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Essential Oil Compositions and Biological Activities of the Genus Chaerophyllum: An Updated Review

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Abstract: The genus Chaerophyllum has economic value-added species. Chaerophyllum belongs to the family, Apiaceae, which comprises about 110 species, including annual and perennial herbal plants widely distributed in temperate and sub-temperate zones of Asia, Africa, and Europe. Chaerophyllum species are used for flavor and fragrance in the food industry. In some countries, a species of this genus is used in cheese production for its strong aroma. Chaerophyllum species were characterized by lignans, polyacetylenes, essential oils, phenolic acids, and flavone derivatives. This review summarizes the essential oil components and isolates of various extracts and their biological activities. In addition, since ancient times, essential oils have been used in many different traditional healing systems worldwide because of their biological activities. Moreover, this review will attract the attention of scientists from the aroma industry, nutritionists, and pharmaceutical industries to improve the use of essential oils for nutraceutical purposes with commercialization to aid and promote a healthy lifestyle, wellness, and wellbeing.

Keywords: Apiaceae; biological activities; essential oils; *Chaerophyllum villosum*. © 2025 ACG Publications. All rights reserved.

1. Introduction

The genus *Chaerophyllum* L. belongs to the family Apiaceae, which includes annual and perennial herbal plants widely distributed in temperate and sub-temperate zones of Asia, Africa, and Europe [1, 2]. The Apiaceae family, including some of the widely used vegetables and aromatic herbs such as celery (*Apium graveolens* L.), carrot (*Daucus carota* L.), gotukola (*Centella asiatica* (L.) Urb), parsley (*Petroselinum crispum* L.), parsnip (*Pastinaca sativa* L.), wild celery (*Angelica archangelica* L.), coriander (*Coriandrum sativum* L.), cumin (*Cuminum cyminum* L.), fennel (*Foeniculum vulgare* Mill.), anise (*Pimpinella anisum* L.), dill (*Anethum graveolens* L.), and caraway (*Carum carvi* L.) are the economically important foods, herbs, and spice plants in the Apiaceae family [3-7]. In the flora of Türkiye, the genus *Chaerophyllum* L. is represented by 15 species, of which four are endemic [1]. From the genus *Chaerophyllum*, out of 72 species, one species from India was reported as *Chaerophyllum villosum* Wall. & DC., is widely distributed in the Himalayas at elevations of 2500 m to 3500 m, from India to Bhutan, Nepal, and China, and widely grows in moist, shady places, roadsides, or open grassy

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places [8]. Chaerophyllum species are used to add flavor and fragrance in the food industry. In the high-altitude tribes of Uttarakhand Himalaya (India), it was commonly known and sold in the name of 'Ganjari or Ganjiadi' and widely used by people in food, spice, and medicine [9]. Chaerophyllum macropodum Boss. (C. macropodum) is traditionally mixed into an herbal cheese, which is a famous dairy product known for its aroma and flavor in Türkiye [10-11]. C. macropodum (local name: mendi, mendo) is widely grown in Van and is used in the production of a very famous cheese known as "Van Otlu Peynir" [12, 13]. In the flora of Iran, the genus consists of 8 species of biennial or perennial herbs [14]. The seeds, leaves, and roots of the Chaerophyllum species have a carrot-parsley aroma and are sometimes used as flavoring agents in Iran, Türkiye, and Caucasian and Eastern Europe. C. macrospermum is the most widespread species of the genus in Iran, where it is known as Djafari-Farangi [15]. The main objective of this review is to focus on the components of essential oils, biological activities, and other nutraceutical potentials of essential oils and major isolates of the genus Chaerophyllum.

2. Taxonomy of the Genus Chaerophyllum

The scientific taxonomy of the Apiaceae started using morphological criteria, but afterwards, there were new diagnostic elements from several disciplines: anatomy, karyology, seedling morphology, palynology, and phytochemistry. The genus *Chaerophyllum* L. has economic value-added species. The Apiaceae, or plants of the 'Carrot Family', have been known by man since ancient times. Many local plants of this family were used in primitive cultures because people soon noticed their odor, flavor, esculence, or toxicity. Various Apiaceae are known in the early languages of China and in Sanskrit [16]. Cesalpino (1583) made the first overall grouping as 'Universum genus Ferulaceum' which comprised about 60 herbs [17]. The 'Umbelliferae' were born.

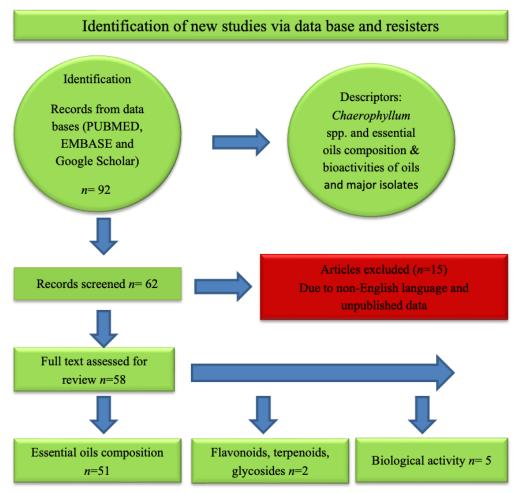


Figure 1. Flow chart showing the selection of the studies

The first monograph of a plant group independent of their uses was carried out by Morison (1672), who chose the Umbelliferae as a model, where he proposed a classification based mainly on fruit morphology [18].

3. Material and Method

This current review focuses on an update on recent studies performed on the chemical composition and biological studies on the genus *Chaerophyllum*. The search engines Google Scholar, PubMed, ScienceDirect, and ResearchGate were used to access the literature. Relevant articles published in English were searched in different databases, specifically Google Scholar, Scopus, Wiley Online Library, Semantic Scholar, and Pub Med (Fig.1). The following terms were used in combination within search strings: "*Chaerophyllum*", "essential oils", "anti-inflammatory", "antioxidant", "antibacterial", "antifungal", "antimicrobial", and "antiviral". The search was restricted to English and experimental studies.

Table 1. Major component from different species of genus Chaerophyllum.

| Chaerophyllum species | Collection Site | Parts of plant used | Major components present | Ref. |
|---|--------------------|---------------------|---|------|
| C. aksekiense A. Duran & H. Duman | Türkiye | Crushed fruits | Heptacosane (10.1%), humulene epoxide II (7.8%), (E)- β -farnesene (6.2%), caryophyllene oxide (6.0%), α -humulene (5.5%), terpinolene (5.5%), nonacosane (5.3%), and terpinen-4-ol (4.6%) | [21] |
| C. aromaticum L. | Türkiye | Aerial parts | Sabinene (28.1%), terpinolene (16.7%) and γ -terpinene (16.1%) | [22] |
| C. aureum L. | Serbia | Fruits | Sabinene (18.5-31.6%), <i>p</i> -cymene (7.9-25.4%), and limonene (1.9-10.9%) | [23] |
| C. azoricum Trel. | Portugal | Leaves & stems | Terpinolene (44-62%) and γ-terpinene (9-31%) of the oil | [24] |
| C. byzantinum Boiss. | Türkiye | Aerial parts | Sabinene (30.0%) and <i>p</i> -cymen-8-ol (16.0%) | [25] |
| C. bulbosum L. ssp. bulbosum) | Greece | Whole plant | Apiol (37%), (<i>E</i>)-nerolidol (8.5%), linalool (7.7%), myristicin (6.9%), and eugenol (5.8%) | [26] |
| | Iran | Aerial parts | (<i>E</i>)-β-Farnesene (22.3%), (<i>Z</i>)-β-ocimene (18.8%), myristicin (17.1%), caryophyllene oxide (6.6%), <i>allo</i> -ocimene (5.1%), and (<i>E</i>)-β-ocimene (4.0%) | [27] |
| C. coloratum L. | Iran | Ripe fruits | (E)-β-Farnesene (79.2%), β-pinene (7.0%), and (Z)-β-ocimene (4.3%) | [28] |
| | | Umbel | (E)-β-Farnesene (68.5%) and (Z)-β-ocimene (5.5%) | [29] |
| C. libanoticum Boiss. & Kotschy | Türkiye | Crushed fruits | β-Phellandrene (17.6%), limonene (15.9%), β-pinene (8.8%), and sabinene (8.5%) | [30] |
| C. macrospermum (Willd. ex Schult.) Fisch. & C.A. Mey | Iran | Inflorescences | 1,8-Cineole (7.2%), linalool (6.7%), δ-3- carene (4.4%), α-terpineol (4.7%), and farnesol (4.0%) | [31] |
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| | Iran | Aerial parts | (<i>E</i>)-β-Ocimene (40.0%), tricyclene (19.4%), δ-3-carene (18.3%), and myrcene (10.1%) | [31] |
|----------------------------|---------|------------------------|--|-------------|
| | Iran | Aerial parts | (E)-β-Ocimene (55.9%), terpinolene (9.8%), α-pinene (7.5%), β-phellandrene (4.3%), and β-pinene (4.2%) | [32] |
| | Iran | Seeds | (E)-β-Farnesene (27.1%), (Z)-β-ocimene (18.8%), p-cymene (14.3 %), α-fenchyl acetate (12.7%), and spathulenol (8.8%) | [33] |
| | Iran | Aerial parts | Terpinolene (21.4%), myristicin (18.9%), <i>p</i> -cymen-8-ol (11.9%) | [34] |
| | Iran | Aerial parts | α-Pinene (23.0%), β-pinene (17.3%), and fenchyl acetate (13.8%) | [34] |
| C. macropodum Boiss. | Iran | Flowering aerial parts | (E)-β-Ocimene (34.5 %), (E)-β-Farnesene (11.8 %), (Z)-β-ocimene (10.4 %), p-cymene (7.3 %) and spathulenol (6.5 %) | [35] |
| | Türkiye | Fruits | <i>p</i> -Cymene (39.3%), spathulenol (7.3%), <i>p</i> -cymen-8-ol (5.9%), octanal (5.2%), and (<i>E</i>)-β-ocimene (4.5%) | [36] |
| | | Flowers, | Myristicin (42.5%) and (<i>E</i>)- β -ocimene (41.0%) | [37] |
| | Iran | Leaf | (<i>E</i>)-β-Ocimene (24.9%), myristicin (15.7%), terpinolene (14.5%), fenchyl acetate (13.9%), (<i>Z</i>)-β-ocimene (6.3%), and sabinene (6.1%) | |
| | | Stem | (<i>E</i>)-β-Ocimene (54.2%), myristicin (22.4%), and sabinene (8.9%) | [38- 39] |
| C. prescotti DC. | Iran | Root | Myristicin (39.2 %), terpinolene (23.1 %), (<i>E</i>)-β-ocimene (21.9 %) and γ-terpinene (5.4 %). | [40] |
| | Iran | Aerial parts | (<i>E</i>)-β-Ocimene (29.3 and 29.3%), myrcene (14.5 and 14.6%), and terpinolene (14.4 and 14.3%) | [40] |
| | Siberia | Flowering tops | (E)-β-Ocimene (35.6%), (Z)-β-ocimene (19.4%), γ-terpinene (18.8%), myrcene (10.6%), and terpinolene (4.6%) | [40] |
| .C. villosum Wall. & DC | India | Root | Carvacryl methyl ether (31.1%), myristicin (19.1%), thymyl methyl ether (18.6 %), and γ-terpinene (11.7%) | [41, 42] |
| | India | Leaf | γ-Terpinene (74.9%), <i>p</i> -cymene (10.0%), terpinolene (2.9%) and β-pinene (2.5%) | [43] |

4. Essential Oil Compositions of the Genus Chaerophyllum

Essential oils and natural volatiles are one of the industry's most successful commodities since they are used as flavors in lotions and shampoos, as fragrances and for skin and hair rejuvenation as perfumes, in candles, soaps, liquids that sterilize, and in conventional or medical aromatherapy. The oils are in different plant parts such as roots, stems, leaves, flowers, fruits, and even seeds, depending on the plant species. In these plant parts, the essential oil accumulates in cells, secretory cavities, or glandular hairs [19, 20]. There are several publications on *Chaerophyllum* essential oil compositions from various geographical locations, which are summarized in Table 1 and structures of major compounds, are shown in Fig. 2.

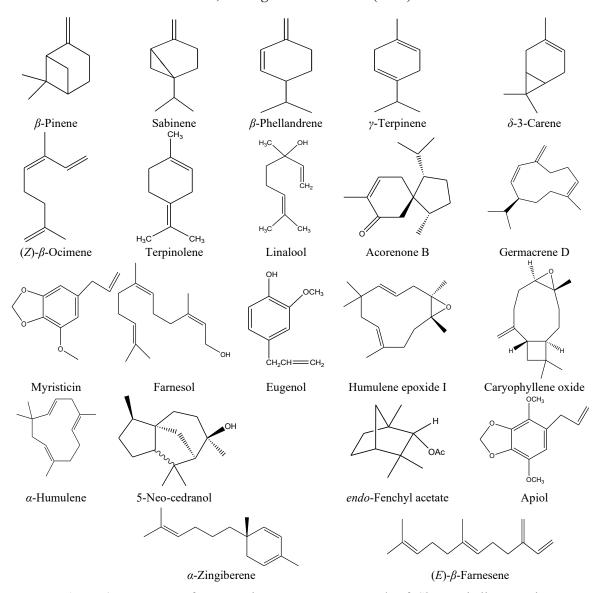


Figure 2. Structures of some major common compounds of Chareophyllum species

5. Phytochemical Constituents Isolated from Different Extracts

Plants contain a large number of bioactive constituents such as alkaloids, tannins, flavonoids, terpenoids, glycosides, etc [44-46]. *C. villosum* were collected from Miranjani top Pakistan and subjected to methanolic, chloroform, and aqueous extract and investigated for bioactive constituents and reported adequate presence of alkaloids, amino acids, saponins, tannins, and glycosides and also showed moderate presence of reducing sugar, fats, oils, and phytosterols. Fresh leaves of *C. byzantinum* were collected from Samsun, Türkiye. The samples were extracted with 80% methyl alcohol and the total phenolic content (TPC) was determined by measuring the phenolic compounds with a Folin-Ciocalteu solution in an alkaline medium. The results of analysis on leaves of the plants were found as; dry matter $16.43 \pm 0.41\%$, ash $2.03 \pm 0.20\%$, crude cellulose $1.95 \pm 0.23\%$, crude protein $4.36 \pm 0.31\%$, crude fat $0.57 \pm 0.16\%$, total phenolics 2890.15 ± 945.33 mg/kg. From the roots of *C. bulbosum*, which were collected from Türkiye, three known compounds were isolated from hexane extracts as *n*-heptadecanyl eicosanoate, stigmasterol, and β -sitosterol-3-*O*- β -D-glucopyranoside [47-48].

6. Biological Activities

Essential oils play a major role in plants and act as antibacterials, antivirals, antifungals, and insecticides and protect the plants from herbivores. Essential oils are a complex mixture of molecules, which generally contain more than twenty different components of low molecular weight with very

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variable concentrations. In general, monoterpenes and sesquiterpenes are the main components of essential oils, though diterpenes and phenylpropanoids can be present to a different extent. Many of these molecules are found in low concentrations, while few of them are the main components that can represent up to 70% of total oil and will be the main ones responsible for the biological effects of the oil [49-50]. Great efforts are performed to investigate the potential therapeutic effects of oils against several diseases, especially those characterized by excessive cell growth and proliferation such as cancer or bacterial infections [51-53]. Some biological activities shown by different *Chaerophyllum* species are summarized in Table 2.

Table 2. Different types of bioactivities shown by different species of *Chaerophyllum*.

| Chaerophyllum species | Different biological activities | Ref. |
|-------------------------|---|------|
| C. aureum L. | Antibacterial activity was evaluated against two Gram-positive and three Gram-negative bacteria. The Gram-positive bacteria used were: <i>Bacillus subtilis</i> and <i>Staphylococcus aureus</i> . The Gram-negative bacteria utilized in the assay were <i>Escherichia coli, Pseudomonas aeruginosa,</i> and <i>Salmonella abony</i> . The results of antibacterial assay showed that the oil was not active at concentration of 3 and 5 mg per disk. | [54] |
| C. bulbosum L. | The essential oil was evaluated for its in vitro antibacterial activity against two Gram-positive and three Gram-negative bacteria by disk diffusion assay. The obtained results showed that essential oil isolated from dry inflorescences of <i>C. bulbosum</i> had significant antibacterial activity against Gram-positive bacteria, <i>Staphylococcus aureus</i> and <i>Bacillus subtilis</i> subsp. <i>spizizenii</i> (diameter of inhibition zone 24 mm and 21 mm, respectively) and moderate antibacterial activity against <i>Escherichia coli</i> , <i>Salmonella abony</i> and <i>Pseudomonas aeruginosa</i> with inhibition zones of 17 mm, 15 mm, and 14 mm, respectively. | [55] |
| C. macropodum Boiss. | The essential oil of <i>Chaerophyllum macropodum</i> was tested for inhibition of ACE which is regarded as fundamental of hypertension treatment. <i>Chaerophyllum macropodum</i> have shown inhibitory effects on human plasma ACE with an IC ₅₀ value of 1.14 mg/mL. | [56] |

7. Conclusions

Species of Chaerophyllum have been used as traditional herbal medicines. This review demonstrated the high chemical and biological potential of essential oils from their species. C.bulbosum from Greece, Iran, Türkiye and Serbia is dominated by apiol, (E)- β -farnesene, (Z)- β -ocimene and myristicin. C. macrospermum reported from Iran was dominated by (E)- β -ocimene, tricyclene, δ -3carene, (E)-β-farnesene, and (Z)-β-ocimene. C. macropodum essential oils from Iran and Türkiye were dominated by α -pinene, β -pinene, p-cymene myristicin, (E)- β -ocimene, and (Z)- β -ocimene. C. crinitum from Türkiye and Iran was dominated by (E)- β -ocimene, and α -terpinolene, and some other species including C. aureum, C. prescotti, C. byzantinum, C. libanoticum, C. villosum, C. temulum, C. hirsutum, and C. aromaticum from different regions were dominated by some common components like sabinene, (E)- β -ocimene, β -phellandrene, carvacryl methyl ether, γ -terpinene, (Z)-falcarinol, and acorenone B. Chaerophyllum species are also used as flavor and fragrance additives in the food industry and also used as spices. This review shows that the genus Chaerophyllum is very rich in beneficial chemical components in essential oils and in different extracts. Essential oils and extracts exhibit significant biological and pharmacological activities. Several species are very useful in the present scenario for daily life in fragrance and perfumery. As well as in aromatherapy and many other purposes, nowadays, aromatic plants and essential oils are used in the cosmetic industries. We can conclude that the genus

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Chaerophyllum is very useful in many ways in daily life, so more studies and commercial production of essential oils will be useful to society, perfume, cosmetics, and pharma industries.

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References

- [1] P. H. Davis (1972). Flora of Turkey and the East Aegean Islands, Edinburgh at the University Press, Volume 4, Edinburgh.
- [2] H. Duman (2000). *Chaerophyllum* L. Flora of Turkey and the East Aegean Islands, University Press, (Edinburgh), Volume II. A3.
- [3] M. G. Acimovic (2019). Nutraceutical Potential of Apiaceae. In Bioactive Molecules in Food, Mérillon, J. M., Ramawat, K. G., Eds.; Reference Series in Phytochemistry; Springer International Publishing: Cham, Switzerland, pp. 1311–1341.
- [4] M. G. Acimovic and N. B. Milicb (2017). Perspectives of the Apiaceae hepatoprotective effects—A Review, *Nat. Prod. Commun.* **12**, 309–317.
- [5] D. Stesevic, M. Bozovic, V. Tadic, D. Rancic and Z. D. Stevanovic (2016). Plant-part anatomy related composition of essential oils and phenolic compounds in *Chaerophyllum coloratum*, a Balkan endemic species, *Flora* **220**, 37-51.
- [6] V. Vajs, S. Milosavljevic, V. Tesevic, P. Zivanovic, R. Jancic, B. Todorovic and V. Slavkovska (1995). *Chaerophyllum coloratum* L.: Essential oils of ripe fruits and Umbels, *J. Ess. Oil Res.* **5**(7), 529-531.
- [7] T. Üstüner, K. Kordali and A. Usanmaz Bozhüyük (2018). Herbicidal and fungicidal effects of *Cuminum cyminum*, *Mentha longifolia* and *Allium sativum* essential oils on some weeds and fungi, *Rec. Nat. Prod.* 12(6), 619-629.
- [8] O. Polunin and A. Stainton (1984). Flowers of the Himalaya, Oxford Press, New Delhi.
- [9] B. M. Sharma, S. Pratap and C. K. Atal (1969). Pharmacognosy of root of *Chaerophyllum villosum*, *Indian J. Med. Res.* **4**, 68-72.
- [10] N. Coruh, A. G. Sagdicoglu Celep and F. Ozgokce (2007). Antioxidant properties of *Prangos ferulacea* (L.) Lindl. *Chaerophyllum macropodum* Boiss. and *Heracleum persicum* Desf. from Apiaceae family used as food in Eastern Anatolia and their inhibitory effects on glutathione-S-transferase, *Food Chem.* **100** (3), 1237-1242.
- [11] A. H. Ebrahimabadi, Z. Djafari-Bidgoli, A. Mazoochi, F. J. Kashi and H. Batooli (2010). Essential oils composition, antioxidant and antimicrobial activity of the leaves and flowers of *Chaerophyllum macropodum* Boiss, *Food Control*, **21**, 1173-1178.
- [12] Z. Tarakci, E. Sagun and H. Durmaz (2006). The effect of mendi (*Chaerophyllum* sp.) on ripening of vacuum-packed herby cheese, *Int. J. Dairy Technol.* **59**(1), 35-39.
- [13] S. E. Celik, M. Ozyurek, M. Altun, B. Bektasoglu, K. Guclu, K.I. Berker, F. Ozgokce and R. Apak (2008). Antioxidant capacities of herbal plants used in the manufacture of Van herby cheese 'Otlu peynir', *Int. J. Food Prop.* **11(4)**, 747-761.
- [14] A. Shafaghat (2009). Antibacterial activity and composition of essential oils from flower, leaf and stem of *Chaerophyllum macropodum* Boiss. from Iran, *Nat. Prod. Commun.* **4(6)**, 861-864.
- [15] K. H. Rechinger (1987). Ferulago, In: *Flora Iranica, Umbelliferae*. No. 162. Edits., K. H. Rechinger and I. C. Hedge, p. 430, Akademische Druck and Verlagsanstalt, Graz, Austria.
- [16] L. Constance (1971). History of the classification of Umbelliferae (Apiaceae), J. Linn. Soc. 64, 1–11.
- [17] A. Rustaiyan, N. Neekpoor, M. Rabani, H. Komeilizadeh, S. Masoudi and A. Monfared (2002). Composition of the essential oil of *Chaerophyllum macrospermum* (Spreng.) Fisch. and CA Mey. from Iran, *J. Ess. Oil Res.* **14(3)**, 216-217.
- [18] N. J. Sadgrove (2018). The new paradigm for androgenetic alopecia and plant-based folk remedies: 5α-reductase inhibition, reversal of secondary microinflammation and improving insulin resistance, *J. Ethnopharmacol*, **227**, 206–236.
- [19] N. J. Sadgrove and M. S. J. Simmonds (2021). Topical and nutricosmetic products for healthy hair and dermal antiaging using dual-acting (2 for 1) plant-based peptides, hormones, and cannabinoids, *FASEB Bioadv.* **3**, 601–610.
- [20] K. H. C. Baser (2010). Handbook of Essential Oils: Science, Technology and Applications. University of Wien, Vienna, Austria, 978-1-4200-6315-8.

- [21] K. H. C. Baser, N.Tabanca, T. Ozek, B. Demirci, A. Duran and H. Duman (2000). Composition of the essential oil of *Chaerophyllum aksekiense* A. Duran et Duman, a recently described endemic from Turkey, *Flavour Fragr. J.* **15**(1), 43-44.
- [22] M. Kurkcuoglu, A. Sen, L. Bitis, S. Birteksoz Tan, A. Dogan and K. H. C. Baser (2018). Chemical composition, anti-inflammatory, antioxidant and antimicrobial activity of essential oil from aerial parts of *Chaerophyllum aromaticum* L. from Turkey, *J. Ess. Oil Res.* 21(2), 563-569.
- [23] B. Lakusic, V. Slavkovska, M. Pavlovic, M. Milenkovic, J. A. Stankovic and M. Couladis (2009). Chemical composition and antimicrobial activity of the essential oil from *Chaerophyllum aureum* L. (Apiaceae), *Nat. Prod. Commun.* **4(1)**, 115-118.
- [24] L. G. Pedro, J. A. da Silva, J. G. Barroso, A. C. Figueiredo, S. G. Deans, A. Looman and J. J. Scheffer (1999). Composition of the essential oil of *Chaerophyllum azoricum* Trel., an endemic species of the Azores archipelago, *Flavour Fragr. J.* **14(5)**, 287-289.
- [25] M. Kurkcuoglu, K. H. C. Baser, G. Iscan, H. Malyer and G. Kaynak (2006). Composition and anticandidal activity of the essential oil of *Chaerophyllum byzantinum* Boiss, *Flavour Fragr. J.* 21(1), 115-117.
- [26] E. Kokkalou and E. Stefanou (1989). The volatiles of *Chaerophyllum bulbosum* L. ssp. *bulbosum* growing wild in Greece, *Pharm. Acta Helv.* **64(5-6)**, 133-134.
- [27] S. Masoudi, A. Faridchehr, S. Alizadehfard, N. Zabarjadshiraz, F. Chalabian, R. Taghizadfarid and A. Rustaiyan (2011). Chemical composition and antibacterial activity of the essential oils of *Semenovia frigida* and *Chaerophyllum bulbosum* from Iran, *Chem. Nat. Compd.* 47, 829-832.
- [28] V. Vajs, S. Milosavljevic, V. Tesevic, P. Zivanovic, R. Jancic, B. Todorovic and V. Slavkovska (1995). *Chaerophyllum coloratum* L.: Essential oils of ripe fruits and Umbels, *J. Ess. Oil Res.* **5**(7), 529-531.
- [29] D. Stesevic, M. Bozovic, V. Tadic, D. Rancic and Z. D. Stevanovic (2016). Plant-part anatomy related composition of essential oils and phenolic compounds in *Chaerophyllum coloratum*, A Balkan endemic species, *Flora*, **220**, 37-51.
- [30] B. Demirci, M. Koşar, F. Demirci, M. Dinc and K. H. C. Baser (2007). Antimicrobial and antioxidant activities of the essential oil of *Chaerophyllum libanoticum* Boiss. et Kotschy, *Food Chem.* **105**(4), 1512-1517.
- [31] S. A. Mamedova, (1994). Essential oil of *Chaerophyllum macrospermum*, *Chem. Nat. Compd.* **30(2)**, 267-268.
- [32] A. Rustaiyan, N. Neekpoor, M. Rabani, H. Komeilizadeh, S. Masoudi and A. Monfared (2002). Composition of the essential oil of *Chaerophyllum macrospermum* (Spreng.) Fisch. and CA Mey. from Iran, *J. Ess. Oil Res.* **14(3)**, 216-217.
- [33] F. Sefidkon and M. Abdoli (2005). Essential oil composition of *Chaerophyllum macrospermum* from Iran, *J. Ess. Oil Res.* **17(3)**, 249-250.
- [34] S. M. Razavi and S. Nejad-Ebrahimi (2010). Essential oil composition of *Chaerophyllum macrospermum* (Spreng.) Fisch CA Mey seeds, *J. Ess. Oil Res.* **13(2)**, 205-210.
- [35] F. Nematollahi, M. R. Akhgar, K. Larijani, A. Rustaiyan and S. Masoudi (2005). Essential oils of *Chaerophyllum macropodum* Boiss. and *Chaerophyllum crinitum* Boiss. from Iran, *J. Ess. Oil Res.* **17**(1), 71-72.
- [36] A. Ghannadi, S. E. Sajjadi, A. J. Kukhedan and S. M. Mortazavian (2011). Volatile constituents of flowering aerial parts of *Chaerophyllum macropodum* Boiss. from Iran, *J. Ess. Oil Res.* **14(4)**, 408-412.
- [37] K. H. C. Baser, G. Ozek and T. Ozek (2006). Composition of the essential oil of *Chaerophyllum macropodum* Boiss. fruits obtained by microdistillation, *J. Ess. Oil Res.* 18, 515-517.
- [38] A. Shafaghat (2009). Antibacterial activity and composition of essential oils from flower, leaf and stem of *Chaerophyllum macropodum* Boiss. from Iran, *Nat. Prod. Commun.* **4(6)**, 861-864.
- [39] A. Shafaghat (2009). Chemical composition of the essential oil of the roots of *Chaerophyllum macropodum* Boiss. from Iran, *J. Essent. Oil Bearing Plant.* 12(5), 615-619.
- [40] W. Letchamo, E. A. Korolyk and A.V. Tkachev (2005). Chemical screening of essential oil-bearing flora of Siberia V. composition of the essential oil of *Chaerophyllum prescotti* DC. tops from Altai region, *J. Essent. Oil Res.* 17, 560-562.
- [41] R.K. Joshi (2013). Root essential oil composition of *Chaerophyllum villosum* Wall. ex DC. from Uttarakhand, India, *Am. J. Essent.* **1**(1), 34-36.
- [42] R. K. Joshi (2013). Antimicrobial activity of leaf essential oil of *Chaerophyllum villosum* Wall. Ex. DC. From Kumaun Himalayan of Uttarakhand, *IAJPR*. **3(2)**, 1503-1509.
- [43] R. K. Joshi and C. S. Mathela (2013). Volatile oil composition and antioxidant activity of leaf of *Chaerophyllum villosum* Wall. ex DC. from Uttarakhand, India, *RRST* **5**(1), 25-28.
- [44] T. Ustuner, A. Kordalı, B. Usanmaz and M. Kesdek (2018). Investigation of pesticidal activities of essential oil of *Eucalyptus camaldulensis* Dehnh, *Rec. Nat. Prod.* **12(6)**, 557-568.

- [45] T. Ustuner, A. Usanmaz Bozhuyuk, A. Komaki and S. Kordali (2019). Assessment of the growth inhibiting effect of *Satureja essential* oils on different *Fusarium* species from Wheat, *Fresenius Environ. Bull.* **28**(11), 8199-8206.
- [46] M. Al. Sakran, K. Almhemed, T. Ustuner and S. Dal (2021). Effect of aqueous extract of *Sorghum halepense* (L.) Pers. on germination and growth of some weed species. *IJSRP*. **11**(1), 404-408.
- [47] A. Muhammad, D. Ghulam, B. Jehan and A. Ambrin (2020). Phytochemical screening and antimicrobial activity of medicinally importance *Achillea millefolium* L. and *Chaerophyllum villosum* Wall. ex DC, *Pak. J. Bot.* **52**(3), 971-974.
- [48] G. Tel-Cayan, E. Deveci, Z. Molo, M. E. Duru and M. Ozturk (2021). Isolation and characterization of chemical constituents from *Chaerophyllum bulbosum* roots and their enzyme inhibitory and antioxidant effects, *Z. Naturforsch C J Biosci.* 77(1-2), 1-9.
- [49] F. Bakkali, S. Averbeck, D. Averbeck and M. Idaomar (2008). Biological effects of essential oils—A review, *Food Chem. Toxicol.* **46**, 446–475.
- [50] C. Zitzelsberger and G. Buchbauer (2015). Essential oils as a cry for help: A review, *Nat. Prod. Commun.* **10**, 1127–1138.
- [51] S. Burt (2004). Essential oils: their antimicrobial properties and potential applications in foods: A review, *Int. J. Food Microbiol.* **94**, 223–253
- [52] I. A. Freires, C. Denny, B. Benso, S. M. de Alencar and P. L. Rosalen (2015). Antibacterial activity of essential oils and their isolated constituents against cariogenic bacteria: A systematic review, *Molecules* **20**, 7329–7358.
- [53] R. Russo, M. T. Corasaniti, G. Bagetta and L. A. Morrone (2015). Exploitation of cytotoxicity of some essential oils for translation in cancer therapy. *Evid. Based. Complement. Alternat. Med.* **2015**, 397821.
- [54] G. S. Jelena, M. P. Goran, S. S. Gordana, S. D. Aleksandra and K. Z. Bojan (2016). *Chaerophyllum aureum* L. Volatiles: Composition, antioxidant and antimicrobial activity, *Rec. Nat. Prod.* **10(2)**, 245-250.
- [55] G. Jelena, Stamenkovic, P. Goran and D. Aleksandra (2021). Chemical composition and antibacterial activity of essential oil obtained from *Chareophyllum bulbosum* L. dry inflorescence, *Arh. Farm.* **71**, 52-53.
- [56] F. Çelikezen, H. Türkez, M. Firat, M. Enes Arslan, S. Öner (2022). *In vitro* evaluation of selective cytotoxic activity of *Chaerophyllum macropodum* Boiss. on cultured human SH SY5Y neuroblastoma cells, *Neurotox. Res.* **40**, 1360-1368.

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