

Heptaelliptoic Acid A, A New Betulinic Acid Saponin from the Leaves of *Heptapleurum ellipticum*

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Abstract: One new 3-*O*-glycoside of betulinic acid, named heptaelliptoic acid A (1), together with one known betulinic acid analogue (2), and four other compounds (3–6) were separated by combinatively chromatographic techniques. For the first time, all of the purified compounds (1–6) were reported from the *H. ellipticum* species. Their structures were obviously elucidated basing exhaustive and pervasive UV-VIS, FT-IR, HR-MS-ESI, and NMR experiment data. Compounds 2-4 were significantly displayed the *in vitro* α -glucosidase inhibition (IC₅₀ values of 11.53, 28.75, and 10.90 μ M, respectively) better than the acarbose positive drug (IC₅₀ value of 214.50 μ M).

Keywords: Araliaceae; *Heptapleurum ellipticum*; heptaelliptoic acid A; α -glucosidase inhibition; betulinic acid saponin. © 2024 ACG Publications. All rights reserved.

1. Plant Source

Heptapleurum ellipticum (synonym: *Schefflera elliptica*) distributing fifty-six species in Vietnam is used as a traditional medicine [1-3]. The leaves of *H. ellipticum* collected at An Giang

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province, were scientifically characterization by Assoc. Prof. Dr. Van Son Dang, ITB. The herbal specimen in Kien Giang University was made with coded KGU/NGHIA-SE0523.

2. Previous Studies

In our published studies on the phytochemistry of this species, five triterpenoids, three flavonoids, and two cerebrosides were reported [3-4].

3. Present Study

The leaves of *H. ellipticum* (10 kg) were dried and extracted at room temperature in ethanol 96° to provide the total extract (SEEt). This extract (960 g) was respectively subjected to a liquid-liquid fractionation with *n*-hexane, and EtOAc solvents to deliver SEH (220 g) and SEE (150 g) extracts, along with an aqueous layer. The aqueous portion (700 g) was eluted by Diaion HP-20 column with H₂O:MeOH (0:100→100:0, v/v), and then gave five major fractions (I-V), respectively. Fraction IV (70 g) was subjected to silica gel column chromatography (CC) with mobile phase (EtOAc:MeOH) gradient (0→100%), and gave six sub-fractions (IV.1-IV.6). Fraction IV.1 (12 g) was chromatographed on silica gel with solvent system CHCl₃:MeOH:H₂O (90:10:0.1→80:20:0.2, v/v/v), and then separated by RP-18 using gradient mixtures of MeOH:H₂O (4:1, v/v) to give **1** (15 mg), **2** (20 mg). The SEE extract was objected to a normal-phase CC via a mixture of solvents containing *n*-hexane, EtOAc, and MeOH (the ratio of 25:75:0 to 0:80:20, v/v/v) to give six fractions, SEE.I-SEE.VI. The SEE.II fraction (27.0 g) was chromatographed on a silica gel column with the solvent system of *n*-hexane:EtOAc (80:20→30:70, v/v) to yield six sub-fractions, SEE.II.1-SEE.II.6. The SEE.II.1 sub-fraction (4.0 g) was separated on a normal-phase CC using CHCl₃:MeOH (99:1, v/v), and further on a reversed phase-C₁₈ CC using gradient mixtures of MeOH:H₂O (70:30, v/v) to obtain **3** (15.0 mg) and **4** (10 mg). Likewise, **5** (10 mg) and **6** (9 mg) were respectively yielded from sub-fractions SEE.II.6 (6.5 g) and SEE.II.2 (3.5 g).

Heptaelliptic acid A (1): Amorphous powder (MeOH); $[\alpha]_D^{25}$ -1.0 (c 0.1, MeOH). UV (MeOH, λ_{max} nm): 203. IR (KBr, ν_{max} , cm⁻¹): 3416, 2942, 2871, 1692, 1616, 1077, 1044. HR-ESI-MS m/z 773.4464 [M+Na]⁺ (calcd for C₄₁H₆₆O₁₂Na, 773.4452). ¹H NMR (600 MHz, methanol-d₄, J/Hz), and ¹³C NMR (125 MHz, methanol-d₄): see in Table 1.

Compound (**1**) was supplied as an amorphous powder. Its formular was determined as C₄₁H₆₆O₁₂ basing on the HR-ESI-MS data m/z 773.4464 [M+Na]⁺ (calcd for C₄₁H₆₆O₁₂Na⁺, 773.4452). The FT-IR spectroscopy of **1** clearly exhibited the specific absorptions of the hydroxyl (3416 cm⁻¹), carboxyl (1692 cm⁻¹), and C-O stretch (1044 cm⁻¹) functional groups. The ¹³C & ¹H NMR, and integrating HSQC spectrum of **1** (Table 1) certificated forty-one carbons comprising one carboxylic carbon at δ_C 181.7 (C-28), one olefinic carbon at δ_C 152.4 (C-20), one exocyclic methylene carbon at δ_C 109.9 (C-29)/ δ_H 4.70 (1H, d, J = 1.8 Hz, H-29a), 4.57 (1H, d, J = 1.2 Hz, H-29b), two anomeric carbons with one of them at δ_C 105.3 (C-1')/ δ_H 4.42 (1H, d, J = 7.8 Hz, H-1'), eight oxygenated methine carbons with five of them at δ_C 91.0 (C-3)/ δ_H 3.12 (1H, dd, J = 10.2, 4.2 Hz, H-3), 83.3 (C-2'), 78.3 (C-3'), 71.5 (C-4'), 77.5 (C-5'), two oxygenated methylene carbons with one of them at δ_C 62.7 (C-6'), five quaternary carbons, ten methylene carbons, five methine carbons, and six methyl carbons, were disclosed a betulinic acid skeleton bearing one 3-*O*- β -D-glucopyranosyl (Glc) unit likely those of 28-*O*- β -D-glucopyranosylbetulinic acid 3-*O*- β -D-glucopyranoside [5]. However, the presence of one anomeric carbons at δ_C 106.3 (C-1'')/ δ_H 4.51 (1H, d, J = 7.2 Hz, H-1''), three oxygenated methine carbons at δ_C 76.3 (C-2''), 77.8 (C-3''), 71.2 (C-4''), one oxygenated methylene carbon at δ_C 67.2 (C-5'')/ δ_H 3.80 (1H, dd, J = 11.4, 5.4 Hz, H-5''a), 3.13 (1H, dd, J = 10.8, 6.6 Hz, H-5''b), and the multiplicities and large coupling constants of proton H-4'' at δ_H 3.45 (1H, ddd, J = 10.2, 8.4, 5.4 Hz) were clearly distinguished one *O*- β -D-xylopyranosyl (Xyl) unit, by comparising with those of a Xyl

moiety in 2 α -hydroxy-3 β -[(*O*- β -D-xylopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosyl)oxy]lup-20(29)-en-28-oic acid α -L rhamnopyranosyl ester (Table 1) [6], and those of Arabinopyranosyl (Ara) unit [δ_C 107.1 (C-1''), 72.8 (C-2''), 74.3 (C-3''), 69.5 (C-4'')/ δ_H 3.82 (1H, dd, $J = 3.5, 3.0$ Hz), 66.3 (C-5'')] in chenoalbusoside A [7]. Furthermore, the R_f comparison between the aqueous layer from the hydrolysis of compound **1** and the D-glucose, D-xylose authentic standards by TLC.

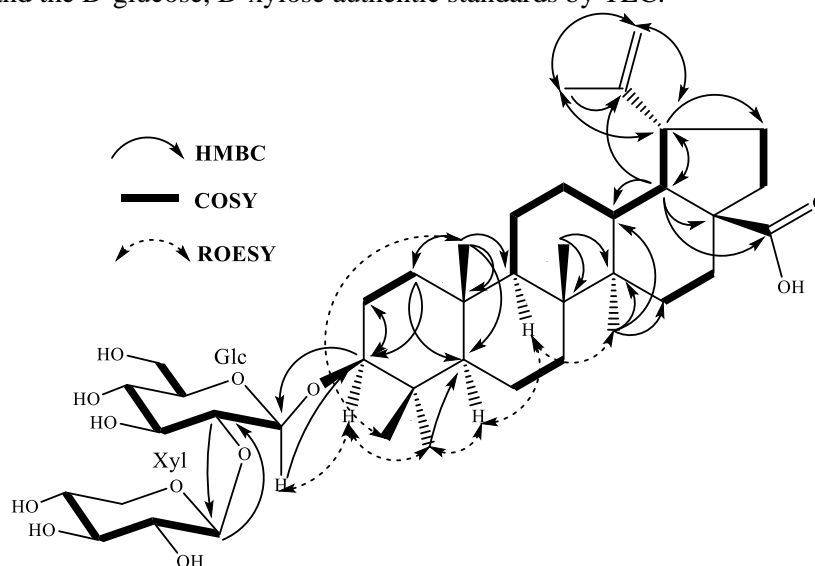


Figure 1. Selected COSY, HMBC, and ROESY correlations of **1**

Moreover, the HMBC spectrum of **1** (Figure 1) displayed two anomeric protons at δ_H 4.42 (1H, d, $J = 7.8$ Hz, H-1') and 4.51 (1H, d, $J = 7.2$ Hz, H-1'') respectively correlated with two oxymethine carbons at δ_C 91.0 (C-3) and 83.3 (C-2'), which were plainly recognized the glycoside chain of **1** was 3-*O*- β -D-Xyl-(1 \rightarrow 2)- β -D-Glc, instead of the 3-*O*- α -L-Ara-(1 \rightarrow 2)- β -D-Glc sugar chain in coccinioside-K [8]. On the other hand, the α -oriented methyl protons δ_H 1.05 (3H, s, H-23), and methine proton δ_H 0.73 (1H, d, $J = 10.8$ Hz, H-5) correlated with oxymethine proton δ_H 3.12 (1H, dd, $J = 10.2, 4.2$ Hz, H-3) in the ROESY spectroscopy of **1** (Figure 1), together with the doublet of doublet splitting and large-small coupling constants ($J = 10.2, 4.2$ Hz) of proton (H-3), which were significantly affirmed a hydroxyl moiety at carbon C-3 was 3β orientation. Based on data of HRMS-ESI, 1D & 2D-NMR, and comparison to the published spectral data [5-6], the structure of **1** was designated as 3-*O*- β -D-xylopyranosyl-(1 \rightarrow 2)- β -D-glucopyranosylbetulinic acid, and trivially named heptaelliptic acid A.

The HRMS-ESI, NMR data of isolated compounds were consistent with those in the published papers for 3-*O*- β -D-xylopyranosyl-(1 \rightarrow 2)- β -D-glucuronopyranosylbetulinic acid (**2**) [9], and four known compounds, oleanolic acid (**3**) [10], ursolic acid (**4**) [11], 3-*O*- β -D-glucopyranosylstigmasterol (**5**) [12], and 3'-*O*-methyluteolin (**6**) [13] (Figure 2).

Hydrolysis of glycoside: The acid hydrolysis of new compound **1** was detailed reported papers in the literature [3,14].

α -Glucosidase Inhibition Assay: The *in vitro* α -glucosidase inhibitory activities of all isolated compounds (**1-6**) were examined as our published method [3]. As the results, the isolated compounds **2-4** showed meaningfully better α -glucosidase inhibition (IC_{50} values of 11.53, 28.75, and 10.90 μ M, respectively) than the acarbose control (IC_{50} values of 214.50 μ M), whereas compounds **1, 5** and **6** did not exhibit activity (Table S1). Additionally, these results were completely consistent with the α -glucosidase inhibitions of those compounds that were previously evaluated [15, 16].

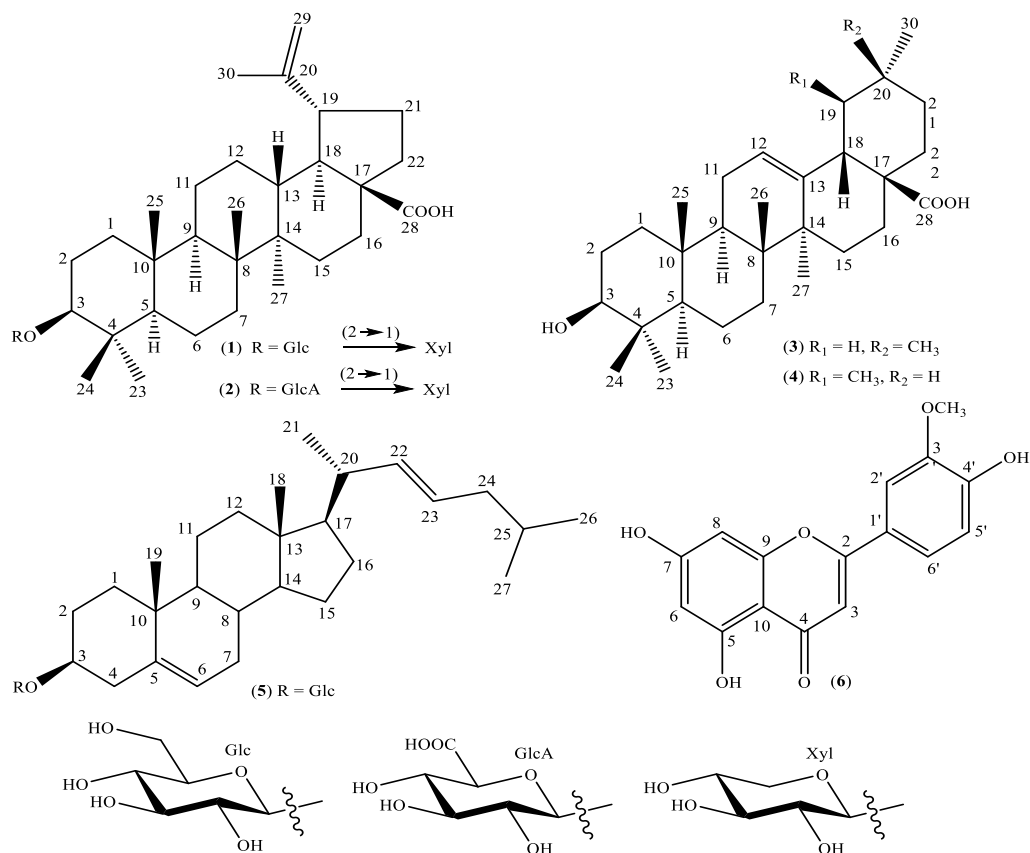
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Figure 2. Chemical structures of 1-6

Table 1. NMR spectral data for compound **1** in methanol-d₄ (δ in ppm, J in Hz)

No.	1	
	δ_{H}	δ
1	1.69 (1H, m)	40.1
	0.92 (1H, m)	
2	1.94 (1H, m)	27.3
	1.70 (1H, m)	
3	3.12 (1H, dd, $J = 10.2, 4.2$)	91.0
4	-	40.4
5	0.71 (1H, d, $J = 10.8$)	57.3
6	1.55 (1H, m)	19.3
	1.42 (1H, m)	
7	1.42 (1H, m)	35.7
8	-	42.0
9	1.30 (1H, m)	52.1
10	-	38.1
11	1.41 (1H, m)	22.2
	1.26 (1H, m)	
12	1.05 (1H, m)	27.0
	1.71 (1H, m)	
13	2.39 (1H, td, $J = 12.0, 0.6$)	39.6
14	-	43.6
15	1.15 (1H, m)	31.0
	1.56 (1H, m)	
16	2.23 (1H, d, $J = 12.0$)	33.8

1.39 (1H, m)		
17	-	57.3
18	1.61 (1H, m)	50.7
19	3.07 (1H, td, $J = 12.0, 3.0$)	48.5
20	-	152.4
21	1.92 (1H, m)	31.9
	1.34 (1H, m)	
22	1.92 (1H, m)	38.5
	1.40 (1H, m)	
23	1.03 (3H, s)	28.2
24	0.81 (3H, s)	16.4
25	0.86 (3H, s)	16.8
26	0.98 (3H, s)	16.8
27	0.99 (3H, s)	15.1
28	-	181.7
29	4.70 (1H, d, $J = 1.8$)	109.9
	4.57 (1H, d, $J = 1.2$)	
30	1.69 (3H, s)	19.6
3-O-Glc		
1'	4.42 (1H, d, $J = 7.8$)	105.3
2'	3.42 (1H, dd, $J = 9.0, 7.8$)	83.3
3'	3.53 (1H, m)	78.3
4'	3.32 (1H, m)	71.5
5'	3.22 (1H, m)	77.5
	3.84 (1H, m)	
6'	3.67 (1H, dd, $J = 12.0, 5.4$)	62.7
Xyl		
1''	4.51 (1H, d, $J = 7.2$)	106.3
2''	3.21 (1H, m)	76.3
3''	3.34 (1H, m)	77.8
4''	3.45 (1H, ddd, $J = 10.2, 8.4, 5.4$)	71.2
	3.80 (1H, dd, $J = 11.4, 5.4$)	
5''	3.13 (1H, dd, $J = 10.8, 6.0$)	67.2

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Supporting Information

Supporting Information accompanies this paper on <http://www.acgpubs.org/journal/records-of-natural-products>

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