

Rec. Nat. Prod. 16:6 (2022) 538-549

records of natural products

A Comprehensive Review on Traditional Uses, Chemical Constituents, and Diverse Pharmacological Importance of the Genus *Breynia*

Malik Saadullah ¹, Muhammad Asif ², Sania Arif ¹ and Bisma Kanwal ¹

¹Department of Pharmaceutical Chemistry, Faculty of Pharmaceutical Sciences, Government College University, Faisalabad, Pakistan

² Department of Pharmacology, The Islamia University of Bahawalpur, Pakistan

(Received December 03, 2021; Revised March 20, 2022; Accepted March 23, 2022)

Abstract: The genus *Breynia* (family Phyllanthaceae) is widely distributed in Australia, Vietnam, Malaysia, and some regions of India. Traditionally, species of this genus were used to treat various disorders, like skin diseases, pain, cough, tonsillitis, dysentery, and headache. Various studies about this genus are available, but reviews highlighting its pharmacology and phytochemistry are inadequate. This review highlights the pharmacology, phytochemistry, and chemotaxonomic classification of the phytochemicals of ten species of the genus *Breynia*. About 90 compounds have been isolated from *Breynia* species, including glycosides, flavonoids, terpenoids, steroids, alkaloids, lignans, phenolic compounds, and catechins. Their structure and presence in each species are presented in tabular form. In pharmacological medicines, the crude extracts and metabolites of the genus *Breynia* have been found to exhibit diverse biological activity, including, antioxidant, antimicrobial, antidiabetic, anti-inflammatory, anticancer, antiviral, and activity against various blood disorders. Few isolated compounds show enzyme inhibitory activity (tyrosinase, xanthine oxidase, and elastase inhibition).

Keywords: *Breynia*; Phyllanthaceae; chemotaxonomic classification; phenolic compounds; glycosides; pharmacology. © 2022 ACG Publications. All rights reserved.

1. Introduction

Natural products which are derived from plants play an important role in the maintenance of human health. In ancient times, plants were the main source of medicines. Plant-derived medicinal products have negligible side effects on human health. Medicinal plants are the main source of various bioactive compounds having diverse therapeutic properties. The therapeutic effects interrelated with medicinal plants include antiviral, anti-inflammatory, antidiabetic, antitumor, antioxidant, and analgesic properties [1].

Breynia is a plant genus in the family Phyllanthaceae which is one of the five segregates of Euphorbiaceae sensu lato, an angiosperm phylogeny group. This family contains about 2000 species in 59 presently accepted genera, 10 tribes, and 2 subfamilies [2]. Most of the plants in the Phyllanthaceae family are trees, shrubs, or herbs, some of them are climbers, and one species is aquatic [3]. The

The article was published by ACG Publications

http://www.acgpubs.org/journal/records-of-natural-products November-December 2022 EISSN:1307-6167 DOI: http://doi.org/10.25135/rnp.314.2112.2280

Available online: April 12, 2022

^{*} Corresponding author: E-Mail: arifsania456@gmail.com, Phone: +92 318 7021480.

Saadullah et.al., Rec. Nat. Prod. (2022) 16:6 538-549

species of the genus *Breynia* are mostly found in Australia, Vietnam, Malaysia, and some regions of India. It is not a native plant of Pakistan but four species are grown in Pakistan, in Karachi and Lahore.

The genus *Breynia* is comprised of evergreen trees and shrubs. They grow to a small size, known as shrubs, and have colourful leaves; mostly their leaves are flattened. The flowers are in small size. *Breynia* species are monoecious. Their leaves are simple, distichous and petioles are short in size. In flowers, the calyx is six-lobed, petals and discs are absent. The fruit is a small fleshy capsule, about 5 mm wide, usually red in colour. The seed is triangular in transverse section, black in colour, and covered with a thin aril. This plant is mostly grown in warmer areas; the soil must be humus-rich. Fifty species of *Breynia* are known in different regions, of which 30 species are found in the above-mentioned regions [4]. The most common species of *Breynia* are *B. racemosa*, *B. vitis-idaea*, *B. disticha*, *B. discigera*, *B. fruticosa*, *B. rostrata*, *B. retusa*, *B. androgyna* (*Sauropus androgynus*), and *B. officinalis*.

2. Traditional Uses

The plants in this genus are mostly used for medicinal purposes, especially their leaves [5]. Traditionally, the leaves of *B. discigera* are used for the treatment of kidney disorders and *B. racemosa* with turmeric is used for various skin disorders [6]. In the Philippines, the bark of *B. vitisidaea* is used as an astringent to stop haemorrhages, and its dried leaves as a treatment for tonsillitis and skin diseases. The leaves of *B. vestita* are rubbed over the body for the treatment of malaria and other fevers, also being used for the treatment of dysentery, cough, and headache. In various regions, their young leaves are used as a vegetable [7]. *B. glauca* and *B. fruticosa* are used as anti-inflammatory agents [7, 8]. Traditionally in Indonesia, *B. androgyna* is used as a lactation enhancer, due to the presence of vitamin A (vitamin A synthesizes retinol, and retinol reacts with fatty acids (triggering release of the hormone prolactin)), increasing the production of breast milk [9]. *B. oblongifolia* is used as an antiviral [10], and many other species are used for different medicinal purposes.

3. Phytochemical Studies

Phytochemicals are biologically active compounds, found naturally in plants, protect plants from environmental hazards, and also provide colour, aroma, and flavour to plants. These chemicals provide different health benefits to humans when their dietary intake is significant. Phytochemicals are majorly used for disease prevention, as anticancer, anti-inflammatory, antimicrobial, and antidiabetic agents, and work as various immunity-potentiating agents. They are accumulated in different parts of plants, such as in roots, stems, leaves, flowers, seeds, and fruits [11]. Compounds **1–90** were isolated from species of the genus *Breynia* (Table 1, Table S1 in supporting information). Commonly, glycosides are the major constituents of this genus. Many alkaloids, terpenoids, lignin, steroids, catechins, aromatic ketones, nucleosides, tannins, and aromatic heterocyclic compounds are reported in this genus.

Glycosidal compounds isolated from genus *Breynia*: among the reported compounds, 13–80 are glycosides. Flavonoidal, sulphur-containing spiroacetal, ionol, phenolic, megastigmane, isoprenoid, spiroketal, and simple glycosides were isolated from *B. fruticosa* (whole plant extract), *B. officinalis* (leaf extract), *B. rostrata* (leaf extract), *B. glauca* (leaf extract), *B. androgyna* (leaf extract), and *B. retusa* (leaf extract) (Table 1).

Flavonoids, including compounds **10–12**, were isolated from *B. retusa* (leaf and fruit extract), *B. glauca* (leaf extract), and *B. fruticosa* (leaf extract).

Terpenoids, including compounds 87–89, were isolated from *B. rostrata* (leaf extract) and *B. fruticosa* (leaf extract).

Alkaloids: there are a total of five alkaloids, separated from *B. coronata* (leaf extract), numbered 1-5; three of them (2-4) are the structural derivative of **1**. In this genus, alkaloids are reported in less quantity as compared to glycosides and flavonoids.

Catechins, isolated from *B. retusa* (leaf, fruit, and stem bark extract) are numbered 7-9 in Table 1.

Aromatic ketones: only one aromatic ketone compound was isolated from *B. fruticosa* (leaf extract), numbered 6 in Table 1.

Lignins (phenylpropane units joined together with β , β -bonds) and neolignan (phenylpropane units linked together with other carbon–carbon bonds) are phenolic compounds. Among the reported compounds, **81** was lignin, and **82–84** were neolignans, isolated from *B. fruticosa* (leaf extract), *B. rostrata* (leaf extract), and *B. androgyna* (leaf extract).

Steroidal compounds: isolated from *B. androgyna* (leaf extract), numbered **85** and **86** in Table 1.

Tannins: only one compound, numbered 90 was isolated from *B. rostrata* (leaf extract).

All these mentioned compounds were isolated and detected by ¹H-NMR, ¹³C-NMR, MS, LC-MS, HPLC, LC-DAD-MSn, a two-column LC method, FTIR, and IR spectroscopy. The extraction of roots, leaves, bark, stem, flowers, and seeds of species of the genus *Breynia* was done with ethanol (EtOH), methanol (MeOH), chloroform (CHCl₃), n-butanol (n-BuOH), ethyl acetate (EtOAc), petrol, dichloromethane (CH₂Cl₂), dimethyl sulphoxide (DMSO), and water (H₂O).

This mentioned data revealed the chemotaxonomic classification of phytochemicals present in ten species of the genus *Breynia*.

3. Pharmacological Activity

The pharmacological or biological activity of a plant extract describes its beneficial or adverse effect on human health [42]. The genus *Breynia* has medicinal importance due to the presence of secondary metabolites such as glycosides, terpenoids, alkaloids, lignins, steroids, nucleosides, tannins, and phenolic compounds. The biological activities of various species of the genus *Breynia* are highlighted below.

Antimicrobial activity is activity that inhibits the growth of microbes or may destroy microbial colonies. EtOH extract of *B. retusa* leaves shows effective results against The Salmonella Typhimurium, E. coli, and Enterobacter aerogenes and is less effective against Micrococcus luteus. Also, it is more effective against Trichophyton rubrum, Aspergillus niger, and Penicillium sp. and less effective against Cryptococcus sp. fungi. This plant is rich in tannins (best for wound healing). This activity is highly sensitive with a high concentration [43]. Petrol, CH_2Cl_2 , BuOH, EtOAc, and MeOH extracts of B. cernua stem, leaves, root bark, and heartwood show broadspectrum antimicrobial activity but MeOH extract of root bark shows the best activity against microbes, and CH₂Cl₂ extract of stem bark shows good activity against all fungi [44]. MeOH and EtOH extracts of B. androgyna leaves show significant antibacterial activity against Bacillus cereus, Proteus vulgaris, Staphylococcus aureus, E. coli, Klebsiella pneumonia, and Gram-positive bacteria. They also show activity against Candida albicans and Aspergillus flavus fungi [45, 46, 47]. Tannins are responsible for this activity. MeOH extract of B. disticha shows activity against S. aureus and E. coli, with a 12 mm zone of inhibition. It also shows activity against S. cerevisiae fungi with an 8.0 mm zone of inhibition [48]. EtOH extract of B. nivosa shows activity against B. subtilis, S. aureus, and Salmonella Typhi. No activity is recorded against C. albicans, A. fumigatus, or E. coli [49].

An *in vitro* study on 96% EtOH extracts of *B. cernua* and three fractions, namely ethyl acetate, n-hexane, and water fractions, showed cytotoxicity against MCF-7 breast cancer cells with IC_{50} values of 245.841, 562.57, 165.65, and 713.78 ppm, respectively [50]. EtOH and MeOH extract (250–2500 mg/L) of *B. androgyna* possesses a cytotoxic effect on human mesenchymal stem cells, showing less cytotoxicity at higher concentrations, and more cytotoxic activity at lower concentrations [9].

 Table 1. Isolated compounds of genus Breynia

Compound	Compound Name	Sources	Ref.
No	Alleologide		
1	Alkaloids Securinine	D concernents (loof outpost)	[10]
1	Allosecurinines	<i>B. coronata</i> (leaf extract)	[12]
2 3	Virosecurinine	<i>B. coronata</i> (leaf extract)	[12]
	Virosecurinine	<i>B. coronata</i> (leaf extract)	[12]
4 5		<i>B. coronata</i> (leaf extract)	[12]
3	Ent-Phyllanthidine Aromatic ketones	B. coronata (leaf extract)	[12]
6		P functioned (loof outroot)	[20]
6	2, 4-dihydroxy-6-methoxy-3-methyl-acetophenone Catechins	B. fruticosa (leaf extract)	[30]
7	Epicatechin	B. retusa (leaf, fruit, and stem	[13]
1	Epicateenin	bark extract)	[15]
8	Epicatechin-7-O-sulphate	<i>B. retusa</i> (stem bark extract)	[13]
9	Procyanidin B2	<i>B. retusa</i> (stell bark extract) <i>B. retusa</i> (leaf and fruit	[13]
,	Flavonoids	extract)	[15]
10	Kaempferol	<i>B. glauca</i> (leaf extract), <i>B.</i>	[7]
10	Kachipicioi	<i>rtusa</i> (leaf and fruit extract)	[7]
11	Gallic acid	<i>B. retusa</i> (leaf and fruit	[13]
11	Game actu	extract)	[15]
12	5-Hydroxy-7, 8, 4'-trimethoxy flavone	<i>B. fruticosa</i> (leaf extract)	[14]
14	Glycosides	D. francosa (lear extract)	[14]
	i. Flavonoidal glycosides		
13	Naringenin-6,8-di-C-glucoside	B. retusa (leaf extract)	[13]
13	Naringenin 7- O - β -D-glucopyranoside	<i>B. fruticosa</i> (whole plant	[15]
14	Naringenin 7-0- p-D-grucopyranoside	extract)	[15]
15	3- <i>O</i> -β-D-Glucosyl-7- <i>O</i> -α-L-rhamnosyl-kaempferol	<i>B. androgyna</i> (leaf extract)	[16]
16	$3-O-\beta$ -D-Glucosyl- $(1\rightarrow 6)-\beta$ -D-glucosyl-kaempferol	<i>B. androgyna</i> (leaf extract)	[16]
10	$3-O-\beta-D$ -Glucosyl- $(1 \rightarrow 6)-\beta$ -D-glucosyl- $(2 \rightarrow 6)-\beta$ -D-glucosyl- $($	<i>B. androgyna</i> (leaf extract)	[16]
17	rhamnosyl-kaempferol	D. anarogyna (lear extract)	[10]
18	Hydroquinone O -[6-(3-hydroxyisobutanoyl)]- β -	B. fruticosa (leaf extract)	[17]
10	galactopyranoside	D. francosa (loar oxador)	[1,]
19	3-Acetyl-(-)-epicatechin 7-O-β-glucopyranoside	B. fruticosa (leaf extract)	[17]
20	3-Acetyl-(-)-epicatechin 7-O-(6-isobutanoyloxyl)- β-	<i>B. fruticosa</i> (leaf extract)	[17]
20	glucopyranoside	D. francosa (lear extract)	[1/]
21	3-Acetyl-(-)-epicatechin 7- <i>O</i> -[6-(2-methyl	B. fruticosa (leaf extract)	[17]
-1	butanoyloxyl)]- β -glucopyranoside	D. francosa (lear extract)	[1/]
22	(2R, 3R)-3-Acetyl-7-methoxy-(–)-epicatechin 5-O-(6-	B. fruticosa (whole plant	[18]
	isobutanoyl)-β-D-glucopyranoside	extract)	[10]
23	(2R, 3R)-3-Acetyl-7-methoxy-(–)-epicatechin 5-O-[6-	<i>B. fruticosa</i> (whole plant	[18]
20	(2m, 5R) 5 receipt 7 methody () epicateenin 5 6 [6 (2-methylbutanoyl)]- β -D-glucopyranoside	extract)	[10]
	ii. Isoprenoid glycosides	entruety	
24	3α , 6α -Dihydroxymegastigman-7-en-9-one-3- O - β -D-	B. fruticosa (leaf extract)	[19]
	apiofuranosyl- $(1\rightarrow 6)$ - β -D-glucopyranoside	Diffiniteosa (lear endace)	[1/]
25	Friedelan-3b-ol	B. fruticosa (leaf extract), B.	[7, 17]
		<i>glauca</i> (leaf extract)	L', 1/
26	Friedelin	<i>B. fruticosa</i> (leaf extract), <i>B</i> .	[7, 17]
		<i>glauca</i> (leaf extract)	L', 1/
27	β-Sitosterol	<i>B. glauca</i> (leaf extract)	[7]
28	3-Oxo-sitosterone	<i>B. glauca</i> (leaf extract)	[7]
20 29	Corchoionoside C	<i>B. androgyna</i> (leaf extract), <i>B.</i>	[16, 20
_/		<i>rostrata</i> (leaf extract)	LIU, 20
30	Sauroposide	<i>B. androgyna</i> (leaf extract)	[16]

31	Betulalbuside A	B. officinalis (leaf extract)	[21]
32	(5Z)-6-[5-(2-Hydroxypropan-2-yl)-2-methyl-	B. fruticosa (leaf extract)	[17]
	tetrahydrofuran-2-yl]-3-methylhexa-1, 5- dien-3-O-b-		
	glucopyranoside		
33	Isolariciresinol 3R- O - β -D-glucopyranoside	B. rostrata (leaf extract)	[22]
	iii. Ionol glycosides		
34	Alangionoside A	B. fruticosa (leaf extract)	[23]
35	Alangionoside B	B. fruticosa (leaf extract)	[24]
	iv. Megastigmane glucosides	.	
36	Icariside B2	B. fruticosa (leaf extract)	[25]
37	Boscialin 4'- O - β -D-glucopyranoside	B. fruticosa (leaf extract)	[26]
38	Byzantionoside B	<i>B. rostrate</i> (leaf extract)	[27]
39	(3S, 5R, 6S, 9S)-Megastigman-7-ene-3, 6, 9-triol 9-O-	B. officinalis (leaf extract)	[21]
	β-D-glucopyranoside		
40	Breyniaionoside A	B. officinalis (leaf extract)	[21]
41	Breyniaionoside B	<i>B. officinalis</i> (leaf extract)	[21]
42	Breyniaionoside C	<i>B. officinalis</i> (leaf extract)	[21]
43	Breyniaionoside D	<i>B. officinalis</i> (leaf extract)	[21]
-0	v. N-glycosides	D. officinaris (lear extract)	[21]
44	Guanosine	B. androgyna (leaf extract)	[16]
45	Adenosine	<i>B. androgyna</i> (leaf extract)	[16]
46	5'-Deoxy-5'-methylsulphinyl-adenosine	<i>B. androgyna</i> (leaf extract)	[16]
47	Uridine	<i>B. androgyna</i> (leaf extract)	[16]
	vi. Phenolic glycosides	<i>D. unurogynu</i> (lear extract)	[10]
48	<i>Cis-p</i> -coumaric acid 4- O -(2'- O - β -D apiofuranosyl)- β -	B. fruticosa (leaf extract)	[28]
-10	D-glucopyranoside	<i>B. francosa</i> (lear extract)	[20]
49	<i>Trans-p</i> -coumaric acid 4- O -(2'- O - β -D-apiofuranosyl)-	B. fruticosa (leaf extract)	[28]
47	β-D-glucopyranoside	<i>B. fruitcosu</i> (leaf extract)	[20]
50	Robustaside A	D officiants (loof outroot)	[21]
50 51	Examin	B. officinalis (leaf extract)	[21]
51 52	Isorobustaside A	B. officinalis (leaf extract)	[21]
52 53		B. officinalis (leaf extract)	[21]
55 54	2-Phenylethyl - β D-glucopyranoside	B. fruticosa (leaf extract)	[29]
	Benzyl 6-O- β -D-apiofuranosyl- β -D-glucopyranoside	<i>B. rostrata</i> (leaf extract)	[17]
55	4-(4-O-β-Glucopyranosyl-phenoxy)-1-O-β-	B. fruticosa (leaf extract)	[17]
=(β glucopyranosyl-1, 3-benzenediol		[17]
56	7, 8-Erythro-dihydroxy- 3, 4, 5-trimethoxy-phenyl-	B. fruticosa (leaf extract)	[17]
57	propane8- O - β -glucopyranoside	D functioner (loof outwort) D	[7 20 21]
57	Arbutin	<i>B. fruticosa</i> (leaf extract), <i>B.</i>	[7, 30, 31]
		rostrata (leaf extract), B.	
-0		glauca (leaf extract)	[20]
58	Phlebotrichin	<i>B. rostrata</i> (leaf extract)	[32]
59	$Cis-p$ -coumaric acid 4-O β -D-glucopyranoside	<i>B. rostrata</i> (leaf extract)	[28]
60	$(3S, 6R)$ -Cis-linalool-3,6-oxide β -D-glucopyranoside	<i>B. rostrata</i> (leaf extract)	[23]
61	Kaempferol-3-O-rutinoside	<i>B. glauca</i> (leaf extract)	[7]
62	Quercetin-3-O-glucoside	B. glauca (leaf extract), B.	[7] [13]
		rtusa (leaf and fruit extract)	
63	6-O-Benzoylarbutin	B. vitis-idaea (leaf extract)	[33]
64	Breynioside B	B. vitis-idaea (leaf extract),	[21, 33]
		B.officinalis (leaf extract)	
65	Breynioside A	B. officinalis (leaf extract)	[21]
66	6- <i>O</i> -Benzoyl-α-D-glucose	B. vitis-idaea (leaf extract)	[33]
67	Syringin	B. fruticosa (leaf extract)	[34]
68	(+)-3α-O-β-Glucosyl-isolariciresinol	B. androgyna (leaf extract)	[16]
69	(-)-3α-O-β-Glucosyl-isolariciresinol	B. androgyna (leaf extract)	[16]

70	(-)- 3α - O - β -Apiofuranosyl-($1\rightarrow 2$)- O - β -glucosyl- isolariciresinol	<i>B. androgyna</i> (leaf extract)	[16]
71	(+)-Di-<i>O</i>-β-glucosyl-syringaresinolvii. Simple Glycoside	B. androgyna (leaf extract)	[16]
72	6-O-Methylpropanoyl-alpha-D-glucopyranoside	B. rostrata (leaf extract)	[35]
73	6, 7-Dimethylbenzofuranol 5- O - β -xylopyranosyl- (1 \rightarrow 6)- β -glucopyranoside	<i>B. rostrata</i> (leaf extract)	[17]
74	(-)-5'- β -D-Glucopyranosyloxyjaamonic acid	B. fruticosa (leaf extract)	[36]
75	 (-)-5'- β -D-Glucopyranosyloxyjaamonic acid methyl ester viii. Sulfur-containing spiroacetal glycoside 	B. rostrata (leaf extract)	[36]
76	4-[(Carboxymethyl)thio]-5'-hydroxyphyllaemblic acid O - β -D-glucopyranosyl-(1 \rightarrow 2)- β -D- glucopyranoside ester ix. Spiroketal glycosides	<i>B. fruticosa</i> (whole plant extract)	[18]
77 78	Breinin B Breinin D	<i>B. fruticosa</i> (whole plant extract), <i>B. retusa</i> (stem bark extract) <i>B. fruticosa</i> (whole plant	[8, 13]
		extract), <i>B. retusa</i> (stem bark extract)	[8, 13]
79	Epibreinin B	<i>B. fruticosa</i> (whole plant extract)	
80	Epibreinin D	<i>B. fruticosa</i> (whole plant extract), <i>B. retusa</i> (stem bark extract)	[8] [8, 13]
	Lignans	extract)	[0, 15]
81	Aviculin	B. fruticosa (leaf extract)	
01	Neolignan	D. Jrancosa (lear extract)	
	2R, 3R-2,3-Dihydro-2-(40-hydroxy-30-		[37]
82	methoxyphenyl)-3-(glucosyloxymethyl)-7- hydroxy-5-	B. fruticosa (leaf extract)	[37]
0	benzofuranpropanol 9,9'- Hydroxy-3,4- methylenedioxy-3'-methoxy [7-O-	<i>B. rostrate</i> (leaf extract)	[38]
83	4', 8–, 8-5'5'] neolignan	D. rostrate (lear extract)	
84	8- <i>O</i> -4'-Neolignan 3'- <i>O</i> - β -glucoside Steroids	B. rostrate (leaf extract)	[39]
85	3β, 20β-diol-stigmast-5-ene	B. androgyna (leaf extract)	[40]
86	3β, 20β-diol-stigmasta-5,24(28)-diene	<i>B. androgyna</i> (leaf extract)	[10]
00	Terpenoids	D. anarogyna (rear extract)	[16]
87	Dihydrophaseic acid	B. rostrata (leaf extract)	[16]
88	Arborinone	<i>B. fruticosa</i> (leaf extract)	[10]
89	Isoarborinol	<i>B. fruticosa</i> (leaf extract)	
0,	Tannins	D. francosti (leur extract)	[41]
90	1-O-galloyl-beta-D-glucopyranoside	B. rostrata (leaf extract)	[41] [14] [14]
			[35]

The EtOAc extract of B. vitis-idaea leaves shows antioxidant and tyrosinase inhibitory activity. To detect antioxidants, the most widely used chromogens are the stable free radical DPPH (2,2-diphenyl-1-picrylhydrazyl) and ABTS (2,2'-azino-bis(3-ethylbenzothiazoline)-6-sulphonic acid), respectively. The DPPH or ABTS assay on the EtOAc extract of leaves shows the highest activity with an IC₅₀ value of $85.2 \pm 0.1 \ \mu \text{g/mL}$; the *n*-BuOH extract of leaves shows tyrosinase inhibition activity with an IC₅₀ value of $650 \pm 0.9 \,\mu\text{g/mL}$. Due to the presence of phenolic compounds (hydroxyl group), they show scavenging activity [28]. Hexane, EtOAc, and MeOH extracts of B. glauca leaves show antioxidant activity at a concentration of more than 50 μ g/mL (due to the presence of kaempferol-3-Orutinoside and quercetin-3-O-glucoside) [7]. Different assays for xanthine oxidase (XOD) and elastase were performed on petroleum ether, MeOH, DMSO with water, or simply with DMSO extracts of B. disticha leaves and bark. The results indicate that the leaf and bark extract of B. disticha exhibits 32%, 16%, 44%, and 32% inhibition of XOD and elastase [51]. EtOH and CHCl₃ extracts of B. retusa leaves and stem show activity with an IC₅₀ value of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays, respectively, at a concentration of 100 µg/mL (leaves have better antioxidant activity than the stem) [52]. Proteins are the major constituents of *B. androgyna* leaves. In antioxidant testing, EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, and 0.198 Abs, respectively, high at a concentration of 50 μ g/mL (strong activity) compared to tannic acid, standard curcumin, curcumin, and vitamin E (0.54 Abs, 62.31%, 75.38%, and 0.15 Abs, respectively). These results reveal that the leaf extract of *B. androgyna* is effective against all those diseases which are mediated by free radicals [53].

In general, phenolic compounds (block virus entry or attachment to host cells), flavonoids (selectively inhibit viral RNA propagation), glycosides (reduce viral protein expression), lignin (immunomodulatory effect), etc. are effective against viral activities [54]. In this genus, antiviral activity has been studied in only one species, *B. oblongifolia*. Aqueous extract of the whole plant shows activity against duck hepatitis B virus (DHBV), with 50% inhibition at 500 μ g/mL, due to the presence of these phytochemicals. The inhibitory activity is dose-dependent (inhibition increases with an increase in dose or concentration) [10].

The hexane, EtOAc, and MeOH extracts of B. glauca leaves show anti-inflammatory activity at concentrations of more than 50 μ g/mL (due to the presence of friedelan-3β-ol, β-sitosterol, and kaempferol) because phenolic compounds are best for anti-inflammatory activity [7]. The MeOH and n-BuOH whole plant extracts of *B. fruticosa* contain sesquiterpenoids and four breynins; their names are mentioned above. This extract prevents rats from arthritis deterioration, showing 50% inhibition at a concentration of 0.2 mg/kg; this inhibition rate was compared with that of indomethacin at a dose of 2 mg/kg. The results indicate that breynins are the toxic components of *B. fruticosa*, having strong anti-inflammatory effects [8]. Aqueous extract of B. retusa leaves shows 87.3% and 63.77% membrane stabilization at concentrations of 1000 and 62.5 µg/mL, respectively. The results indicate anti-arthritic and membrane stabilizing activity at higher concentrations of extract [55]. EtOH extract of B. androgyna leaves shows 74.17% protection in a hypotonic-induced haemolysis model and 83.60% protection in a protein denaturation model at 100 µg/mL. It protects and stabilizes red blood cell membranes [53]. This extract inhibits the discharge of lysosomal content from polyphenolic neutrophils (due to the presence of flavonoids) [9]. EtOH and MeOH extracts of B. nivosa leaves show 18% and 22% inhibition at 100 and 200 mg/mL (approximately equal to the percentage inhibition of diclofenac). This is done by inhibiting histamine or serotonin [49].

Ethyl acetate and MeOH extracts of *B. retusa* leaves show 98% inhibition of α -amylase at 60 µg/mL [56]. Supplementation with 90 mg/kg aqueous leaf extract of *B. androgyna* reduces LDL, VLDL, HDL, and cholesterol (due to the presence of alkaloids). Digestion of 5 g/100 mL of leaf extract significantly lowers the blood glucose level (referred to as diabetic greens) [57]. Aqueous and EtOH extracts of *B. vitis-idaea* leaves elicit a 60.89% and 63.37% decline in glucose level at 300 µg/mL of both extracts, respectively, having significant hypoglycaemic (p < 0.001) and hypolipidaemic (p < 0.01) activity [58]. Aqueous MeOH extract of *B. officinalis* whole plant shows hypocholesterolaemic activity at 0.02, 0.005, 0.01, and 0.025 mg/kg/day (due to the presence of breynin A and B). The breynin B activity is about one-fifth of the activity of breynin A [59].

EtOH extract of *B. fruticosa* whole plant shows tyrosinase inhibition with IC_{50} values of 0.89 and 0.82 respectively (due to the presence of spiroacetal glycosides, i.e. compounds **19**, **21**, **22**, and **23**). Tyrosinase is a copper-containing enzyme that catalyses the production of melanin. Tyrosinase inhibitors specifically inhibit melanogenesis in the cell without side effects [13].

Gold and silver nanoparticles synthesized from the stem bark of *B. rhamnoides* show an efficient reduction of 4-nitrophenol to 4-aminophenol [60]. In the body, accumulation of 4-nitrophenol causes blood disorders, such as reducing the oxygen-carrying capacity of the blood [61].

EtOH extract of leaves is more effective against Salmonella [43] B. retusa EtOH extract of leaves is more effective against Salmonella [43] Typhimurium, E. coli, and Enterobacter aerogenes and less effective against Trichophyton rubrum, Aspergillus niger, and Penicillium sp. and less effective against Cryptococcus sp. fungi [44] Antimicrobial activity B. cernua Petrol, CH-CL, BuOH, EtOAc, and MeOH extracts of leaves are effective against Cryptococcus aureus, Ieon and less effective against Cryptococcus aureus, Ieon and EtOH extracts of leaves are effective against Corptococcus aureus, Ieon and AE (MeOH extracts of leaves are effective against Corptococcus aureus, IeoH extract of leaves is effective against Corptococcus aureus, IeoH extract of leaves is effective against Corptococcus aureus, IeoH extract of leaves is effective against Corptococcus aureus, IeoH extract of leaves is effective against Corptococcus aureus, IeoH extract of leaves is effective against Corptococcus and EtOH extract of leaves is effective against Corptococcus and EtOH extract of leaves is effective against Corptococcus and EtOH extract of leaves is effective against Corptococcus and EtOH extract of leaves and EtOH extract (250–2500 mg/L) shows a [9] [9] Cytotoxic B. ernua n-Hexane fraction shows good cytotoxic activity against indee averant of leaves and EtOH extracts of leaves and EtOH extract of leaves and EtOH extract of leaves show activity (ICs_0 650 ± 0.9 µg/mL)	Pharmacological activity	Plant sources	Results	Referenc
B. retusa effective against Micrococcus luteus EtOH extract of leaves is more effective against Tricholphyton rubrum, Aspergillus riger, and Penicillium sp. and less effective against Ifungi Antimicrobial activity B. cernua Petrol, CH_2C1, BuOH, EtOAc, and MeOH extracts of [44] Antimicrobial activity B. androgyna [45, 46] Proteus vulgaris, Bacillus cereus, Staphylococcus aureus, [47] [47] Activity B. androgyna [48] B. disticha MeOH extract of leaves is effective against I fungi [48] B. disticha MeOH extract of leaves is effective against B. subtilis, [49] [48] Cytotoxic B. nivosa EtOH extract is effective against B. subtilis, [49] [50] S. cernua n-Hexane fraction shows good cytotoxic activity against [50] Cytotoxic B. androgyna EtOH and MeOH leaf extract (250–2500 mg/L) shows a [9] [9] Cytotici (Cigo 650 ± 0.9 µg/mL) B. androgyna EtOH and MeOH extracts of leaves shows activity against tideaa [9] Cytotici (Cigo 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extracts of leaves shows activity (Cigo 650 ± 0.9 µg/mL) [17] Activity B. androgyna DPPH or ABTS assay of EtOAc extract of leaves and sterivity (Cigo 650 ± 0.9 µg/mL) [17] <td></td> <td></td> <td></td> <td>[43]</td>				[43]
EtOH extract of leaves is more effective against <i>Trichophyton rubrum, Aspergillus niger, and Penicillium sp.</i> and less effective against <i>Aspergillus niger, and Penicillium sp.</i> and less effective against <i>Cryptococcus sp.</i> fungi 		B. retusa		
$ \begin{array}{c} Trichophyton rubrum, Aspergillus niger, and Penicillium sp. and less effective against Cryptococcus sp. fungi \\ Petrol, CH_2Cb, BuOH, EtOAc, and MeOH extracts of [44] \\ leaves are effective against all fungi \\ MeOH and EtOH extracts of leaves are effective against [45, 46] \\ Proteus vulgaris, Bacillus cereus, Staphylococcus aureus, [47] \\ activity B. androgyna E. coli, Klebsiella pneumonia, Gram-positive bacteria (MeOH extract of leaves is effective against Candida albicans and Aspergillus flavus B. disticha MeOH extract is effective against S. aureus and E. coli and effective against S. cerevisiae fungus EtOH extract of leaves is effective against B. subtilis, [49] S. aureus, and Salmonella Typhi B. androgyna EtOH extract of leaves is effective against B. subtilis, [49] S. aureus, and Salmonella Typhi B. androgyna EtOH extract of leaves is effective against B. subtilis, [49] S. aureus, and Salmonella Typhi B. androgyna EtOH extract of leaves is effective against B. subtilis, [40] S. aureus, and Salmonella Typhi B. androgyna EtOH extract of leaves hows good cytotoxic activity against [50] DPPH or ABTS assay of EtOAc extract of leaves shows a [9] cytotoxic effect on human mesenchymal stem cells DPPH or ABTS assay of EtOAc extract of leaves shows activity (IC50 650 ± 0.9 µg/mL) and n-BuOH extract of leaves shows tyrosinase inhibition activity (IC50 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extracts of leaves and stem show activity (IC50 650 ± 0.9 µg/mL). Hexane, EtOAc and MeOH extracts of leaves and [51] bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl1 extracts of leaves and stem show activity [52] with IC50 of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, [53] DPPH, reducing power, alkaline DMSO, and phosphomolydehum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL [451]$				
and less effective against <i>Cryptococcus</i> sp. fungi [44] Petrol, CH ₂ Cl ₂ , BuOH, EtOAc, and MeOH extracts of [44] leaves are effective against all fungi [45, 46 Antimicrobial MeOH and EtOH extracts of leaves are effective against <i>Proteus vulgaris, Bacillus cereus, Staphylococcus aureus,</i> [47] activity B. androgyna [45, 46 Proteus vulgaris, Bacillus cereus, Staphylococcus aureus, [47] activity B. androgyna [48] B. disticha MeOH extract shows more inhibition than EtOH extract) [48] EtOH extract is effective against <i>S. aureus</i> and <i>E. coli</i> and effective against <i>S. cerevisiae</i> fungus [49] B. nivosa EtOH extract of leaves is effective against <i>B. subtilis</i> , [49] S. aureus, and Salmonella Typhi [49] B. eernua n-Hexane fraction shows good cytotoxic activity against activity B. androgyna [50] B. androgyna EtOH and MeOH leaf extract (250–2500 mg/L) shows a [9] [21] cytotoxic DPPH or ABTS assay of EtOAc extract of leaves shows [28] highest activity (ICs ₀ 65.2 ± 0.1 µµ/mL) and n-BuOH Proteus crutice fleaves shows tyrosinase inhibition activity (ICs ₀ 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extrac				
B. cernua Petrol, CH ₂ Cl ₂ , BuÕH, EtoÅc, and MeÕH extracts of [44] leaves are effective against all fungi [45, 46] Antimicrobial activity B. androgyna [47] [47] B. androgyna E. coli, Klebsiella pneumonia, Gram-positive bacteria (MeOH extract shows more inhibition than EtOH extract) EtOH extract of leaves is effective against S. aureus, and Aspergillus flavus [47] B. disticha MeOH extract is effective against S. aureus and E. coli and effective against S. cerevisiae fungus [48] B. nivosa EtOH extract of leaves is effective against S. subbilis, S. aureus, and Salmonella Typhi [49] Cytotoxic activity B. cernua n-Hexane fraction shows good cytotoxic activity against (Cytotoxic effect on human mesenchymal stem cells [9] DPPH or ABTS assay of EtOA extract of leaves shows a [9] [20] [21] [23] Vitoxic activity B. glauca at concentrations greater than 50 µg/mL. [41] Hexane, EtOA and MeOH extracts of leaves show activity (IC ₃₀ 650 ± 0.9 µg/mL.) [23] [32] [32] Highest activity (IC ₃₀ 85.2 µg/mL.) Hexane, EtOA-cuttons delease by 32%, 16%, 44%, and 32%, respectively [51] Antioxidant activity B. disticha bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively [52] B. retusa with IC				
Antimicrobial activityleaves are effective against and EtOH extracts of leaves are effective against Proteus vulgaris, Bacillus cereus, Staphylococcus aureus, E. coli, Klebsiella pneumonia, Gram-positive bacteria (MeOH extract shows more inhibition than EtOH extract) EtOH extract of leaves is effective against <i>Candida albicans</i> and Aspergillus flavus[47]B. distichaMeOH extract of leaves is effective against <i>S. aureus</i> and <i>E. coli</i> and effective against <i>S. cerevisiae</i> fungus B. nivosa[48] effective against <i>S. cerevisiae</i> fungus B. nivosa[49] S. aureus, and Salmonella TyphiB. cernuan-Hexane fraction shows good cytotoxic activity against cytotoxic effect on human mesenchymal stem cells[50]Cytotoxic activityB. androgynaEtOH and MeOH leaf extract (250–2500 mg/L) shows a cytotoxic effect on human mesenchymal stem cells[28]DPH or ABTS assay of EtOAc extract of leaves shows idaea[28]DPH or ABTS assay of EtOAc extract of leaves shows toxic effect on human mesenchymal stem cells[28]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside)[51]B. retusabark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl ₃ extracts of leaves and stracts of leaves and stem show activity exited activity for 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, EtOH extract of leaves shows, activity in hydroxyl radical, <b< td=""><td></td><td>B. cernua</td><td></td><td>[44]</td></b<>		B. cernua		[44]
Antimicrobial activity B. androgyna Proteus vulgaris, Bacillus cereus, Staphylococcus aureus, E. coli, Klebsiella pneumonia, Gram-positive bacteria (MeOH extract shows more inhibition than EtOH extract) EtOH extract of leaves is effective against Candida albicans and Aspergillus flavus [47] B. disticha MeOH extract of leaves is effective against Candida albicans and Aspergillus flavus [48] B. nivosa EtOH extract of leaves is effective against S. aureus and E. coli and effective against S. cerevisiae fungus [49] Cytotoxic B. nivosa EtOH extract of leaves is effective against B. subtilis, S. aureus, and Salmonella Typhi [49] B. cernua n-Hexane fraction shows good cytotoxic activity against MCF-7 breast cancer cells (ICso 165.65 ppm) [9] extivity B. androgyna EtOH and MeOH leaf extract (250–2500 mg/L) shows a cytotoxic effect on human mesenchymal stem cells [28] bPPH or ABTS assay of EtOAc extract of leaves shows idaea highest activity (ICs ₀ 85.2 ± 0.1 µg/mL) and n-BuOH [47] extract of leaves shows trosinase inhibition activity (ICso 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extracts of leaves show activity (ICso 650 ± 0.9 µg/mL) [7] Antioxidant activity B. disticha bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl ₃ extracts of leaves and stem show activity (ICO oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical,				
activityB. androgynaE. coli, Klebsiella pneumonia, Gram-positive bacteria (MeOH extract shows more inhibition than EtOH extract) EtOH extract of leaves is effective against Candida albicans and Aspergillus flavusB. distichaMeOH extract is effective against S. aureus and E. coli and effective against S. cerevisiae fungus[48]B. nivosaEtOH extract of leaves is effective against B. subtilis, s. aureus, and Salmonella Typhi[49]B. cernuan-Hexane fraction shows good cytotoxic activity against cytotoxic[50]B. androgynaEtOH extract of leaves is effective against S. 5pm)[9]Cytotoxic activityB. androgynaEtOH and MeOH leaf extract (250–2500 mg/L) shows a cytotoxic effect on human mesenchymal stem cells[9]DPPH or ABTS assay of EtOAc extract of leaves shows (ICso 650 ± 0.9 µg/mL)[28]Breynia vitis- idaeahighest activity (ICso 85.2 ± 0.1 µg/mL) and n-BuOH extract of leaves show strostinase inhibition activity (ICso 650 ± 0.9 µg/mL)[7]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH and CMCH extracts of leaves and 32%, respectively[51]B. retusawith ICso 0 52, 63, 31, 2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging asays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, phosphomolybdenum assays of 55, 62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL				[45, 46]
(MeOH extract shows more inhibition than EtOH extract) EtOH extract of leaves is effective against <i>Candida albicans</i> and <i>Aspergillus flavus</i> B. disticha MeOH extract of leaves is effective against <i>S. aureus</i> and <i>E. coli</i> and effective against <i>S. cerevisiae</i> fungus B. nivosa EtOH extract of leaves is effective against <i>B. subtilis</i> , <i>S. aureus</i> , and <i>Salmonella</i> Typhi B. cernua n-Hexane fraction shows gootytotxic activity against Cytotoxic MCF-7 breast cancer cells (ICs0 165.65 ppm) B. androgyna EtOH and MeOH leaf extract (250–2500 mg/L) shows a [9] cytotoxic effect on human mesenchymal stem cells DPPH or ABTS assay of EtOAc extract of leaves shows [28] Breynia vitis- idaea extract of leaves shows tyrosinase inhibition activity (ICs0 650 ± 0.9 µg/mL) [4] Hexane, EtOAc and MeOH extracts of leaves show activity (ICs0 650 ± 0.9 µg/mL) [7] B. glauca at concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) [51] Petroleum ether, MeOH, and DMSO extracts of leaves and 32%, respectively [52] [52] B. retusa with ICs0 of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH and CHCl3 extracts of leaves shows activity [52] B. androgyna DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%,	Antimicrobial		Proteus vulgaris, Bacillus cereus, Staphylococcus aureus,	[47]
EtOH extract of leaves is effective against Candida albicans and Aspergillus flavusB. distichaMeOH extract is effective against S. aureus and E. coli and effective against S. cerevisiae fungus[48]B. nivosaEtOH extract of leaves is effective against B. subtilis, S. aureus, and Salmonella Typhi[49]S. aureus, and Salmonella TyphiB. cernuan-Hexane fraction shows good cytotoxic activity against (Cytotoxic B. androgyna[50]DETOH extract of leaves is effective against B. subtilis, S. aureus, and Salmonella TyphiactivityB. cernuan-Hexane fraction shows good cytotoxic activity against (Cytotoxic effect on human mesenchymal stem cellsDPPH or ABTS assay of EtOAe extract of leaves showsDPPH or ABTS assay of EtOAe extract of leaves shows (ICso 650 \pm 0.9 µg/mL)Hexane, EtOAc and MeOH extracts of leaves show activity (ICso 650 \pm 0.9 µg/mL)Berynia vitis- highest activity (ICso 650 \pm 0.9 µg/mL)Hexane, EtOAc and MeOH extracts of leaves show activity (ICso 650 \pm 0.9 µg/mL)Berynia vitis- highest activity (ICso 650 \pm 0.9 µg/mL)B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and 32%, respectivelyB. retusawith ICso of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at	activity	B. androgyna	E. coli, Klebsiella pneumonia, Gram-positive bacteria	
and Aspergillus flavus [48] B. disticha MeOH extract is effective against S. aureus and E. coli and effective against S. cerevisiae fungus [49] B. nivosa EtOH extract of leaves is effective against B. subtilis, subtil			(MeOH extract shows more inhibition than EtOH extract)	
B. distichaMeOH extract is effective against S. aureus and E. coli and effective against S. cerevisiae fungus[48]B. nivosaEtOH extract of leaves is effective against B. subtilis, S. aureus, and Salmonella Typhi[49]Cytotxic activityB. cernuan-Hexane fraction shows good cytotoxic activity against MCF-7 breast cancer cells (ICso 165.65 ppm)[50]B. androgynaEtOH and MeOH leaf extract (250–2500 mg/L) shows a cytotoxic effect on human mesenchymal stem cells[9]DPPH or ABTS assay of EtOAc extract of leaves shows idaea[28]Breynia vitis- idaeahighest activity (ICso 85.2 ± 0.1 µg/mL) and n-BuOH (ICso 650 ± 0.9 µg/mL)[7]Hexane, EtOAc and MeOH extracts of leaves show activity (ICso 650 ± 0.9 µg/mL)[7]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and 32%, respectively[51]Antioxidant activityB. distichabark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively[52]B. retusawith ICso 0 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL[53]			EtOH extract of leaves is effective against Candida albicans	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $			and Aspergillus flavus	
B. nivosaEtOH extract of leaves is effective against B. subtilis, S. aureus, and Salmonella Typhi[49]B. cernuan-Hexane fraction shows good cytotoxic activity against MCF-7 breast cancer cells (ICs0 165.65 ppm)[50]activityB. androgynaEtOH and MeOH leaf extract (250–2500 mg/L) shows a cytotoxic effect on human mesenchymal stem cells[9]DPPH or ABTS assay of EtOAc extract of leaves shows[28]bight activityICs0 650 ± 0.1 µg/mL)[7]idaeaextract of leaves shows tyrosinase inhibition activity (ICs0 650 ± 0.9 µg/mL)[7]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside)[51]Antioxidant activityB. distichabark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[52]B. retusawith ICs0 0f 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL		B. disticha	MeOH extract is effective against S. aureus and E. coli and	[48]
$\begin{array}{c c c c c c c c c c c c c c c c c c c $			effective against S. cerevisiae fungus	
B. cernuan-Hexane fraction shows good cytotoxic activity against[50]activityB. androgynaEtOH and MeOH leaf extract (250–2500 mg/L) shows a[9]cytotoxic effect on human mesenchymal stem cellspPPH or ABTS assay of EtOAc extract of leaves shows[28]Breynia vitis- idaeahighest activity (IC ₅₀ 650 ± 0.1 µg/mL) and n-BuOHextract of leaves shows tyrosinase inhibition activity (IC ₅₀ 650 ± 0.9 µg/mL)[7]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside)[51]Antioxidant activityB. distichabark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl ₃ extracts of leaves and stem show activity[52]B. retusawith IC ₅₀ of 2, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, pPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL		B. nivosa	EtOH extract of leaves is effective against B. subtilis,	[49]
Cytotoxic activityMCF-7 breast cancer cells (IC $_{50}$ 165.65 ppm)MCF-7 breast cancer cells (IC $_{50}$ 165.65 ppm)activityB. androgynaEtOH and MeOH leaf extract (250–2500 mg/L) shows a cytotoxic effect on human mesenchymal stem cells[9]DPPH or ABTS assay of EtOAc extract of leaves shows idaea[28]Breynia vitis- idaeahighest activity (IC $_{50}$ 85.2 ± 0.1 µg/mL) and n-BuOH extract of leaves shows tyrosinase inhibition activity (IC $_{50}$ 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extracts of leaves show activity at concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and 32%, respectively EtOH and CHCl ₃ extracts of leaves and stem show activity at 02.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, toxide, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL			S. aureus, and Salmonella Typhi	
activityB. androgynaEtOH and MeOH leaf extract (250–2500 mg/L) shows a cytotoxic effect on human mesenchymal stem cells[9]DPPH or ABTS assay of EtOAc extract of leaves shows[28]Breynia vitis- idaeahighest activity (IC ₅₀ 85.2 ± 0.1 µg/mL) and n-BuOH extract of leaves shows tyrosinase inhibition activity (IC ₅₀ 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extracts of leaves show activity[7]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity with IC ₅₀ of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL		B. cernua	n-Hexane fraction shows good cytotoxic activity against	[50]
cytotoxic effect on human mesenchymal stem cellscytotoxic effect on human mesenchymal stem cellsDPPH or ABTS assay of EtOAc extract of leaves showsBreynia vitis- idaeaidaeaBreynia vitis- idaeaidaeaBreynia vitis- idaeaidaeaBreynia vitis- idaeaBreynia vitis- idaeaidaeaBreynia vitis- idaeaBreynia vitis- idaeaAntioxidant a colspan="2">B. glaucaBreynia vitis- presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and stats by 32%, 16%, 44%, and 32%, respectivelyEtOH and CHCl3 extracts of leaves and stem show activityBreynia vitis- Stats by 32% of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract	Cytotoxic		MCF-7 breast cancer cells (IC ₅₀ 165.65 ppm)	
Breynia vitis- idaeaDPPH or ABTS assay of EtOAc extract of leaves shows[28]Breynia vitis- idaeahighest activity (IC50 85.2 ± 0.1 µg/mL) and n-BuOH extract of leaves shows tyrosinase inhibition activity (IC50 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extracts of leaves show activity[7]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[51]B. retusawith IC50 of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL[53]	activity	B. androgyna	EtOH and MeOH leaf extract (250-2500 mg/L) shows a	[9]
Breynia vitis- idaeahighest activity ($IC_{50} 85.2 \pm 0.1 \mu g/mL$) and <i>n</i> -BuOH extract of leaves shows tyrosinase inhibition activity ($IC_{50} 650 \pm 0.9 \mu g/mL$) Hexane, EtOAc and MeOH extracts of leaves show activity[7]B. glaucaat concentrations greater than 50 $\mu g/mL$ (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and 32%, respectively EtOH and CHCl ₃ extracts of leaves and stem show activity[51]B. retusabark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl ₃ extracts of leaves and stem show activity[52]B. retusawith IC_{50} of 26, 31.2, and 25.5 $\mu g/mL$ in DPPH, nitric oxide, and SOD radical scavenging assays at 100 $\mu g/mL$ EtOH extract of leaves shows activity in hydroxyl radical, phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 $\mu g/mL$			cytotoxic effect on human mesenchymal stem cells	
idaeaextract of leaves shows tyrosinase inhibition activity (IC $_{50}$ 650 ± 0.9 µg/mL) Hexane, EtOAc and MeOH extracts of leaves show activity[7]B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and Bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[51]B. retusawith IC $_{50}$ of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL			DPPH or ABTS assay of EtOAc extract of leaves shows	[28]
$\begin{array}{cccc} (IC_{50} 650 \pm 0.9 \ \mu g/mL) & Hexane, EtOAc and MeOH extracts of leaves show activity [7] \\ B. glauca & at concentrations greater than 50 \ \mu g/mL (due to the presence of kaempferol-3-O-rutinoside) \\ Petroleum ether, MeOH, and DMSO extracts of leaves and [51] \\ bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively \\ EtOH and CHCl3 extracts of leaves and stem show activity [52] \\ B. retusa & with IC_{50} of 26, 31.2, and 25.5 \ \mu g/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 \ \mu g/mL \\ EtOH extract of leaves shows activity in hydroxyl radical, [53] \\ B. androgyna & DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 \ \mu g/mL \end{array}$		Breynia vitis-	highest activity (IC ₅₀ 85.2 \pm 0.1 μ g/mL) and <i>n</i> -BuOH	
B. glaucaHexane, EtOAc and MeOH extracts of leaves show activity[7]at concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[51]B. retusawith IC ₅₀ of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL		idaea	extract of leaves shows tyrosinase inhibition activity	
B. glaucaat concentrations greater than 50 µg/mL (due to the presence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[51]B. retusawith IC ₅₀ of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL			$(IC_{50} 650 \pm 0.9 \ \mu g/mL)$	
Antioxidant activityB. distichapresence of kaempferol-3-O-rutinoside) Petroleum ether, MeOH, and DMSO extracts of leaves and bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[51]B. retusawith IC ₅₀ of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL				[7]
Antioxidant activityB. distichaPetroleum ether, MeOH, and DMSO extracts of leaves and bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[51] bark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectivelyB. retusaB. retusawith IC ₅₀ of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL		B. glauca		
Antioxidant activityB. distichabark inhibit XOD and elastase by 32%, 16%, 44%, and 32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[52]B. retusawith IC50 of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL				
activity32%, respectively EtOH and CHCl3 extracts of leaves and stem show activity[52]B. retusawith IC50 of 26, 31.2, and 25.5 µg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 µg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL[53]				[51]
EtOH and CHCl3 extracts of leaves and stem show activity[52]B. retusawith IC50 of 26, 31.2, and 25.5 μg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 μg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 μg/mL		B. disticha		
B. retusawith IC ₅₀ of 26, 31.2, and 25.5 μg/mL in DPPH, nitric oxide, and SOD radical scavenging assays at 100 μg/mL EtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 μg/mL[53]	activity			
oxide, and SOD radical scavenging assays at 100 μg/mLEtOH extract of leaves shows activity in hydroxyl radical,DPPH, reducing power, alkaline DMSO, andphosphomolybdenum assays of 55.62%, 50%, 0.286 Abs,72.51%, 0.198 Abs, respectively, high at 50 μg/mL			•	[52]
B. androgynaEtOH extract of leaves shows activity in hydroxyl radical, DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL[53]		B. retusa		
<i>B. androgyna</i> DPPH, reducing power, alkaline DMSO, and phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL				
phosphomolybdenum assays of 55.62%, 50%, 0.286 Abs, 72.51%, 0.198 Abs, respectively, high at 50 µg/mL		_		[53]
72.51%, 0.198 Abs, respectively, high at 50 μg/mL		B. androgyna		
[10]			72.51%, 0.198 Abs, respectively, high at 50 μg/mL	
				[10]

Table 2. Biological	activity of species	of the genus Breynia

Antiviral activity B. oblongifolia

500 µg/mL aqueous extract of whole plant shows 50% inhibition against duck hepatitis B virus (DHBV)

	B. glauca	Hexane, EtOAc, and MeOH extracts of leaves show activity at concentrations greater than 50 μ g/mL (due to the presence of friedelan-3 β -ol, β -sitosterol, and kaempferol)	[7]
Anti-	B. fruticosa	MeOH and n-BuOH whole plant extracts prevent rats from arthritis deterioration with 50% inhibition at 0.2 mg/kg (due to the presence of acception provide and hermite)	[8]
inflammatory activity	B. retusa	to the presence of sesquiterpenoids and breynins) Aqueous extract of leaves shows 87.3% and 63.77% membrane stabilization at 1000 and 62.5 µg/mL and moderate anti-arthritic activity	[55]
	B. androgyna	EtOH extract of leaves shows 74.17% and 86.88% protection and stabilization to membranes of red blood cells at 100 μ g/mL	[53] [9]
	B. nivosa	EtOH and MeOH leaf extracts show 18% and 22% inhibition at 100 and 200 mg/mL	[49]
		EtOAc and MeOH extracts of leaves show 98% inhibition	[56]
	B. retusa	of α -amylase at 60 µg/mL	
	B. androgyna	Supplementation with 90 mg/kg aqueous extract of leaves reduces LDL, VLDL, HDL, and cholesterol; digestion of leaves lowers the blood glucose level	[57]
Antidiabetic activity	B. vitis-idaea	Aqueous and EtOH extracts of leaves show a 60.89% and 63.37% decline in glucose level at 300 μ g/mL of both extracts, respectively, having significant hypoglycaemic (p < 0.001) and hypolipidemic (p < 0.01) activity	[59]
	B. officinalis	Aqueous MeOH extract of whole plant shows hypocholesterolaemic activity at 0.02, 0.005, 0.01, and 0.025 mg/kg/day	[59]
Enzyme inhibition	B. fruticosa	EtOH extract of the whole plant shows tyrosinase inhibition with IC_{50} values of 0.89 and 0.82, respectively	[13]
Blood disorder	B. rhamnoides	Gold and silver nanoparticles of this species reduced 4- nitrophenol to 4-aminophenol (accumulation of 4- nitrophenol causes blood disorder)	[61]

4. Conclusion

The species of this genus are widely distributed in Australia, Malaysia, Vietnam, and India. A total of 90 compounds are mentioned in this review; these secondary metabolites are very effective for the treatment of several diseases, like cancer, arthritis, diabetes, fever, cough, and blood disorders, and act as antioxidants. This antioxidant activity can be proved best for the treatment of many diseases, which are caused due to oxidative stress, like for the treatment of neurodegenerative disorders. There are about 38 species in this genus, out of which the pharmacology and phytochemistry of ten species have been identified. In this way, much more consideration ought to be paid to sort *Breynia* for the disclosure of novel phytochemicals and their pharmacological assessment. This will help to cope with various diseases by introducing novel therapeutic agents to the world health community.

Supporting Information

Supporting information accompanies this paper on <u>http://www.acgpubs.org/journal/records-of-natural-products</u>

ORCID 回

Malik Saadullah: <u>0000-0002-2453-5769</u> Muhammad Asif: <u>0000-0001-8989-9945</u> Sania Arif: <u>0000-0002-5167-8001</u> Bisma Kanwal: <u>0000-0003-3192-6191</u>

References

- [1] M. M. Aye, H. T. Aung, M. M. Sein and C. Armijos (2019). A review on the phytochemistry, medicinal properties and pharmacological activities of 15 selected Myanmar medicinal plants, *Molecules* **24**(2), 293.
- [2] P. Hoffmann, H. Kathriarachchi and K. J. Wurdack (2006). A phylogenetic classification of Phyllanthaceae (Malpighiales; Euphorbiaceae sensu lato), *Kew Bull.* **61**(1), 37-53.
- [3] M. S. Vorontsova and P. Hoffmann (2008). A phylogenetic classification of tribe Poranthereae (Phyllanthaceae, Euphorbiaceae sensu lato), *Kew Bull.* **63**(1), 41-59.
- [4] P. Van Welzen, K. Pruesapan, I. Telford, H. J. Esser and J. Bruhl (2014). Phylogenetic reconstruction prompts taxonomic changes in *Sauropus, Synostemon*, and *Breynia* (Phyllanthaceae tribe Phyllantheae), *Blumea.* **59(2)**, 77-94.
- [5] A. T. Khalil, Z. K. Shinwari, M. Qaiser and K. B. Marwat (2014). Phyto-therapeutic claims about Euphorbiaceous plants belonging to Pakistan; an ethnomedicinal review, *Pak. J. Bot.* **46**(**3**), 1137-1144.
- [6] D. Holdsworth, B. Pilokos and P. Lambes (1983). Traditional medicinal plants of New Ireland, Papua New Guinea Part. II. New Hanover Island, *Int. J. Crude Drug Res.* **21**(4), 161-168.
- [7] B. Supudompol, S. Wongseripipatana and K. Likhitwitayawuid (2005). Chemical constituents of *Breynia glauca* leaves, *Songklanakarin J. Sci. Technol.* **27**(**Suppl. 2**), 563-7.
- [8] X. L. He, J. J. Lv, X. Wang, Q. Zhang, B. Zhang, K. Cao, L. L. Liu and Y. Xu (2019). The identification and isolation of anti-inflammatory ingredients of ethno medicine *Breynia fruticosa*, *J. Ethnopharmacol.* **239**, 111894.
- [9] F. Fikri and M. T. Purnama (2020). Pharmacology and phytochemistry overview on *Sauropus* Androgynous, Sys. Rev. Pharm. **11(6)**, 124-128.
- [10] A. Shead, K. Vickery, A. Pajkos, R. Medhurst, J. Freiman, R. Dixon and Y. Cossart (1992). Effects of Phyllanthus plant extracts on duck hepatitis B virus in vitro and in vivo, *Antivir. Res.* **18**(2), 127-138.
- [11] M. Saxena, J. Saxena, R. Nema, D. Singh and A. Gupta (2013). Phytochemistry of medicinal plants, *J. Pharmacogn. Phytochem.* **1**(6), 168-182.
- [12] N. Lajis, O. B. Guan, M. Sargent, B. Skelton and A. White (1992). Viroallosecurinine and entphyllanthidine from the leaves of *Breynia coronata* (Euphorbiaceae), *Aust. J. Chem.* **45**(11), 1893-1897.
- [13] S. Dall, K. I. Sinan, I. Ferrarese, S. Sut, K. Bene, M. F. Mahomoodally, N. Bibi Sadeer, G. Ak and G. Zengin (2020). Chromatographic separation of *Breynia retusa* (Dennst.) Alston bark, fruit and leaf constituents from bioactive extracts, *Molecules* **25**(**23**), 5537.
- [14] G. Fu, Z. Xu, B. Yu and D. Zhu (2004). Studies on the chemical constituents from the aerial parts of *Breynia fruticosa, Chin. Med. J.* **29**(**11**), 1052-1054.
- [15] M. Yuldashev, É. K. Batirov, V. Malikov and P. K. Yuldashev (1993). Acylated flavanone glycosides from *Ricinus communis, Chem. Nat. Compd.* 29(3), 303-305.
- [16] B. D. Zhang, J. X. Cheng, C. F. Zhang, Y. D. Bai, W. Y. Liu, W. Li, K. Koike, T. Akihisa, F. Feng and J. Zhang (2020). *Sauropus androgynus* L. Merr. A phytochemical, pharmacological and toxicological review, J. Ethnopharmacol. 257, 112778.
- [17] D. Meng, J. Wu and W. Zhao (2010). Glycosides from *Breynia fruticosa* and *Breynia rostrata*, *Phytochemistry* **71**(2-3), 325-331.
- [18] W. W. Peng, Z. Q. Wang, M. Y. Ji, Z. L. Liao, Z. Q. Liu and P. Wu (2017). Tyrosinase inhibitory activity of three new glycosides from *Breynia fruticosa*, *Phytochem. Lett.* **22**, 1-5.
- [19] P. Picerno, T. Mencherini, L. Rastrelli, A. Piccinelli and R. Aquino (2008). Isoprenoid glycosides from *Liriosma ovata, J. Nat. Prod.* 71(2), 265-268.
- [20] İ. Çalış, A. Kuruüzüm, P. A. Lorenzetto and P. Rüedi (2002). (6S)-Hydroxy-3-oxo-α-ionol glucosides from *Capparis spinosa* fruits, *Phytochemistry* **59**(**4**), 451-457.
- [21] H. Morikawa, R. Kasai, H. Otsuka, E. Hirata, T. Shinzato, M. Aramoto and Y. Takeda (2004). Terpenic and phenolic glycosides from leaves of *Breynia officinalis, Chem. Pharm. Bull.* **52**(**9**), 1086-1090.
- [22] M. Wang, J. Li, M. Rangarajan, Y. Shao, E. J. LaVoie, T. C. Huang and C. T. Ho (1998). Antioxidative phenolic compounds from sage (*Salvia officinalis*), *J. Agric. Food Chem.* **46**(**12**), 4869-4873.
- [23] L. Jiang, H. Kojima, K. Yamada, A. Kobayashi and K. Kubota (2001). Isolation of some glycosides as aroma precursors in young leaves of Japanese pepper (*Xanthoxylum piperitum DC.*), *J. Agric. Food Chem.* **49**(**12**), 5888-5894.
- [24] H. Otsuka, K. Kamada, C. Ogimi, E. Hirata, A. Takushi and Y. Takeda (1994). Alangionosides A and B, ionol glycosides from leaves of *Alangium premnifolium*, *Phytochemistry* **35**(**5**), 1331-1334.

- [25] T. Miyase, A. Ueno, N. Takizawa, H. Kobayashi and H. Oguchi (1988). Studies on the glycosides of *Epimedium grandiflorum* Morr. var. thunbergianum (Miq.) Nakai. III, Chem. Pharm. Bull. 36(7), 2475-2484.
- [26] N. Pauli, U. Séquin and A. Walter (1990). Boscialin and Boscialin 4'-O-Glucoside, Two new compounds isolated from the leaves of *Boscia salicifolia*, *Helv. Chim. Acta* **73**(**3**), 578-582.
- [27] C. Masuoka, M. Ono, Y. Ito, M. Okawa and T. Nohara (2002). New megastigmane glycoside and aromadendrane derivative from the aerial part of *Piper elongatum, Chem. Pharm. Bull.* **50**(10), 1413-1415.
- [28] Y. Lu and L. Y. Foo (2000). Flavonoid and phenolic glycosides from *Salvia officinalis, Phytochemistry* **55**(**3**), 263-267.
- [29] S. J. Ma, M. Mizutani, J. Hiratake, K. Hayashi, K. Yagi, N. Watanabe and K. Sakata (2001). Substrate specificity of β -primeverosidase, a key enzyme in aroma formation during oolong tea and black tea manufacturing, *Biosci. Biotechnol. Biochem.* **65**(12), 2719-2729.
- [30] A. M. Pawlowska, M. D. Leo and A. Braca (2006). Phenolics of *Arbutus unedo* L.(Ericaceae) fruits: Identification of anthocyanins and gallic acid derivatives, *J. Agric. Food Chem.* **54**(**26**), 10234-10238.
- [31] C. H. Li, X. D. Yang, J. F. Zhao and L. Li (2006). The chemical constituents of *Breynia rostrata, Yao Xue Xue Bao.* **41**(2), 125-7.
- [32] M. Takido, K. Fukuhara, S. Yamanouchi and S. Takahashi (1983). Phlebotrichin, a phenolic compound from the fresh leaves of *Viburnum phlebotrichum*, *Phytochemistry* **22**(**1**), 223-225.
- [33] T. M. Nguyen, X. T. Le and M. T. K. Nguyen (2017). Chemical constituents of *Breynia vitis-idaea* (Burm. f.) CEC Fischer in AIP Conference Proceedings, *AIP Publishing LLC*. **1878**(1), 020042 (1-7).
- [34] K. Sano, S. Sanada, Y. Ida and J. Shoji (1991). Studies on the constituents of the bark of *Kalopanax pictus Nakai, Chem. Pharm. Bull.* **39**(**4**), 865-870.
- [35] C. Li, X. Yang, J. Zhao and L. Li (2006). The chemical constituents of *Breynia rostrata*, Acta *Pharm. Sin. B.* **41(2)**, 125-127.
- [36] T. Fujita, K. Terato and M. Nakayama (1996). Two jasmonoid glucosides and a phenylvaleric acid glucoside from *Perilla frutescens, Biosci. Biotechnol. Biochem.* **60**(4), 732-735.
- [37] H. J. Kim, E. R. Woo and H. Park (1994). A novel lignan and flavonoids from *Polygonum aviculare*, *J. Nat. Prod.* **57**(**5**), 581-586.
- [38] B. Baderschneider and P. Winterhalter (2001). Isolation and characterization of novel benzoates, cinnamates, flavonoids, and lignans from Riesling wine and screening for antioxidant activity, *J. Agric. Food Chem.* **49**(**6**), 2788-2798.
- [39] S. T. Chang, S. Y. Wang and Y. H. Kuo (2003). Resources and bioactive substances from Taiwania (*Taiwania cryptomerioides*), J. Wood Sci. **49**(**1**), 0001-0004.
- [40] N. Matsuda and M. Kikuchi (1996). Studies on the Constituents of *Lonicera* Species. X. Neolignan Glycosides from the Leaves of *Lonicera gracilipes* var. glandulosa, Chem. Pharm. Bull. 44(9), 1676-1679.
- [41] T. Masamune, M. Anetai, A. Fukuzawa, M. Takasugi, H. Matsue, K. Kobayashi, S. Ueno and N. Katsui (1987). Glycinoeclepins, natural hatching stimuli for the soybean cyst nematode, *Heterodera glycines*. I. Isolation, *Bull. Chem. Soc. Jpn.* 60(3), 981-999.
- [42] R. Finkel, M. A. Clark and L. X. Cubeddu (2009). Pharmacology, Lippincott Williams & Wilkins.
- [43] J. Vanithamani and M. Kamaraj (2018). Screening of phytochemical constituents, trace metal concentrations and antimicrobial efficiency of *Breynia retusa*, **6**(**6**), 374-380.
- [44] M. Khan and A. Omoloso (2008). Antibacterial and antifungal activities of *Breynia cernua*, *Fitoterapia* 79(5), 370-373.
- [45] K. Laveena and M. Chandra (2018). Evaluation of bioactive compounds, antioxidant, and antibacterial properties of medicinal plants *Sauropus androgynus* L. and *Erythrina variegata* L, *Asian J. Pharm. Clin. Res.* **11**, 313-7.
- [46] V. Ariharan, V. M. Devi and P. N. Prasad (2013). Antibacterial activity of *Sauropus* and *Rogynous* leaf extracts against some pathogenic bacteria, *Rasayan J. Chem.* **6**(2), 134-137.
- [47] S. Selvi and A. Basker (2012). Phytochemical analysis and GC-MS profiling in the leaves of *Sauropus androgynus* (1) MERR, *Int. J. Drug Dev. Res.* **4**(1), 162-7.
- [48] S. Abid and S. Touqeer (2015). Antimicrobial and antioxidant activity of *Breynia disticha* and *Vernonia elaeagnifolia*, *J. Appl. Pharm.* **7**, 178-182.
- [49] F. A. Onyegbule, I. O. Louno, P. M. Eze, C. C. Abba and V. U. Chigozie (2014). Evaluation of the analgesic, anti-inflammatory and antimicrobial activities of leaf extracts of *Breynia nivosa, Chem. Sci. Rev. Lett.* **3(12)**, 1126-1134.
- [50] S. Dirgantara, R. H. Tanjung, H. K. Maury and E. Meiyanto (2018). Cytotoxic activity and phytochemical analysis of *Breynia cernua* from Papua, *Indones. J. Pharm.* 1(1), 31-36.

Saadullah et.al., Rec. Nat. Prod. (2022) 16:6 538-549

- [51] E. Bosisio, D. Mascetti and P. Caballion (2000). Screening of plants from New Caledonia and Vanuatu for inhibitory activity of xanthine oxidase and elastase, *Pharm. Biol.* **38**(1), 18-24.
- [52] Y. Bhagyasri, N. S. Subramanian and M. V. Akram (2017). Phytochemical studies and in-vitro antioxidant activity of *Breynia retusa* dennst, *World J. Pharm. Res.* 6(11), 641-651.
- [53] C. S. Madhu, H. M. G. Manukumar and P. Basavaraju (2014). New-vista in finding antioxidant and anti-inflammatory property of crude protein extract from *Sauropus androgynus* leaf, *Acta Sci. Pol. Technol. Aliment.* **13**(4), 375-383.
- [54] S. Mukherjee, K. B. S. Chouhan, R. Tandey, N. Yadav, M. Dhobi and V. Mandal (2021). A status report with critical analysis of research trends in exploring medicinal plants as antiviral: Let us dig into the history to predict the future, *Phytother. Res.* **35**(**8**), 4284-4296
- [55] M. I. Chowdury, M. H. Rahman, M. N. Alam, S. Chowdhury, M. S. Biozid, M. N. Bhuiyan and S. A. Alam (2015). Ex-vivo anti-arthritic and membrane stabilizing activity of aqueous extract of *Breynia retusa*, *World J. Pharm. Pharm. Sci.* (*WJPPS*). **4**(**9**), 89-95.
- [56] K. Kripa, R. Sangeetha, P. Madhavi and P. Deepthi (2011). Phytochemical screening and in vitro amylase inhibitory effect of the leaves of *Breynia retusa*, *Pak. J. Biol. Sci.* **14**(**19**), 894.
- [57] K. Sai and N. Srividya (2002). Blood glucose lowering effect of the leaves of *Tinospora cordifolia* and *Sauropus androgynus* in diabetic subjects, *J. Nat. Remedies* **2**(**1**), 28-32.
- [58] J. C. Nagar and L. S. Chauhan (2016). Evaluation of antihyperglycemic and antihyperlipidemic activity of leaf extracts of *Breynia vitis-idaea* in alloxan induced diabetic rats, *Pharmacogn. J.* **8**(3), 259-263.
- [59] H. Koshiyama, M. Hatori, H. Ohkuma, F. Sakai, H. Imanishi, M. Ohbayashi and H. Kawaguchi (1976). Breynins, new sulfur-containing glycosides with hypocholesterolemic activity, *Chem. Pharm. Bull.* 24(1), 169-172.
- [60] A. Gangula, R. Podila, L. Karanam, C. Janardhana and A. M. Rao (2011). Catalytic reduction of 4nitrophenol using biogenic gold and silver nanoparticles derived from *Breynia rhamnoides, Langmuir* 27(24), 15268-15274.
- [61] A. Godain, M. W. Spurr, H. C. Boghani, G. C. Premier, E. H. Yu and I. M. Head (2020). Detection of 4-nitrophenol, a model toxic compound, Using multi-stage microbial fuel cells, *Front. Environ. Sci.* 8, 5.

A C G publications