant effects, some flavonoids were demonstrated to show other activities, including anti-inflammatory, anti-cancer, immunomodulatory and anti-hyperglycemic activities.

Four kaempferol glycosides (**160-162** and **166**) from *C. osmophloeum* leaves exerted a dosedependent inhibition on the production of NO, TNF-a and IL-12 from LPS/IFNc-activated macrophages. Among them, compound **162** showed the highest inhibitory activity, with significant inhibition at 10 μ M, and 41% of TNF-a production and 35% of IL-12 production of the positive control at 20 μ M [66].

Two flavonoids (**155** and **156**) were repeorted to show potent inhibitory activities in lung cancer cell line (A549 and NCI-H460) and breast cancer cell line (MCF-7 and MDA-MB-231), with IC₅₀ values ranging from 1.6 to 8.4 μ g / mL. Both of them show highest inhibition on NCI-H460 cell line, with IC₅₀ values of 4.6 and 1.6 μ g/mL, respectively [67].

In vitro tests conducted by Liu *et al.* showed that compound **187** stimulated cell proliferation of splenocytes and peritoneal macrophages, significantly enhanced the cytotoxicity of natural killer cells and increased CD^{4+} and CD^{8+} cell populations, showing good immunomodulatory activity. Moreover, **187** also induced effective phagocytic activation in macrophages [68].

Proanthocyanidins from *C. osmophloeum* twig extracts, including **188** and **190**, were found to be associated with anti-hyperglycemic capacity. Moreover, it was also found that the higher the degree of polymerization of the proanthocyanidins, the better the inhibition of α -Glucosidase [37]. Proanthocyanidins were also reported to show inhibitory effects against cyclooxygenase-2 (COX-2). In the assay, compounds **188-191** were tested for their inhibitory activities against the COX-2 enzyme isolated from human recombinant Sf9 cells and all of them exerted significant inhibition at doses of 10, 100, and 1000 µg/mL. Moreover, the tetramers (**190**, **191**) showed stronger inhibition than the trimers (**188**, **189**) [69].

No.	Compounds	Plants	Ref.	No.	Compounds	Plants	Ref.
					kaempferol 3-O-β-D-apiofuranosyl-		
129	tricetin-7-methyl ether	d	[70]	162	$(1\rightarrow 2)$ - α -L arabinofuranosyl-7-O- α -L-	0	[1]
					rhamnopyranoside		
130	4',6,7-trimethoxyflavone	d	[42]	163	herbacetin	р	[71]
131	apigenin	k	[40]	164	kaempferol-3-O-β-D-glucose(6→1)-α-L- rhamnoside	d	[48]
132	genkwanin	k	[40]	165	kaempferol-3-O- α -L-rhamnopyranosyl- (1 \rightarrow 2)- α -L rhamnopyranoside	0	[1]
133	isorhoifolin	р	[61]	166	kaempieroi 3-O-p-D-apioruranosy- (1 \rightarrow 4)-α-L-rhamnopyranosyl-7-O-α-L- rhamnopyranoside	0	[1]
134	luteolin	d	[70]	167	naringenin 5-O-β-D-glucopyranoside	s	[72]
135	luteolin-7-O-β-D glucoside	d	[70]	168	taxifolin	d	[73]
136	kaempferol	j,k	[40, 60]	169	dihydrokaempferol	d	[42]
137	kaempferol 3-O-ß-D-glucopyranoside	c,d,s	[28, 72]	170	phloridzin	s	[72]
138	kaempferol-3-0-a-L-rhamnopyranoside	d,o,p,s	[1, 71, 72]	171	3'-methoxyl-(-)-epicatechin	c	[37]
139	kaempferol-3-0-β-rutinoside	d	[72]	172	(-)-epicatechin	c,k,p,s	[44, 60]
140	kaempferol-7-O-a-L-rhamnopyranoside	0	[1]	173	5,7-dimethyl-3',4'-di-O-methylene-(±)- epicatechin	c	[31, 32]
141	kaempferol-3-O-α-L-rhamnopyranoside- 7-O-α-L rhamnopyranoside	0	[1]	174	5,3',dimethoxyl-(-)-epicatechin	c	[37]
142	quercetin	d,k,z	[1, 42, 60]	175	(-)-(2R,3R)-4'-hydroxy-5,7,3'- trimethoxyflavan-3-ol	b,c,k	[31, 40, 43]
143	quercetin 3-O-ß-D-glucopyranoside	c,d	[28, 70]	176	4'-methoxyl-(+)-catechin	с	[37]
144	quercetin 3-O-a-L-rhamnopyranoside	c,d,p,z	[1, 28, 29, 71]	177	7,4' -dimethoxyl-(+)-catechin	c	[37]
145	quercetin 3-O-α-D-arabinopyranoside	с	[28]	178	5,7,4' -trimethoxyl-(+)-catechin	c	[37]
146	rutin	d,p,z	[29, 61]	179	(+)-catechin	k,s,t	[40, 41, 45]
147	nicotiflorin	р	[61]	180	(-)-catechin	k,t	[40, 74]
148	isorhamnetin-3-O-β-D-glucopyranoside	d	[33]	181	(-)-afzelechin	с	[37]
149	isorhamnetin-3-O-β-rutinoside	d	[48]	182	(2S,3S)-3'-hydroxy-5,7,4'-trimethoxy- flavan-3-ol	d	[32]
150	quercetin 3-O- $(3'',4''-di-trans-p-coumaroyl)-\alpha-L-rhamnopyranoside$	с	[28]	183	(-)-(2R,3R)-5,7-dimethoxy-3',4'- methylenedioxy-flavan-3-ol	d	[42]
151	quercetin 3-O- $(2^{\circ}, 4^{\circ})$ -di- trans-p- coumaroyl)- α -L-rhamnopyranoside	c	[28]	184	(-)-epicatechin-3-O-β-glucoside	c	[37]
152	3"-trans-p-coumaroylquercitrin	с	[28]	185	(-)-epicatechin-6-β-glucoside	c	[37]
153	4"-trans-p-coumaroyl-kaempferol-3-O-α- L-rhamnoside	с	[28]	186	(-)-epicatechin-8-\beta-glucoside	c	[37]
154	4"-cis-p-coumaroyl-kaempferol-3-O-α-L- rhamnoside	c	[28]	187	proanthocyanidin A-1	c	[37]
155	kaempferol-3-O-(2",4"-di-E-p- coumaroyl)-α-L-rhamnopyranoside	r	[63]	188	cinnamtannin B-1	c,s,z	[29, 69, 72]
156	kaempferol-3-O-(3",4"-di-E-p- coumaroyl)-α-L-rhamnopyranoside	r	[63]	189	cinnamtannin D-1	c,s	[69, 72]
157	kaempferol 3-O-(3",6"-di-trans-p- coumaroyl)-B-D-glucopyranoside	с	[28]	190	parameritannin A-1	c	[69]
158	tiliroside	с	[28]	191	cassiatannin A	c	[69]
159	kaempferol 3-O-(3",6"-di-trans-p- coumaroyl)-ß-D-galactopyranoside	с	[28]	192	epicatechin- $(4\beta \rightarrow 8)$ -epicatechin- $(4\beta \rightarrow 8)$ -epicatechin	c	[34]
160	kaempferitrin	j,o	[1, 75]	193	cinnamomoside A	c	[17]
	kaempferol 3-O-β-D-glucopyranosyl-						
161	$(1\rightarrow 4)$ - α -L-rhamnopyranosyl-7-O- α -L- rhamnopyranoside	0	[1]				

Table 3. Flavonoids from Cinnamomum genus

159 $R_1 = R_2 = trans-p$ -coumaroyl **157** $R_1 = R_2 = trans-p$ -coumaroyl **158** R_1 =H R_2 =*trans-p*-coumaroyl

HO

HO

ĠН Ĉ

HO

ö ĠН

Figure 3. The structures of flavonoids from Cinnamomum

2.4. Phenylpropanoids

Figure 4. The structures of phenylpropanoids from *Cinnamomum*

Cinnamomum plants are characterized by the aroma related to phenylpropanoids. Also, phenylpropanoids are common in *Cinnamomum* species with high content, especially in their volatile oils. A total of 76 phenylpropanoids (**194-269**) have been isolated from *Cinnamomum* plants (Table 4 & Figure 4). Only four coumarins (**247-250**) were obtained from *Cinnamomum* but they have shown great biological activities. Compounds **247** and **248** were found in three *Cinnamomum* species respectively and moreover, **248** and **249** have been reported to show potent anti-inflammatory effects. Coumacasia (**250**) was obtained for the first time from *C. cassia* in 2013 and exhibited significant cytotoxic activity [76]. Cinnamaldehyde (**194**) is the main component of essential oil from *C. cassia*. In 2012, Ngoc *et al.* separated cinnacasolide B (**208**) for the first time from in *C. cassia* [77]. Cinnamomdiol A (**237**) is a 3-(3,4-methylenedioxyphenyl)-propane-1,2-diol glycoside which was

isolated for the first time from *C. camphora* [73]. Some phenylpropanoids are common in *Cinnamomum* species, including compounds **202**, **203**, **209** and **227**. This finding supported the chemotaxonomic relationship among *Cinnamomum* species.

Table 4. Phenylpropanoids from <i>Cinna</i>	<i>momum</i> genus	S
--	--------------------	---

No.	Compounds	Plants	Ref.	No.	Compounds	Plants	Ref.
194	cinnamaldehyde	b,c	[78, 82]	232	erthro-guaiacy lglycerol	с	[37]
195	2-methoxycinnamaldehyde	с	[82]	233	4-methoxy guaiacyl glycerol 7-O-β-D- glucopyranoside	c	[37]
196	2-hydroxycinnamaldehyde	с	[82]	234	D-threo-guaiacylglycerol 7-O-β-D- glucopyranoside	s	[83]
197	coniferaldehyde	с	[82]	235	cinnacassoside D	с	[13]
198	cassiferaldehyde	с	[82]	236	3-(3,4-methylenedioxyphenyl)-1,2- propanediol	d,p	[73]
199	4-methoxycinnamaldehyde	с	[34]	237	cinnamomdiol A	d	[36, 73]
200	cinnamyl acetate	b	[43]	238	methyl-phenylpropanoate-2-O-β-D- apio-furanosyl-(1→6)-O-β-D- glucopyranoside	с	[2]
201	trans-cinnamaldehyde	b,m	[43, 84]	239	dihydomelilotoside	с	[82]
202	cinnamyl alcohol	b,c,j,m	[43, 75, 84]	240	methyl dihydromelilotoside	с	[82]
203	cinnamic acid	b,c,j,m,n,z	[43, 84-86]	241	dihydrocinnacasside	с	[82]
204	O-coumaric acid	C,S	[82, 87]	242	p-dihydrocoumaric acid	r	[88]
205	2-hydroxy-cinnamyl alcohol	с	[37]	243	3-phenylpropanol	с	[34]
206	E)-3-(2-methoxyphenyl)prop-2- en-1-ol	с	[37]	244	benzenepropanal	c	[34]
207	rosavin	с	[77, 82]	245	2-ethyl-5-propylphenol	с	[34]
208	cinnacasolide B	c	[77]	246	stearyl ferulate	d	73
			[44, 45, 58,				[38, 43, 47,
209	ferulic acid	k,r,s,t,z	88]	247	coumarin	b,c,j,m	84]
210	trans-methyl p-coumarate	r	[88]	248	scopoletin	b.d.p	[38, 42, 61]
211	trans-coumaric acid	k	[40]	249	6,7-dimethoxycoumarin	d	[42]
212	(E)-3-(3- methoxyphenyl)acrylaldehyde	с	[37]	250	coumacasia	с	[76]
213	3-(3,4-dimethoxyphenyl)-2- propenal	с	[37]	251	3,4-methylenedioxycinnamaldehyde	s	[46]
214	3,4-dimethoxycinnamaldehyde	S	[46]	252	methyl trans-3-(3,4-dimethoxyphenyl)- 3-propenoate	s	[46]
215	3,4-methylenedioxycinnamyl alcohol	S	[46]	253	2-methoxyphenylacetone	c	[34]
216	isoeugenol	k	[41]	254	phenethyl (E)-3-[4-methoxyphenyl]-2- propenoate	c	[37]
217	kobusinol B	b,z	[1, 38]	255	trans-cinnamyl 3-phenylpro pionate	b,c	[17, 43]
218	caffeic acid	Z	[85]	256	(E)-cinnamyl-(E)-cinnamate	с	[17]
219	methyl cinnamate	8	[46]	257	1,2-dimethoxy-4-(1-E- propeny1)benzene	а	[15]
220	cis-2-methoxycinnamic acid	с	[34]	258	1,2-dimethoxy-4-(l-Z-propenyl)benze	а	[15]
221	linocinnamarin	с	[13]	259	rosarin	с	[13]
222	E-(3,4-dimethoxyphenyl)-2- propenal	а	[15]	260	2-[4-(3-hydroxypropyl)-2- methoxyphenoxy]-1,3-propanediol	c	[13]
223	sinapaldehyde	b,c	[28, 43]	261	E-(3,4-dimethoxyphenyl)-2-propenal	а	[15]
224	trans-ferulaldehyde	b	[43]	262	Cinnacassiol	с	[1]
225	trans-3,4,5-trimethoxycinnamic alcohol	с	[28]	263	dimethylmatairesinol	d	[48, 70]
226	4-allylcatechol	t	[45]	264	linocinnamarin	с	[13]
227	eugenol	d,j,m,n,u,s, z	[46, 84, 85, 89]	265	cinnacassin N	c	[17]
228	methyl-eugenol	s.t	[46, 90]	266	cinnacassin O	с	[17]
229	safrole	p	[36]	267	cinnacassin L	с	[17]
230	(7R, 8S)-syringoylglycerol	c	[37]	268	cinnamic aldehyde cyclic syringyl glycerol 1,3-acetal	b	[43]
231	(7S, 8S)-syringoylglycerol	с	[37]	269	cinnacassin K	c	[17]

Phenylpropanoids **194** and **195** were found to exhibit potent anti-inflammatory effects by inhibiting transcriptional activity of NF- κ B induced by LPS, and their IC₅₀ values were 43 and 31 μ M, respectively [78]. The phenylpropanoid (**250**) induced cell death in the HL-60 and A-549 cell lines with IC₅₀ values of 8.2 and 11.3 μ M, respectively. Compounds **194-197** showed moderate inhibitory effects with IC₅₀ values ranging from 20.5 to 65.6 μ M [76]. The anti-proliferation activities of **96** and **109** were evaluated against human HT29 and MCF-7 cancer cell lines, and their IC₅₀ values ranged from 3.3 to 25.8 μ M [51].

Cinnamaldehyde (194) has been demonstrated to show various activities and some *in vivo* experiments have been conducted. It was confirmed to exert *in vivo* anti-inflammatory effects and significantly reduced synovial inflammation in adjuvant arthritis rats due to suppressing IL-1 β through modulating succinate/HIF-1 α axis and inhibition of NLRP3 [79]. In another *in vivo* experiment, treatment with **194** was demonstrated to exhibit neuroprotective activity against subarachnoid hemorrhage-induced early brain injury through increasing the cross-sectional areas of the basilar artery and reducing the arterial wall thickness in rabbits [80]. Moreover, pretreatment with **194** significantly protected against and ameliorated intestinal ischemia/ reperfusion injuries by synergistic inhibition of NF- κ B and p53 in rats [81].

2.5. Alkaloids

Although *Cinnamomum* plants are rich in many types of constituents, alkaloids are not common in the genus and only 19 alkaloids (**270-288**) have been isolated up to now (Table 5 & Figure 5). These compounds include five piperidines (**270-274**), two pyrrolidines (**275**, **276**), nine amines (**277-285**) and three chlorophylls (**286-288**). The pyridine alkaloids (**270-274**) are all from *C. philippinense*. Compound **270** was first isolated from this species in 2012 and compound **272** was in 2015 [91, 92]. However, alkaloids from *Cinnamomum* didn't show significant biological activities according to existing activity tests.

No.	Compounds	Plants	s Ref.	No.	Compounds	Plant	s Ref.
270	2-(4'-hydroxypyridin-3'-yl)- acetic acid	q	[91]	280	cinnabutamine	b	[93]
271	corydaldine	q	[91]	281	N-cis-feruloyltyramine	r	[88]
272	Cinnapine	q	[92]	282	(E)-3-(4-hydroxy-3-methoxyphenyl)-N- phenthy-lacrylamide	c	[28]
273	glaziovine	q	[91]	283	N-trans-caffeoyl-5-hydroxytyramine	b	[93]
274	zenkerine	q	[91]	284	N-trans-feruloyl-5-methoxytyramine	b,c,r	[28, 39, 94]
275	3-glyceroylindole	с	[28]	285	N-trans-feruloyltyramine	b,r	[88, 93]
276	indole-3-carboxaldehyde	с	[28]	286	pheophytin b	b	[38]
277	Cinnaretamine	b,q,r	[52, 93, 94	4] 287	pheophytin a	b,s	[38, 53]
278	dihydroferuloyltyramine	r	[58]	288	aristophyll C	S	[53]
279	N-cis-feruloyl-5- methoxytyramine	b,r	[93, 94]				

Table 5. Alkaloids from Cinnamomum genus

Figure 5. The structures of alkaloids from Cinnamomum

2.6. Other Compounds

In addition to lignans, butanolides, flavonoids, phenylpropanoids and alkaloids, other compounds consist of 17 phenylethanols (**289-305**), 69 simple benzenoids (**306-374**) and 6 steroids (**375-380**). Compounds **292-296** are 4-hydroxy-3-methoxyphenethyl derivatives and all were obtained from *C. reticulatum* [39]. phenylethyl glycosides include compounds **300-304** and all are from *C. cassia* [32, 95]. Twelve dibenzocycloheptatrienes (**306-317**) have been obtained from *Cinnamomum* plants. Compounds **306-312** were first isolated from *C. subavenium* in 2012 [72]. Some benzenoids (**318-320**, **322**, **323** and **329**) are common in *Cinnamomum* species. Compounds **375-380** are steroids and **375-379** can be easily found in the genus *Cinnamomum*. These compounds are shown in Table 6.

No.	Compounds	Plants	Ref.	No.	Compounds	Plants	Ref.
289	phenylethyl alcohol	с	[34]	335	3,4,5-trimethoxyphenyl-1- O-β-D-glucoside	S	[83]
290	hydroxytyrosol	S	[72]	336	isovanillin	S	[46]
291	3,4-dimethoxyphenethyl alcohol	с	[34]	337	3,4-dihydroxybenzoate	с	[28]
292	4-hydroxy-3-methoxyphenethyl butyrate	r	[39]	338	ethyl 3,4- dihydroxybenzoate	с	[28]
293	4-hydroxy-3-methoxyphenethyl hexyrate	r	[39]	339	1,2-dimethoxy-4-(2- propenyl)benzen	a	[15]
294	4-hydroxy-3-methoxyphenethyl pentadecyrate	r	[39]	340	3,4- dimethoxybenzaldehyde	a	[15]
295	4-hydroxy-3-methoxyphenethyl stearate	r	[39]	341	ethyl 3,5-dihydroxy-4- nitrobenzoate	t	[90]
296	4-hydroxy-3-methoxyphenethyl heneicosyrate	r	[39]	342	leonuriside	с	[13]
297	4,4'-diacetyl-2,2'- dimethoxydiphenyl ether	n	[86]	343	methyl 3-methoxy-4-(β-D- allopyranosyloxy) benzoate	с	[13]
298	cinnamic alcohol	b,c	[43, 82]	344	gallic acid	Z	[85]
299	icariside DC	с	[82]	345	isotachioside	с	[95]
300	cinnacasolide A	с	[77]	346	3,4-dimethoxyphenol- β -D- apiofuranosyl(1 \rightarrow 6)-b-D- glucopyranoside	с	[13, 96]
301	2-phenylethyl-O-β-D- glucopyranoside	с	[95]	347	kelampayoside A	С	[96]
302	2-O-β-D-glucosyl-(1S)- phenylethylene glycol	с	[95]	348	glucosyringic acid	с	[31]
303	cinnamic aldehyde cyclic glycerol 1,3-acetal(9,2'-trans)	с	[32]	349	dihydrisosubamol	S	[72]
304	cinnamic aldehyde cyclic glycerol 1,3-acetal(9,2'-cis)	с	[32]	350	2,2',7a,7a',7b,7b'- hexamethyldiphenyl ether	S	[46]
305	galactitol 3'R,4'S-acetal	с	[31]	351	2-hydroxybenzaldehyde	с	[34]
306	Subavenoside A	S	[72]	352	ethyl ester	d	[42]
307	Subavenoside B	S	[72]	353	benzyl benzoate	C	[34]
308	Subavenoside C	S	[/2]	354	$(2P, 4S, 6P), 4 \in dibudrouv$	a	[48]
309	Subavenoside D	S	[72]	355	de-O-methyllasiodiplodin		[96]
310	Subavenoside E	S	[72]	356	8-methyl-anthraquinone (3R, 4R, 3'R,4'R)-6,6'-	d	[42]
311	Subavenoside F	S	[72]	357	dimethoxy-3,4,3',4'- tetrahydro-2H,2'H- [3,3']bichromenyl-4,4'-diol 2 3-dihydro-6 6-	d,p	[42, 71]
312	9,12-Di-O-methylsubamol	S	[72]	358	dimethylbenzo- [b][1,5]dioxocin-4(6H)-one	t	[65]
313	5'-hydroxy-5-hydroxy methyl- 4",5"-methylenedioxy-1,2,3,4- dibenzo-1,3,5-cycloheptatriene	b	[43]	359	cinnamophilin D	q	[97]
314	Subamol	S	[50, 87]	360	cinnacasolide C 3 4-dimethoxyphenol-β-D-	с	[77]
315	burmanol	b	[38]	361	apiofuranosyl($1 \rightarrow 6$)- β -D- glucopyranoside	с	[34]
316	tenuifolin	r,t	[45, 88]	362	3,4,5-trimethoxyphenol-β- D-apiofuranosyl-(1→6)-O- β-D-glucopyranoside 3-trimethoxy-4-	с	[2, 13]
317	reticuol	b,m,r	[98]	363	hydroxyphenol-β-D- apiofuranosyl (1→6)-β-D- glucopyranoside	с	[2, 13]
318	vanillin	b,c,d,k,s	[38, 41, 42]	364	cinnacassoside C	с	[13]
<u>319</u>	4-hydroxybenzaldehyde	c,k,p,r,s,t	[40, 45, 58]	365	cinnacasolide E	с	[2]

Table 6. Other compounds from Cinnamomum genus

			120 40 42		phenol-β-D-apiofuranosyl-		
320	protocatechuic acid	b,c,d,k,r,z	[29, 40, 42,	366	(1→6)-O-β-D-	с	[2, 32]
			00]		glucopyranoside		
321	benzoic acid	c,k	[40]	367	cinnacasolideb	с	[1]
			[20 5 0 6 1		phenylmethanol O- α -L-		
322	p-hydroxybenzoic acid	b,c,d,j,k,p,r,s	[30, 30, 01,	368	arabinofuranosyl $(1 \rightarrow 6)$ -	с	[95]
			00]		β-D-glucopyranoside		
			[20 50 61		phenylmethanol O-α-L-		
323	vanillic acid	b,c,k,r,s	[30, 30, 01,	369	arabinopyranosyl $(1\rightarrow 6)$ - β -	с	[95]
			00]		D-glucopyranoside		
324	benzaldehyde	с	[34]	370	cinnacassinol	с	[82]
225	varatraldahyda	C	[46]	271	1,4-diphenyl-1,4-	0	[17]
345	veratiatdenyde	8	[40]	5/1	butanedione	C	[1/]
326	protocatechualdehyde	d	[42]	372	evofolin B	с	[13, 17]
327	1,2,4-trihydroxybenzene	р	[71]	373	cinncassin B	с	[34]
328	benzene,1,3-dimethyl	с	[34]	374	cinncassin C	с	[34]
220	auringaldahuda	badka	[40, 41, 43,	275	B sitestarol	adikmnnatu	[27, 75,
349	synngaldenyde	0,c,u,ĸ,s	44]	515	p-sitosteror	c,u,j,ĸ,m,n,p,s,t,u	90, 99]
330	suringic acid	ret	[30 00 04]	376	stigmasterol	dkistu	[40, 44,
550	synngic acid	1,8,0	[39, 90, 94]	570	stigmasteroi	и,к,ј,ѕ,ι,и	89, 99]
331	myristicin	s t	[45 67 100]	377	daucosterol	dknst	[44, 70,
551	mynstem	5,1	[43, 07, 100]	511	dateosteror	и,к,р,з,г	82]
332	3,4-methylenedioxy-5-	s	[67 100]	378	stigmasteryl-3-O-β-D-	dks	[27, 41,
554	methoxycinnamyl alcohol	3	[07, 100]	570	glucoside	и,к,з	44]
333	myristicic acid	s	[100]	379	B-sitostenone	hikmnrst	[39, 45,
555	mynstele deld	5	[100]	517	p-sitostenone	0,1,1,11,11,1,5,0	46, 86]
334	3 -hydroxy- 4 , 5 -dinethoxyphenyl- β -	s	[83]	380	stigmasta-4 22-dien-3-one	k	[40]
554	D glucopyranoside	3	[00]	500	511511115tu +,22-0101-5-0110	ĸ	[40]

3. Conclusion

The medicinal value of the genus *Cinnamomum* has attracted much attention around the world and a great deal of phytochemical and biological investigations have been done. According to the findings, there have been many unique constituents isolated from *Cinnamomum* plants, with various novel skeletons and significant biological activities. The research on *Cinnamomum* species can provide abundant bioactive compounds and promote the further development and utilization of new drugs.

The *Cinnamomum* genes is rich in resources which contains approximately 250 species. However, only a few species have been studied, most of which are only given focus on the investigation of essential oils. Chemical research on the bioactive components from *Cinnamomum* plants have only focused on less than 20 species, such as *C. cassia, C. camphora, C. kotoense* and *C. subavenium*. Hence, the research range of species of the genus *Cinnamomum* need to be widened and the active ingredients and their pharmacological activities need to be further explored.

The compounds obtained from *Cinnamomum* show various significant activities, especially lignans and butanolides. The lignans from *Cinnamomum* have high content and have shown potent neuroprotective, anti-hyperlipidemic, anti-inflammatory, anticancer and other effects. However, most of the compounds only stays in the study of cell activity *in vitro* except sesamin, which was demonstrated to show various activities *in vivo* and *in vitro* and some clinical experiments have been conducted. Moreover, anti-tumor ingredients are mainly concentrated in butanolides. Nevertheless, though showing significant effects, most of the compounds have only been tested for *in vitro* activities. More *in vivo* experiments are needed to explore the mechanism of action and provide data for clinical trials.

Acknowledgments

This work was supported by the Open Project of the Key Laboratory of Biological Pesticide and Chemical Biology of the Ministry of Education (No. Keylab2019-05), Forestry Science and Technology

Promotion Demonstration Project of Central Finance (No.Min[2020]TG07) and Special Funds for Science and Technology Commissioners of Fujian Province (103 / KTP19108A and 103 / K1517070A).

ORCID D Meiting Wu: 0000-0001-5750-8209 Zhiyi Lin: 0000-0002-5898-7173 Bingbing Huang: 0000-0003-2923-8664 Kuo Xu: 0000-0002-0566-9884 Shuangquan Zou: 0000-0002-4125-7916 Lin Ni: 0000-0001-6118-6724 Yuxi Chen: 0000-0002-7826-4212

References

- [1] J. Zhao and J. S. Ma (2016). Phytochemicals and biological activities of the genus *Cinnamomum*, J. *Pharmacognosy Phytochem.* **4**, 27-34.
- [2] J. Zeng, Y. Xue, Y. Lai, G. Yao, Z. Luo, Y. Zhang and J. Zhang (2014). A new phenolic glycoside from the barks of *Cinnamomum cassia*, *Molecules* **19**, 17727-17734.
- [3] S. Kumar, R. Kumari and S. Mishra (2019). Pharmacological properties and their medicinal uses of *Cinnamomum*: a review, *J. Pharm. Pharmacol.* **71**, 1735-1761.
- [4] R. Singh and T. Jawaid (2012). Cinnamomum camphora (Kapur): Review, Pharmacognosy J. 4, 1-5.
- [5] D. R. A. Muhammad, E. Tuenter, G. D. Patria, K. Foubert, L. Pieters and K. Dewettinck (2021). Phytochemical composition and antioxidant activity of *Cinnamomum burmannii* Blume extracts and their potential application in white chocolate, *Food Chem.* **340**, 127983.
- [6] X. Yan, Y. Gao, S. Liu, G. Zhang, J. Zhao, D. Cheng, Z. Zeng, X. Gong, P. Yu and D. Gong (2021). Covalent modification by phenolic extract improves the structural properties and antioxidant activities of the protein isolate from *Cinnamonum camphora* seed kernel, *Food Chem.* **352**, 129377.
- [7] M. A. Alghuthaymi, A. M. Diab, A. F. Elzahy, K. E. Mazrou, A. A. Tayel, S. H. Moussa and L. Arru (2021). Green biosynthesized selenium nanoparticles by cinnamon extract and their antimicrobial activity and application as edible coatings with nano-chitosan, *J. Food Quality* **2021**, 1-10.
- [8] E. Tamilselvi, A. Karuppaiah, G. Shyamala, S. Shobana, P. Thangaraj, S. Hariharan and V. Sankar (2021). Exploring combined herbal extract-loaded phytoniosomes for antimalarial and antibacterial activity against methicillin-resistant Staphylococcus aureus, *3 Biotech* **11**, 177.
- [9] Y. Fan, J. Liu, J. Miao, X. Zhang, Y. Yan, L. Bai, J. Chang, Y. Wang, L. Wang, Y. Bian and H. Zhou (2021). Anti-inflammatory activity of the Tongmai Yangxin pill in the treatment of coronary heart disease is associated with estrogen receptor and NF-kappaB signaling pathway, *J. Ethnopharmacol.* 276, 114106.
- [10] N. Singh, A. S. Rao, A. Nandal, S. Kumar, S. S. Yadava, S. A. Ganaie and B. Narasimhan (2021). Phytochemical and pharmacological review of *Cinnamomum* verum J. Presl-a versatile spice used in food and nutrition, *Food Chem.* 338, 127773.
- [11] M. T. Wu, L. Ni, H. Lu, H. Y. Xu, S. Q. Zou and X. X. Zou (2020). Terpenoids and their biological activities from *Cinnamomum*: A review, *J. Chem.* **2020**, 1-14.
- [12] Y. Lai, T. Liu, R. Sa, X. Wei, Y. Xue, Z. Wu, Z. Luo, M. Xiang, Y. Zhang and G. Yao (2015). Neolignans with a rare 2-Oxaspiro[4.5]deca-6,9-dien-8-one motif from the stem bark of *Cinnamomum subavenium*, *J. Nat. Prod.* **78**, 1740-1744.
- [13] Q. Luo, S.-M. Wang, Q. Lu, J. Luo and Y.-X. Cheng (2013). Identification of compounds from the water soluble extract of *Cinnamonum cassia* barks and their inhibitory effects against high-glucose-induced mesangial cells, *Molecules* 18, 10930-10943.
- [14] T. H. Chen, Y. H. Huang, J. J. Lin, B. C. Liau, S. Y. Wang, Y. C. Wu and T. T. Jong (2010). cytotoxic lignan esters from *Cinnamomum osmophloeum*, *Planta Med.* **76**, 613-619.
- [15] N. M. Cuong, W. C. Taylor and T. V. Sung (2006). A new cyclobutane lignan from *Cinnamomum* balansae, Nat. Prod. Lett. **15**, 331-338.
- [16] C. Yang, X. Zhi and H. Xu (2016). Advances on semisynthesis, total synthesis, and structure-activity relationships of honokiol and magnolol derivatives, *Mini Rev. Med. Chem.* **16**, 404-426.
- [17] X. Liu, J. Fu, X. J. Yao, J. Yang, L. Liu, T. G. Xie, P. C. Jiang, Z. H. Jiang and G. Y. Zhu (2018). Phenolic constituents isolated from the twigs of *Cinnamomum cassia* and their potential neuroprotective effects, *J*.

Nat. Prod. 81, 1333-1342.

- [18] S. Ye, W. Wang, X. Chen and Y. Deng (2020). Sesamin promotes angiogenesis and accelerates wound healing in rats via alleviates TBHP-induced apoptosis in human umbilical vein endothelial cells, *Biosci. Biotechnol. Biochem.* 84, 887-897.
- [19] A. F. Majdalawieh, S. Dalibalta and S. M. Yousef (2020). Effects of sesamin on fatty acid and cholesterol metabolism, macrophage cholesterol homeostasis and serum lipid profile: A comprehensive review, *Eur. J. Pharmacol.* 885, 173417.
- [20] T. H. Pham, S. W. Jin, G. H. Lee, J. S. Park, J. Y. Kim, T. N. Thai, E. H. Han and H. G. Jeong (2020). Sesamin induces endothelial nitric oxide synthase activation via transient receptor potential vanilloid type 1, J. Agric. Food Chem. 68, 3474-3484.
- [21] M. B. Sayhan, S. Oguz, O. Salt, N. Can, T. Ozgurtas and T. D. Yalta (2019). Sesamin ameliorates mucosal tissue injury of mesenteric ischemia and reperfusion in an experimental rat model, *Arch. Med. Sci.* 15, 1582-1588.
- [22] H. Ye, L. Sun, J. Li, Y. Wang, J. Bai, L. Wu, Q. Han, Z. Yang and L. Li (2020). Sesamin attenuates carrageenan-induced lung inflammation through upregulation of A20 and TAX1BP1 in rats, *Int. Immunopharmacol.* **88**, 107009.
- [23] B. Helli, M. M. Shahi, K. Mowla, M. T. Jalali and H. K. Haghighian (2019). A randomized, triple-blind, placebo-controlled clinical trial, evaluating the sesamin supplement effects on proteolytic enzymes, inflammatory markers, and clinical indices in women with rheumatoid arthritis, *Phytother. Res.* **33**, 2421-2428.
- [24] M. C. Barbalace, L. Zallocco, D. Beghelli, M. Ronci, S. Scortichini, M. Digiacomo, M. Macchia, M. R. Mazzoni, D. Fiorini, A. Lucacchini, S. Hrelia, L. Giusti and C. Angeloni (2021). Antioxidant and neuroprotective activity of extra virgin olive oil extracts obtained from quercetano cultivar trees grown in different areas of the tuscany region (Italy), *Antioxidants (Basel)* 10, 10030421.
- [25] S. He, K. W. Zeng, Y. Jiang and P. F. Tu (2016). Nitric oxide inhibitory constituents from the barks of *Cinnamomum cassia*, *Fitoterapia* **112**, 153-160.
- [26] F. Zhao, L. Chen, C. Bi, M. Zhang, W. Jiao and X. Yao (2013). *In vitro* anti-inflammatory effect of picrasmalignan A by the inhibition of iNOS and COX2 expression in LPS activated macrophage RAW 264.7 cells, *Mol. Med. Rep.* 8, 1575-1579.
- [27] T. J. Hsieha, C. H. Chenb, W. L. Loa and C. Y. Chena (2006). Lignans from the stem of *Cinnamomum camphora*, *Nat. Prod. Commun.* **1**,21-25.
- [28] X. Liu, J. Yang, J. Fu, T. G. Xie, P. C. Jiang, Z. H. Jiang and G. Y. Zhu (2018). Phytochemical and chemotaxonomic studies on the twigs of *Cinnamomum cassia* (Lauraceae), *Biochem. Systemat. Ecol.* 81, 45-48.
- [29] G. K. Jayaprakasha, M. O. Kameyama, H. Ono, M. Yoshida and L. J. Rao (2006). Phenolic constituents in the fruits of *Cinnamomum zeylanicum* and their antioxidant activity, *J. Agric. Food Chem.* **54**, 1672-1679.
- [30] J. F. Zeng, H. C. Zhu, J. W. Lu, L. Z. Hu, J. C. Song and Y. H. Zhang (2017). Two new geranylphenylacetate glycosides from the barks of *Cinnamomum cassia*, *Nat. Prod. Res.* **31**, 1812-1818.
- [31] C. Liu, S.-M. Zhong, R.-Y. Chen, Y. Wu and X.-J. Zhu (2009). Two new compounds from the dried tender stems of *Cinnamomum cassia*, *J. Asian Nat. Prod. Res.* **11**, 845-849.
- [32] M. Miyamura, T. Nohara, T. Tomimatsu and I. Nishioka (1983). Seven aromatic compounds from bark of *Cinnamomum cassia*, *Phytochemistry* **22**, 215-218.
- [33] Z. Z. Wang, T. J. Ling, L. Zhang and G. H. Bao (2014). A Study on the chemical constituents in the leaves of *Cinnamomum camphora*, *Nat. Prod. Res. Dev.* **26**, 860-863.
- [34] C. Zhang, L. Fan, S. Fan, J. Wang, T. Luo, Y. Tang, Z. Chen and L. Yu (2019). *Cinnamomum cassia* Presl: A review of its traditional uses, phytochemistry, pharmacology and toxicology, *Molecules* 24, 03473.
- [35] E. R. S. Nkanwen, M. D. Awouafack, J. J. K. Bankeu, H. K. Wabo, S. A. A. Mustafa, M. S. Ali, M. Lamshoft, M. I. Choudhary, M. Spiteller and P. Tane (2013). Constituents from the stem bark of *Cinnamomum zeylanicum* Welw. (Lauraceae) and their inhibitory activity toward Plasmodium falciparum enoyl-ACP reductase enzyme, *Rec. Nat. Prod.* 7, 296-301.
- [36] M. Adfa, R. Rahmad, M. Ninomiya, S. S. Yudha, K. Tanaka and M. Koketsu (2015). Antileukemic activity of lignans and phenylpropanoids of *Cinnamomum parthenoxylon*, *Bioorg. Med. Chem. Lett.* 26, 761-764.
- [37] Q. L. Hu. (2018) The discovery of components and research on bioactivities of *Cinnamomum cassia* and *Carpesium abrotanoides*, Lanzhou University, Lanzhou.
- [38] C.-Y. Chen, Z.-L. Hong, W.-L. Yang, M.-H. Wu, J.-C. Huang and J.-Y. Lee (2012). A novel homosesquiterpenoid from the stems of *Cinnamomum burmanii*, *Nat. Prod. Res.* **26**, 1218-1223.

- [39] I. Lin, Jr., W.-L. Lo, Y.-C. Chia, L.-Y. Huang, T.-M. Cham, W.-S. Tseng, Y.-T. Yeh, H.-C. Yeh, Y.-D. Wang and C.-Y. Chen (2010). Isolation of new esters from the stems of *Cinnamomum reticulatum* Hay, *Nat. Prod. Res.* **24**, 775-780.
- [40] F. C. Chen, C. F. Peng, I. L. Tsai and I. S. Chen (2005). Antitubercular constituents from the stem wood of *Cinnamomum kotoense*, *J. Nat. Prod.* **68**, 1318-1323.
- [41] C. C. H., L. W. L., L. Y. C. and C. C. Y. Chemical and cytotoxic constituents from the leaves of *Cinnamomum kotoense*, J. Nat. Prod. **69**, 927-933.
- [42] Y. R. Li, C. S. Fu, W. J. Yang, X. L. Wang, D. Feng, X. N. Wang, D. M. Ren, H. X. Lou and T. Shen (2018). Investigation of constituents from *Cinnamonum camphora* (L.) J. Presl and evaluation of their anti-inflammatory properties in lipopolysaccharide-stimulated RAW 264.7 macrophages, *J. Ethnopharmacol.* 221, 37-47.
- [43] S. Subehan, S. Kadota and Y. Tezuka (2008). In vitro mechanism-based inactivation of cytochrome P450 3A4 by a new constituent of *Cinnamomum burmani*, *Planta Med.* **74**, 1474-1480.
- [44] C. Y. Chen, C. H. Chen, C. H. Wong, Y. W. Liu, Y. S. Lin, Y. D. Wang and Y. R. Hsui (2007). Cytotoxic constituents of the stems of *Cinnamomum subawenium*, *J. Nat. Prod.* **70**, 103-106.
- [45] R. J. Lin, M. J. Cheng, J. C. Huang, W. L. Lo, Y. T. Yeh, C. M. Yen, C. M. Lu and C. Y. Chen (2009). Cytotoxic compounds from the stems of *Cinnamomum tenuifolium*, *J. Nat. Prod.* **72**, 1816-1824
- [46] G. C. Huang, C. L. Kao, H. C. Yeh, W. J. Li, H. T. Li and C. Y. Chen (2018). A new diphenyl ether from *Cinnamomum subavenium, Chem. Nat. Compd.* **54**, 869-871.
- [47] C. Y. Chen, C. L. Lin, C. L. Kao, C. T. Chen and H. T. Li (2019). Chemical Constituents of the Leaves of *Cinnamomum insulari-montanum, Chem. Nat. Compd.* **55**, 922-923.
- [48] C. Sun, X. Tang, J. Zhou and H. Wu (2014). Study on chemical constituents of *Cinnamomum camphora* leaves, *Nat. Prod. Res. Dev.* **26**, 1793-1796.
- [49] L. T. Yuan, C. L. Kao, C. T. Chen, H. T. Li and C. Y. Chen (2017). A New lignan from *Cinnamomum burmanii, Chem. Nat. Compd.* **53**, 623-625.
- [50] C. H. Lee, C. N. Kuo, H. L. Chen and C. Y. Chen (2013). Review on pharmacological activities of *Cinnamomum subavenium*, *Nat. Prod. Res.* **27**, 988-991.
- [51] S. S. Yang, W. C. Hou, L. W. Huang and T. H. Lee (2006). A new γ-lactone from the leaves of *Cinnamomum kotoense*, *Nat. Prod. Res.* 20, 1246-1250.
- [52] H.-M. D. Wang, C.-Y. Chen and P.-F. Wu (2014). Isophilippinolide a arrests cell cycle progression and induces apoptosis for anticancer inhibitory agents in human melanoma cells, *J. Agr. Food Chem.* **62**, 1057-1065.
- [53] S. Y. Kuo, T. J. Hsieh, Y. D. Wang, W. L. Lo, Y. R. Hsui and C. Y. Chen (2008). Cytotoxic constituents from the leaves of *Cinnamomum subavenium*, *Chem. Pharm. Bull (Tokyo)* **56**, 97-101.
- [54] H. M. Wang, C. Y. Chen and Z. H. Wen (2011). Identifying melanogenesis inhibitors from *Cinnamomum subavenium* with in vitro and *in vivo* screening systems by targeting the human tyrosinase, *Exp. Dermatol.* 20, 242-248.
- [55] C. H. Liu, C. Y. Chen, A. M. Huang and J. H. Li (2011). Subamolide A, a component isolated from *Cinnamomum subavenium*, induces apoptosis mediated by mitochondria-dependent, p53 and ERK1/2 pathways in human urothelial carcinoma cell line NTUB1, *J. Ethnopharmacol.* 137, 503-511.
- [56] J. Y. Hung, C. W. Wen, Y. L. Hsu, E. S. Lin, M. S. Huang, C. Y. Chen and P. L. Kuo (2013). Subamolide a induces mitotic catastrophe accompanied by apoptosis in human lung cancer cells, *Evid. Based. Complement Alternat. Med.* **2013**, 828143.
- [57] H. Fuchino, A. Yazawa, F. Kiuchi, N. Kawahara, Y. Takahashi and M. Satake (2015). Novel monoterpene lactones from *Cinnamomum inunctum*, *Chem. Pharm. Bull* **63**, 833-836.
- [58] I. J. Lin, H. C. Yeh, T. M. Cham and C. Y. Chen (2011). A new butanolide from the leaves of *Cinnamomum reticulatum, Chem. Nat. Compd.* **47**, 43-45.
- [59] Z. Li, Z. Cai, S. Qian and M. Chen (2017). A new lactone from the twigs of *Cinnamomum cassia*, *Chem. Nat. Compd.* **53**, 234-236.
- [60] C. Y. Chen (2006). Butanolides from the stem of *Cinnamomum kotoense*, *Nat. Prod. Commun.* **1**, 453-455.
- [61] A. Pardede, M. Adfa, A. Juliari Kusnanda, M. Ninomiya and M. Koketsu (2017). Flavonoid rutinosides from *Cinnamomum parthenoxylon* leaves and their hepatoprotective and antioxidant activity, *Med. Chem. Res.* **26**, 2074-2079.
- [62] M.-T. Chua, Y.-T. Tung and S.-T. Chang (2008). Antioxidant activities of ethanolic extracts from the twigs of *Cinnamomum osmophloeum*, *Bioresource Technol.* **99**, 1918-1925.
- [63] C.-M. Liu (2015). The antioxidation and antiproliferation activity of new flavonoids from the leaves and stems of *Cinnamomum reticulatum* Hayate, *Med. Chem.* **5** (2), 064-066.
- [64] S. Devi and V. Kumar (2020). Comprehensive structural analysis of cis- and trans-tiliroside and quercetrin from Malvastrum coromandelianum and their antioxidant activities, *Arab. J. Chem.* **13**, 1720-

1730.

- [65] H. L. Chen, S. Y. Kuo, Y. P. Li, Y. F. Kang, Y. T. Yeh, J. C. Huang and C. Y. Chen (2012). A new benzodioxocinone from the leaves of *Cinnamomum tenuifolium*, *Nat. Prod. Res.* **26**, 1881-1886.
- [66] S.-H. Fang, Y. K. Rao and Y.-M. Tzeng (2005). Inhibitory effects of flavonol glycosides from *Cinnamomum osmophloeum* on inflammatory mediators in LPS/IFN-γ-activated murine macrophages, *Bioorg. Med. Chem.* 13, 2381-2388.
- [67] C. Y. Chen, H. M. Wang, S. H. Chung, W. L. Lo, W. L. Yang and S. C. Yang (2010). Chemical constituents from the roots of *Cinnamomum subavenium*, *Chem. Nat. Compd.* **46**, 474-477.
- [68] Y. Z. Liu, Y. G. Cao, J. Q. Ye, W. G. Wang, K. J. Song, X. L. Wang, C. H. Wang, R. T. Li and X. M. Deng (2010). Immunomodulatory effects of proanthocyanidin A-1 derived in vitro from *Rhododendron spiciferum*, *Fitoterapia* **81**, 108-114.
- [69] K. B. Killday, M. H. Davey, J. A. Glinski, P. Duan, R. Veluri, G. Proni, F. J. Daugherty and M. S. Tempesta (2011). Bioactive a-type proanthocyanidins from *Cinnamomum cassia*, J. Nat. Prod. 74, 1833-1841.
- [70] L. Wu, W. Xiong, J.-W. Hu, J. Wu, Z.-J. Li, Y. Gao, D. Liu, Y. Liu, W. Liu, M. Liang, C.-L. Si and Y.-S. Bae (2019). Secondary metabolites from the twigs of *Cinnamomum camphora*, *Chem. Nat. Compd.* 55, 345-347.
- [71] X. Wei, G.-H. Li, X.-L. Wang, J.-X. He, X.-N. Wang, D.-M. Ren, H.-X. Lou and T. Shen (2017). Chemical constituents from the leaves of *Cinnamomum parthenoxylon* (Jack) Meisn. (Lauraceae), *Biochem. Syst. Ecol.* **70**, 95-98.
- [72] H.-C. Lin and S.-S. Lee (2012). Dibenzocycloheptanoids from the Leaves of *Cinnamomum subavenium*, *J. Nat. Prod.* **75**, 1735-1743.
- [73] R. J. Zhong, L. Y. Wu, W. Xiong, E. L. Xie, G. P. Zhou and D. M. Zhang (2011). A new 3-(3,4-methylenedioxyphenyl)-propane-1,2-diol glycoside from the roots of *Cinnamomum camphora*, *Chinese Chem. Lett.* **22**, 954–956.
- [74] C. Y. Chen, Y. D. Wang and C. J. Wang (2011). Chemical constituents from the leaves of *Cinnamomum philippinense*, *Chem. Nat. Compd.* **46**, 941-942.
- [75] T. J. Hsieh, S. F. Hsieh and C. Y. Chen (2010). Chemical constituents from the stems of *Cinnamomum insulari-montanum*, *Chem. Nat. Compd.* **46**, 99-100.
- [76] T. M. Ngoca, N. X. Nhiemb, N. M. Khoia, D. CaoSonc, T. V. Hungc and P. V. Kiem (2014). A new coumarin and cytotoxic activities of constituents from *Cinnamomum cassia*, *Nat. Prod. Commun.* 9, 487-488.
- [77] T. M. Ngoc, N. M. Khoi, D. T. Ha, N. X. Nhiem, B. H. Tai, D. V. Don, H. V. Luong, D. C. Son and K. Bae (2012). Xanthine oxidase inhibitory activity of constituents of *Cinnamonum cassia* twigs, *Bioorg. Med. Chem. Lett.* 22, 4625–4628.
- [78] A. M. Reddy, J. H. Seo, S. Y. Ryu, Y. S. Kim, Y. S. Kim, K. R. Min and Y. Kim (2004). Cinnamaldehyde and 2-methoxycinnamaldehyde as NF-κB inhibitors from *Cinnamomum cassia*, *Planta Med.* **70**, 823-827.
- [79] P. Liu, J. Wang, W. Wen, T. Pan, H. Chen, Y. Fu, F. Wang, J. H. Huang and S. Xu (2020). Cinnamaldehyde suppresses NLRP3 derived IL-1beta via activating succinate/HIF-1 in rheumatoid arthritis rats, *Int. Immunopharmacol.* 84, 106570.
- [80] B. Gurer, H. Kertmen, P. Kuru Bektasoglu, O. C. Ozturk, H. Bozkurt, A. Karakoc, A. T. Arikok and E. Celikoglu (2019). The effects of cinnamaldehyde on early brain injury and cerebral vasospasm following experimental subarachnoid hemorrhage in rabbits, *Metab. Brain Dis.* **34**, 1737-1746.
- [81] M. Almoiliqy, J. Wen, B. Xu, Y. C. Sun, M. Q. Lian, Y. L. Li, E. Qaed, M. Al-Azab, D. P. Chen, A. Shopit, L. Wang, P. Y. Sun and Y. Lin (2020). Cinnamaldehyde protects against rat intestinal ischemia/reperfusion injuries by synergistic inhibition of NF-kappaB and p53, *Acta. Pharmacol. Sin.* 41, 1208-1222.
- [82] T. M. Ngoc, I. S. Lee, D. T. Ha, H. J. Kim, B. S. Min and K. H. Bae (2009). Tyrosinase-inhibitory constituents from the twigs of *Cinnamomum cassia*, *J. Nat. Prod.* **72**, 1205–1208.
- [83] X. Hao, J. Chen, Y. Lai, M. Sang, G. Yao, Y. Xue, Z. Luo, G. Zhang and Y. Zhang (2015). Chemical constituents from leaves of *Cinnamomum subavenium*, *Biochem. Syst. Ecol.* **61**, 156-160.
- [84] W.-L. Yang, Y. C. Guo, H. H. Lin, H. C. Hong and C. Y. Chen (2012). Chemical constituents from the twigs of *Cinnamomum macrostemon*, *Chem. Nat. Compd.* **47**, 1030-1031.
- [85] D. Shahwar, M. A. Raza, R. Shafiq Ur, M. A. Abbasi and A. Ur Rahman (2012). An investigation of phenolic compounds from plant sources as trypsin inhibitors, *Nat. Prod. Res.* **26**, 1087-1093.
- [86] C. L. Lin, M. H. Perng, W. J. Li, H. T. Li and C. Y. Chen (2018). Chemical constituents of the roots of *Cinnamomum randaiense, Chem. Nat. Compd.* **54**, 628-630.

- [87] C. Y. Chen and Y. D. Wang (2011). Wang A novel sequiterpenoid from the leaves of *Cinnamomum subavenium*, *Chem. Nat. Compd.* **47**, 215-217.
- [88] C. Y. Chen (2011). Chemical constituents from the roots of *Cinnamomum reticulatum*, *Chem. Nat. Compd.* **47**, 306-308.
- [89] D. L. Espineli, E. M. G. Agoo, C. C. Shen and C. Y. Ragasa (2014). Chemical constituents of *Cinnamomum trichophyllum, Chem. Nat. Compd.* **50**, 389-390.
- [90] M. J. Cheng, Y. T. Yeh, C. J. Wang, C. Yi and C. Y. Chen (2011). Isolation of a nitrobenzoate from the leaves of *Cinnamomum tenuifolium*, *Nat. Prod. Res.* **25**, 118-122.
- [91] H. T. Li, W. J. Li, H. M. Wu and C. Y. Chen (2012). Alkaloids from *Cinnamomum philippinense*, *Nat. Prod. Commun.***7**, 1581-1582.
- [92] C. L. Kao, C. L. Cho, H. M. Wu, C. T. Huang, W. J. Li, C. T. Li, H. T. Li, C. L. Lin and C. Y. Chen (2015). Cinnapine, a new pyridine alkaloid from *Cinnamomum philippinense*, *Chem. Nat. Compd.* 51, 736-738.
- [93] Z. L. Hong, J. C. Huang, S. Y. Kuo and C. Y. Chen (2011). Amides from the stems of *Cinnamomum burmannii*, *Nat. Prod. Commun.* **6** (9), 1297-1298.
- [94] C.-Y. Chen and H. C. Yeh (2011). A new amide from the stems of *Cinnamomum reticulatum* Hay, *Nat. Prod. Res.* **25**, 26-30.
- [95] Y. Guoruoluo, H. Zhou, W. Wang, J. Zhou, H. A. Aisa and G. Yao (2018). Chemical constituents from the immature buds of *Cinnamomum cassia* (Lauraceae), *Biochem. Syst. Ecol.* **78**, 102-105.
- [96] S. He, Y. Jiang and P. F. Tu (2016). Three new compounds from *Cinnamomum cassia*, J. Asian Nat. *Prod. Res.* **18**, 134-140.
- [97] M. J. Cheng, Y. T. Yeh, C. J. Wang and C. Y. Chen (2012). A new 3,4-dihydronaphthalen-1(2H)-one from the leaves of *Cinnamomum philippinense*, *Nat. Prod. Res.* **26**, 1433-1435.
- [98] H. L. Chen, W. L. Yang, Y. P. Li, Y. F. Kang and H. M. Wu (2012). A novel homosesquiterpene from *Cinnamomum macrostemon* Hayata, *J. Med. Med. Sci.* **3**, 90-92.
- [99] C. Y. Chen and Z. L. Hong (2011). Chemical constituents from the fruits of *Cinnamomum kotoense*, *Chem. Nat. Compd.* **47**, 450-451.
- [100] C.-Y. Chen and Y.-D. Wang (2011). Norcadinane sesquiterpene from the roots of *Cinnamomum subavenium*, *Chem. Nat. Compd.* **47**, 461-462.

A C G publications © 2021 ACG Publications